



USER MANUAL

RNAscopeTM Multiomic LS Detection Kit

For use with BOND RXTM System, from Leica Biosystems

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Citing RNAscope in Publications

When describing a procedure for publication using this product, please refer to it as the RNAscope Assay and cite: Wang F, Flanagan J, Su N, Wang L-C, Bui S, Nielson A, Wu X, Vo H-T, Ma X-J and Luo Y. RNAscope: A Novel *In Situ* RNA Analysis Platform for Formalin Fixed Paraffin Embedded Tissues. *J. Mol. Diagnostics*, 2012, 14:22–29.

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Chapter 1. Product Information



Before using this product, read and understand the information in **Appendix L. Safety** of this document.

IMPORTANT! We recommend reading the entire user manual before beginning any protocols.

About this guide

This user manual provides guidelines and protocols to use the RNAscope Multiomic LS Fluorescent Reagent Kit for use with the BOND RX Research Advanced Staining System.

For questions or support, contact your ACD representative at +1 (877) 576-3636.

Product description

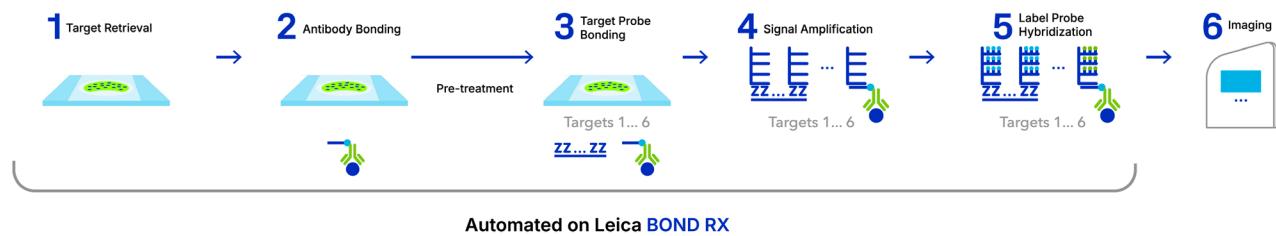
Background

The RNAscope Multiomic LS Assay uses a novel and proprietary method of *in situ* hybridization (ISH) to simultaneously visualize up to six RNA or protein targets in samples mounted on slides. The assay is based on ACD's patented signal amplification and background suppression technology and incorporates signal amplification systems that enable users to investigate expression as well as positional relationship of multiple genes within a cellular context. The RNAscope Multiomic LS Assay allows users to automate the highly sensitive Multiomic Assay using the BOND RX System.

Overview

Figure 1 on page 7 illustrates the RNAscope Multiomic LS Fluorescent Assay procedure, which can be completed on the instrument in ~15 hours. Starting with properly prepared samples, sections are incubated with target retrieval reagents and incubated with antibody to allow binding with their protein targets, then the sections are incubated with pretreatment reagents and RNA-specific probes to hybridize to their target RNAs. The RNAscope Multiomic LS Fluorescent Assay enables up to six independent signal amplification systems each using a different fluorophore to enable independent detection of the six protein or RNA targets or a combination of both. While protein signals are similar to standard fluorescent immunofluorescence signals, the assay is sensitive enough to detect single RNA transcripts which appear as punctate dots that are visible using a fluorescent microscope or slide scanner.

Figure 1. Procedure overview



1. Target retrieval	2. Antibody binding	3. Target probe hybridization	4. Signal amplification and label probe hybridization	5. Label probe hybridization	6. Imaging
Start with properly prepared tissue sections and perform target retrieval to allow access to target.	Incubation with target antibodies.	Perform pretreatments and hybridize multiple sets of gene-specific probe pairs to target RNAs.	RNA scope technology sensitively and specifically amplifies signal.	Label probe hybridization and TSA amplification results in deposition of up to six spectrally distinct fluorophores.	Visualize protein and RNA targets using a fluorescent microscope or slide scanner.

Kit contents and storage

The RNA scope Multiomic LS Assay requires the RNA scope Multiomic LS Probes (for RNA detection), Conjugated antibodies (for protein detection) and the RNA scope Multiomic LS Reagents, available from Advanced Cell Diagnostics.

RNAScope Multiomic LS Probes

The RNAScope Multiomic LS Probes include the user-specified Target Probe and the Positive and Negative Control Probes. Visit <https://acdbio.com/products> to find a gene specific target probe or appropriate control probes. Each target probe contains a mixture of short oligonucleotides designed to bind to a specific target mRNA, and detectable in one of six probe channels C1, C2, C3, C4, C5 or C6. Signal detection is performed using Tyramide Signal Amplification (TSA) linked fluorophores. Different colors are assigned to the C1, C2, C3, C4, C5 and C6 channel tags depending on the TSA Vivid™, Opal™ dye or TSA® plus fluorophore selected for that channel.

Channel C1 target probes are Ready-To-Use (RTU), while channels C2, C3, C4, C5 and C6 probes are shipped as a 50X concentrated stock. To independently detect multiple target RNAs, each target probe must be in a different channel. If you are using only the C2, C3, C4, C5 and C6 probes, you can use Probe Diluent (Cat. No. 300048)

Each probe is sufficient to stain ~30 standard slides. The probes have a shelf life of two years from the manufacturing date when stored as indicated in the following tables:

Target Probes					
<input checked="" type="checkbox"/>	Reagent	Cat. No.	Content	Quantity	Storage
	RNAScope Multiomic LS Target Probe - <i>[species]</i> - <i>[gene]</i> - C1	Various	Ready-To-Use (RTU) probe for channel C1	16 mL x 1 bottle	2–8°C
	RNAScope Multiomic LS Target Probe - <i>[species]</i> - <i>[gene]</i> - C2	Various	50X probe for channel C2	320 µL x 1 tube	2–8°C
	RNAScope Multiomic LS Target Probe - <i>[species]</i> - <i>[gene]</i> - C3	Various	50X probe for channel C3	320 µL x 1 tube	2–8°C
	RNAScope Multiomic LS Target Probe - <i>[species]</i> - <i>[gene]</i> - C4	Various	50X probe for channel C4	320 µL x 1 tube	2–8°C
	RNAScope Multiomic LS Target Probe - <i>[species]</i> - <i>[gene]</i> - C5	Various	50X probe for channel C5	320 µL x 1 tube	2–8°C
	RNAScope Multiomic LS Target Probe - <i>[species]</i> - <i>[gene]</i> - C6	Various	50X probe for channel C6	320 µL x 1 tube	2–8°C
Control Probes					
<input checked="" type="checkbox"/>	Reagent	Cat. No.	Content	Quantity	Storage
	RNAScope Multiomic LS 6-plex Positive Control Probe-Hs	323198	RNAScope Multiomic LS Positive Control Probe for the RNAScope Multiomic LS Fluorescent Assay – <i>POLR2A</i> (C1 channel), <i>PPIB</i> (C2 channel), <i>UBC</i> (C3 channel), <i>HPRT1</i> (C4 channel), <i>ATB</i> (C5 channel), <i>TUBB</i> (C6 channel).	16 mL x 1 bottle	2–8°C
	RNAScope Multiomic LS 6-plex Negative Control Probe	323208	RNAScope Multiomic LS Negative Control Probe for the	16 mL x 1 bottle	2–8°C

		RNAscope Multiomic LS Fluorescent Assay - <i>dapB</i> (<i>Bacillus subtilis</i> strain, C1-C6 channels).		
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RNAscope Antibodies

RNAscope antibodies are specially conjugated primary or secondary antibodies that are based on RNAscope technology and enable protein detection with enhanced sensitivity. Panels of pre-configured conjugated primary antibodies are composed of sets of antibodies that have been qualified to work either singly or with others of the same panel and can be combined with RNA-specific probes in the same assay for simultaneous detection of protein and RNA targets. Conjugated secondary antibodies are available for pairing with user-supplied primary antibodies to enable detection of additional proteins of interest.

RNAscope Human Tumor Infiltrating Lymphocyte (TIL) Primary Antibody Panel

<input checked="" type="checkbox"/>	Reagent	Dilution	Cat. No.	Quantity	Storage
	RNAscope Ab Hs CD4-C3	75x	322949	103 uL, (20 slides)	2-8°C
	RNAscope Ab Hs CD8-C4	75x	322951	103 uL, (20 slides)	2-8°C
	RNAscope Ab Hs PanCK-C5	75x	322952	103 uL, (20 slides)	2-8°C
	RNAscope Ab Hs FoxP3-C6	75x	322953	103 uL, (20 slides)	2-8°C

RNAscope Neural Primary Antibody Panel

<input checked="" type="checkbox"/>	Reagent	Dilution	Cat. No.	Quantity	Storage
	RNAscope™ Ab NeuN-C3	75x	AB0018-C3	105 uL, (20 slides)	2-8°C
	RNAscope™ Ab GFAP-C4	75x	AB0028-C4	105 uL, (20 slides)	2-8°C
	RNAscope™ Ab IBA-1-C5	75x	AB0038-C5	105 uL, (20 slides)	2-8°C

RNAscope Secondary Antibodies

<input checked="" type="checkbox"/>	Reagent	Dilution	Cat. No.	Quantity	Storage
	RNAscope anti-rabbit-C1	25x	322954	930 uL, (60 slides)	2-8°C
	RNAscope anti-mouse-C2	25x	322956	930 uL, (60 slides)	2-8°C

RNAscope Multiomic LS Reagents

To perform the RNAscope Multiomic LS assay, the RNAscope Multiomic LS CORE Reagents along with at least one of the Channel Reagents needs to be purchased. The kits provide enough reagents to stain ~ 20 or ~60 standard slides. The assay reagents are then used with RNA-specific probes (if RNA detection is desired), antibodies (if protein detection is desired), TSA linked fluorophores and mounting medium.

The assay reagents are Ready-To-Use (RTU) except for the TSA buffer, and are stored as indicated in the following tables:

RNAscope Multiomic LS CORE Reagents

<input checked="" type="checkbox"/>	Reagent	Quantity 60-slide kit Cat. No. 322930	Quantity 20-slide kit Cat. No. 323425	Storage
	RNAscope 2.5 LS Protease III	21 mL x 1 bottle	8 mL x 1 bottle	2-8°C
	RNAscope Multiomic LS Rinse	29 mL x 3 bottles	11 mL x 2 bottles	2-8°C
	RNAscope Multiomic LS AMP 1	21 mL x 1 bottle	8 mL x 1 bottle	2-8°C
	RNAscope Multiomic LS AMP 2	21 mL x 1 bottle	8 mL x 1 bottle	2-8°C

	RNAscope Multiomic LS AMP 3	21 mL x 1 bottle	8 mL x 1 bottle	2–8°C
	RNAscope PretreatPro™	21 mL x 1 bottle	8 mL x 1 bottle	2–8°C
	RNAscope Multiomic LS Hydrogen Peroxide	21 mL x 1 bottle	8 mL x 1 bottle	2–8°C
	RNAscope Multiomic LS DAPI	21 mL x 1 bottle	8 mL x 1 bottle	2–8°C
	RNAscope Multiomic Antibody Diluent	29 mL x 3 bottles	14 mL x 3 bottles	2–8°C

RNAscope Multiomic C1 Channel Reagents

<input checked="" type="checkbox"/>	Reagent	Quantity 60-slide kit Cat. No. 322935	Quantity 20-slide kit Cat. No. 323430	Storage
	RNAscope Multiomic TSA Buffer	29 mL x 1 bottle	8 mL x 1 bottle	2–8°C
	RNAscope Multiomic HRP Blocker	29 mL x 1 bottle	8 mL x 1 bottle	2–8°C
	RNAscope Multiomic LS HRP C1	21 mL x 1 bottle	8 mL x 1 bottle	2–8°C

RNAscope Multiomic C2 Channel Reagents

<input checked="" type="checkbox"/>	Reagent	Quantity 60-slide kit Cat. No. 322940	Quantity 20-slide kit Cat. No. 323435	Storage
	RNAscope Multiomic TSA Buffer	29 mL x 1 bottle	8 mL x 1 bottle	2–8°C
	RNAscope Multiomic HRP Blocker	29 mL x 1 bottle	8 mL x 1 bottle	2–8°C
	RNAscope Multiomic LS HRP C2	21 mL x 1 bottle	8 mL x 1 bottle	2–8°C

RNAscope Multiomic C3 Channel Reagents

<input checked="" type="checkbox"/>	Reagent	Quantity 60-slide kit Cat. No. 322945	Quantity 20-slide kit Cat. No. 323440	Storage
	RNAscope Multiomic TSA Buffer	29 mL x 1 bottle	8 mL x 1 bottle	2–8°C
	RNAscope Multiomic HRP Blocker	29 mL x 1 bottle	8 mL x 1 bottle	2–8°C
	RNAscope Multiomic LS HRP C3	21 mL x 1 bottle	8 mL x 1 bottle	2–8°C

RNAscope Multiomic C4 Channel Reagents

<input checked="" type="checkbox"/>	Reagent	Quantity 60-slide kit Cat. No. 322950	Quantity 20-slide kit Cat. No. 323445	Storage
	RNAscope Multiomic TSA Buffer	29 mL x 1 bottle	8 mL x 1 bottle	2–8°C
	RNAscope Multiomic HRP Blocker	29 mL x 1 bottle	8 mL x 1 bottle	2–8°C
	RNAscope Multiomic LS HRP C4	21 mL x 1 bottle	8 mL x 1 bottle	2–8°C

RNAscope Multiomic C5 Channel Reagents

<input checked="" type="checkbox"/>	Reagent	Quantity 60-slide kit Cat. No. 322955	Quantity 20-slide kit Cat. No. 323450	Storage
	RNAscope Multiomic TSA Buffer	29 mL x 1 bottle	8 mL x 1 bottle	2–8°C
	RNAscope Multiomic HRP Blocker	29 mL x 1 bottle	8 mL x 1 bottle	2–8°C
	RNAscope Multiomic LS HRP C5	21 mL x 1 bottle	8 mL x 1 bottle	2–8°C

RNAscope Multiomic C6 Channel Reagents

<input checked="" type="checkbox"/>	Reagent	Quantity 60-slide kit Cat. No. 322960	Quantity 20-slide kit Cat. No. 323455	Storage
	RNAscope Multiomic TSA Buffer	29 mL x 1 bottle	8 mL x 1 bottle	2–8°C

	RNAscope Multiomic HRP Blocker	29 mL x 1 bottle	8 mL x 1 bottle	2–8°C
	RNAscope Multiomic LS HRP C6	21 mL x 1 bottle	8 mL x 1 bottle	2–8°C

Required materials and equipment

Assay design

The RNAscope Multiomic LS assay provides flexibility in assay design to enable detection of up to a total of six protein or RNA targets. Proteins can be detected with user-provided primary antibodies paired with ACD's oligo-conjugated secondary antibodies. These protein targets can be augmented with available pre-qualified conjugated primary antibodies against common human targets. The conjugated secondary antibodies and conjugated primary antibodies are designed to specific RNAscope channels. Probes to RNA targets can be designed to any open channel not used by one of the conjugated antibodies.

Use this table to plan the channels needed for your assay based on your targets.

Target	RNAscope channel assignment					
	C1	C2	C3	C4	C5	C6
Protein targets	User provided primary antibody-rabbit + RNAscope anti-rabbit-C1	User provided primary antibody-mouse + RNAscope anti-mouse-C2	—	—	—	—
	—	—	RNAscope Primary C3	RNAscope Primary C4	RNAscope Primary C5	RNAscope Primary C6
RNA targets	RNAscope C1 probe	RNAscope C2 probe	RNAscope C3 probe	RNAscope C4 probe	RNAscope C5 probe	RNAscope C6 probe

Note: Antibodies are available only in the listed channels. Probes can be designed for any open channel

User-supplied primary antibodies

If you wish to use the RNAscope Multiomic LS assay with your own primary antibodies, these can be paired with the RNAscope conjugated secondary antibodies (listed in the previous tables). This provides the greatest sensitivity and specificity for your selected targets. Your primary antibodies should be hosted in either rabbit (for use with the RNAscope anti-rabbit-C1 antibody) or hosted in mouse (for use with the RNAscope anti-mouse-C2 antibody).

Recommended fluorophores

The RNAscope Multiomic LS assay requires purchase of TSA Vivid from ACD or Opal from Akoya Biosciences. For a 3-plex assay, TSA Vivid dyes are recommended. For a 4 or 6-plex assay, Opal dyes are recommended.

Dilute the fluorophores to the desired working concentration in the TSA Buffer provided in the RNAscope Kit. Choose a dilution factor for each fluorophore based on recommendations from ACD and your specific experimental conditions including target expression levels, tissue quality, or microscope setting. Materials are qualified with 1:1500 dilution for all fluorophores. We cannot guarantee assay results if you use other fluorescent dyes.

Note: To reconstitute dyes, follow the manufacturer instructions available on the tube labels.

Dilute the fluorophores in TSA buffer provided in the Channel Reagent kits.

Below are recommended combinations of fluorophores. Other combinations are also acceptable as long as the spectra are non-overlapping and are compatible with your imaging system while considering tissue autofluorescence.

3-Plex assay using TSA Vivid Fluorophores

<input checked="" type="checkbox"/>	Fluorophores	Cat. No.	Recommended dilution range
	TSA Vivid Fluorophore 520	323271	1:750–1:3000
	TSA Vivid Fluorophore 570	323272	1:750–1:3000
	TSA Vivid Fluorophore 650	323273	1:750–1:3000

Assays using Akoya Biosciences Opal Fluorophores

<input checked="" type="checkbox"/>	Fluorophores	Akoya Biosciences Cat. No.	Recommended dilution range	3-plex ²	4-plex	5- or 6-plex
	Opal 480 Reagent Pack	FP1500001KT	1:750–1:3000			✓
	Opal 520 Reagent Pack	FP1487001KT	1:750–1:3000	✓	✓	✓
	Opal 570 Reagent Pack	FP1488001KT	1:750–1:3000	✓	✓	✓
	Opal 620 Reagent Pack	FP1495001KT	1:750–1:3000			✓
	Opal 690 Reagent Pack	FP1497001KT	1:750–1:3000	✓	✓	✓
	Opal Polaris 780 Reagent Pack ¹	FP1501001KT	TSA-DIG: 1:750–1:3000		✓	✓

<input checked="" type="checkbox"/>	Fluorophores	Akoya Biosciences Cat. No.	Recommended dilution range	3-plex ²	4-plex	5- or 6-plex
			Polaris 780: 1:187.5–1:750			

¹The Opal Polaris 780 Reagent Pack contains two reagents: Opal TSA-DIG and Opal Polaris 780. We recommend diluting Polaris TSA-DIG in TSA buffer, and diluting Opal Polaris 780 in Antibody Diluent/Block from Akoya Biosciences (PN: ARD1001EA). We recommend keeping the dilution factors of Opal TSA-DIG and Opal Polaris 780 at a constant ratio. For example, when using 1:1500 dilution for Opal TSA-DIG, use 1:375 dilution for Opal Polaris 780. When using 1:750 dilution for Opal TSA-DIG, use 1:187.5 dilution for Opal Polaris 780.

²Opal 650 or Polaris 780 may be used instead of Opal 690, depending on your imager configuration.

Required slide scanner or microscope

It is recommended to use a system with multispectral capabilities, especially for imaging tissue with high autofluorescence. For optimal fluorescence detection, we recommend using a high resolution and high sensitivity cooled CCD camera that is 64 µm pixel size or smaller with > 65% peak quantum efficiency. Common models include Orca-Flash 4.0 (Hamamatsu) and Nuance® EX (Perkin Elmer).

Slide scanner or microscope	Optics
<ul style="list-style-type: none"> • Akoya PhenolImager HT • Leica DM series or equivalent • Zeiss Axio Imager, Axioscan or equivalent • Inverted microscope if optics and condenser meet requirements. • Required excitation/emission filter cube for 6-plex: DAPI/Opal480/Opal520/Opal570/Opal620/Opal690/Opal780 	<ul style="list-style-type: none"> • 20X (N.A 0.75) air • 40X (N.A. 0.8) air (recommended) • 40X (N.A. 1.3) oil • 63X (N.A. 1.3) oil – use for low expression targets, if needed • Use 20X and 40X to visualize high expression genes and low expression genes, respectively

Required materials and equipment from Leica Biosystems

The RNAscope Multiomic LS Fluorescent Assay is designed for the Leica BOND RX and requires specific materials and equipment available *only* from Leica Biosystems.

<input checked="" type="checkbox"/>	Component	Cat. No.	Storage
	BOND RX System — automated slide stainer	—	—
	BOND 30 mL Open containers	OP309700	Room temp (20–25°C)
	BOND 6 mL Titration containers*	OPT9049	Room temp (20–25°C)
	BOND Research Detection System	DS9455	Room temp (20–25°C)
	BOND Universal Covertile	S21.4611	Room temp (20–25°C)
	BOND Epitope Retrieval Solution 1-1L (RTU)	AR9961	2–8°C
	BOND Epitope Retrieval Solution 2-1L (RTU)	AR9640	2–8°C
	BOND Dewax Solution – 1L (RTU)	AR9222	2–6°C
	BOND Wash Solution 10X Concentrate – 1L	AR9590	2–8°C
	BOND Aspirating Probe Cleaning System	CS9100	2–8°C
	BOND Mixing Stations	S21.1971	Room temp (20–25°C)

* BOND 7 mL Containers can be used instead but offer less flexibility.

Other user-supplied materials

IMPORTANT! Do not substitute other materials for the SuperFrost® Plus Slides listed in the following table.

<input checked="" type="checkbox"/>	Description	Supplier	Cat. No.
	SuperFrost Plus Slides (required)	Fisher Scientific	12-550-15
	Salmon Sperm DNA, sheared (10mg/ml)	ThermoFisher	AM9680
	Normal Rabbit IgG Control	R&D Systems	MAB1050
	Mouse IgG2A Isotype Control	R&D Systems	MAB003
	ProLong™ Gold Antifade Mountant	Thermo Fisher	P36930; P10144; P36934
	Opal dyes fluorophores (if not using TSA Vivid Dyes from ACD)	Akoya Biosciences	—
	Either BOND Primary Antibody Diluent or Antibody Diluent/Block (if Opal Polaris 780 is used)	Leica Biosystems Akoya Biosciences	AR9352 ARD1001EA
	Xylene	Fisher Scientific/MLS	X3P-1GAL
	100% alcohol (EtOH)	American Master Tech Scientific/MLS*	ALREACS
	10% neutral-buffered formalin (NBF)	MLS	—
	Paraffin wax	MLS	—
	1X PBS	MLS	—
	Microtome	MLS	—
	Drying oven, capable of holding temperature at 60 +/- 1°C (optional)	MLS	—
	Water bath or incubator, capable of holding temperature at 40 +/- 1°C	MLS	—
	Tissue-Tek® Vertical 24 Slide Rack	American Master Tech Scientific/MLS	LWSRA24
	Tissue-Tek Staining Dish (4 required)	American Master Tech Scientific/MLS	LWT4457EA
	Tissue-Tek Clearing Agent Dish, xylene resistant (2 required)	American Master Tech Scientific/MLS	LWT4456EA
	Cover Glass 24 x 50 mm	Fisher Scientific/MLS	12-545-F
	Distilled water	MLS	—
	Fume hood	MLS	—

* Major Laboratory Supplier in North America. For other regions, please check Catalog Numbers with your local lab supplier.

2

Chapter 2. Before You Begin

Prior to running the RNAscope Multiomic LS Fluorescent Assay on your samples for the first time, we recommend that you:

- Become familiar with BOND RX Research Advanced Staining System from Leica Biosystems. Refer to the *BOND RX System User Manual*.
- Run the assay on Control Slides (Cat. No. 310045 for Human HeLa Cell Pellet, and Cat. No. 310023 for Mouse 3T3 Cell Pellet) using the RNAscope Multiomic LS Positive and Negative RNAscope Multiomic Control Probes.

Important procedural guidelines

- Start with properly fixed and prepared sections. Refer to **Chapter 3. Prepare Samples** for preparation of FFPE, fixed-frozen or fresh-frozen slides. For preparation of other sample types, contact support.acd@bio-techne.com.
- Regularly maintain and clean your automated staining instrument.
- Always run positive and negative control probes on your sample to assess sample RNA quality and optimal permeabilization.
- Do not substitute required materials. The assay has been validated with these materials only.
- Follow the protocol exactly for the best results.
- Do not let your sections dry out during the procedure.
- Use good laboratory practices and follow all necessary safety procedures. Refer to **Appendix L. Safety** for more information.

Chapter 3. Prepare Samples

The following protocols describe formalin-fixed, paraffin-embedded (FFPE), fixed frozen and fresh frozen sample preparation.

IMPORTANT! We highly recommend following these guidelines. We cannot guarantee assay results with other preparation methods.

IMPORTANT! RNA-only staining has been validated by ACD for all sample preparations described in this manual, but only FFPE samples have been fully tested for combined RNA + protein staining. Please see the following guidance for details.

Prepare FFPE sections

Materials required

- 10% neutral buffered formalin (NBF)
- 1X PBS
- Paraffin wax
- 95% Ethanol (EtOH)
- Xylene
- Microtome
- Water bath
- SuperFrost Plus slides

Fix the sample

1. Immediately following dissection cut the tissue into blocks of 3–4 mm in thickness.
2. Place the tissue blocks into fixative within **1 HR** of biopsy.
3. Fix the tissue in 10% NBF for **16–32 HRS** at **ROOM TEMPERATURE (RT)**. Fixation time will vary depending on tissue type and size.



CAUTION! Handle biological specimens appropriately.

IMPORTANT! Fixation for **<16 HRS** or **>32 HRS** will impair the performance of the assay.

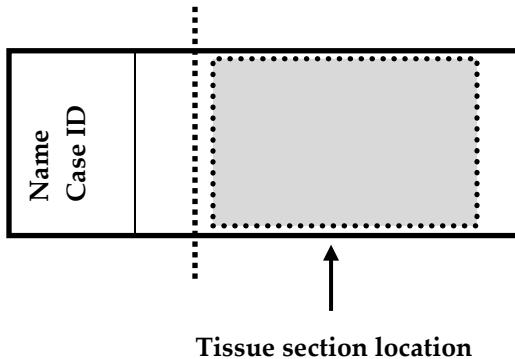
Dehydrate, embed, and cut the sample

IMPORTANT! Use fresh reagents.

1. Wash sample with 1X PBS.
2. Dehydrate sample using a standard ethanol series, followed by xylene.
3. Embed sample in paraffin using standard procedures.

Note: Embedded samples may be stored at room temperature with desiccation. To better preserve RNA quality over a long period (>1 yr), storing at 2–8°C with desiccation is recommended.

4. Trim paraffin blocks as needed and cut embedded tissue into **5 +– 1 µm** sections using a microtome.
5. Place the paraffin ribbon in a **40–45°C** water bath and mount the sections on **SUPERFROST® PLUS SLIDES**. Place tissue as shown for optimal staining:



IMPORTANT! Do not mount more than one section per slide. Place sections in the center of the slide.

6. Air dry slides **OVERNIGHT** at **RT**.

OPTIONAL STOPPING POINT. Use sectioned tissue within three months. Store sections with desiccants at room temperature.

Prepare fixed-frozen sections

Materials required

- 1X PBS
- 10% Neutral Buffered Formalin (NBF) or 4% Paraformaldehyde (PFA)
- 100% alcohol (EtOH)
- Tissue-Tek Vertical 24 Slide Rack
- Tissue-Tek Staining Dishes
- Drying oven

Fix sample

1. If needed, perfuse tissue with freshly prepared 4% Paraformaldehyde (PFA) in 1X PBS, or go directly to step 2.
2. Dissect tissue and place in freshly prepared 4% Paraformaldehyde (PFA) for **24 HRS** at **4°C**.

Freeze tissue

1. Immerse the tissue in 10% sucrose in 1X PBS at **4°C** until the tissue sinks to the bottom of the containers (approximately **18 HRS** for brain tissue.)
Note: The time needed for the tissue to sink varies with the tissue type and size.
2. Immerse the tissue in 20% sucrose in 1X PBS at **4°C** until the tissue sinks to the bottom of the container.
3. Immerse the tissue in 30% sucrose in 1X PBS at **4°C** until the tissue sinks to the bottom of the container.

4. Freeze the tissue in OCT (Optimal Cutting Temperature) embedding media for TFM (Tissue Freezing Media) with crushed dry ice or iso-pentane or liquid nitrogen.
5. Store tissue blocks in an airtight container at -80°C.

Prepare sections

1. Before sectioning, equilibrate the tissue blocks at -20°C **for at least 1 HR in a cryostat**.
2. Section blocks by cutting sections to a thickness of 7 – 15 µm. Mount sections on SuperFrost® Plus slides ONLY (other slide types could result in tissue loss).
3. Air dry the slides for **2 HR** at -20°C and overnight at -80°C. If all slides are not used immediately, store them **at -80°C** for up to **3 MONTHS**.
4. On the day of starting the assay, remove fixed-frozen tissue slides from -80°C.
5. Wash the slides with 200 mL 1X PBS for **5 MIN** while moving the rack to remove OCT.
6. Bake slides in drying oven for **15 - 60 MIN** at **60°C**.
7. *Immediately* post-fix slides by immersing them in prechilled 10% NBF or 4% PFA for **15–60 MIN** at **4°C**.

Note: If experiencing issues with sample detachment, the longer post-fix and baking times could be helpful.

Dehydrate and dry the sections

Reagents can be prepared ahead of time. Ensure all containers remain covered.

1. Prepare 200 mL 50% ethanol, 200 mL 70% ethanol, and 2X 200 mL 100% ethanol in Tissue Tek Staining Dishes.
2. Remove the slides from the 10% NBF or 4% PFA, and immerse them in 50% EtOH for **5 MIN** at **RT**.
3. Place the slides in 70% ethanol for **5 MIN** at **RT**.
4. Place the slides in 100% ethanol for **5 MIN** at **RT**.
5. Place slides in fresh 100% ethanol for **5 MIN** at **RT**.
6. Remove slides from ethanol, and let them dry for **5 MIN** at **RT**.

Prepare fresh-frozen sections

Materials required

-
- 1X PBS
 - 10% Neutral Buffered Formalin (NBF) or 4% Paraformaldehyde (PFA)
 - 100% alcohol (EtOH)
 - Tissue-Tek Vertical 24 Slide Rack
 - Tissue-Tek Staining Dishes
-

Section preparation

1. Cryosection the tissue to 10 – 20 um thickness and place onto SuperFrost Plus slides.
2. Keep the section at -20°C to dry for **1 HR**.
3. Store the sections at -80°C. Sections can be stored for at least **3 MONTHS** at -80°C.

Note: Do not process the slides with any fixative (alcohol or formaldehyde before) this step.

4. The slides can be shipped in dry ice.

Fix the sections

1. Pre-chill 200 ml of 10% NBF or 4% PFA in 1x PBS to **4°C**.
2. Remove fresh-frozen tissue slides from -80°C and place in a Tissue Tek Slide Rack.
3. *Immediately* immerse the slides in 200 mL of 10% NBF or freshly prepared 4% PFA.
4. Incubate the slides for at least **90 MIN** at **ROOM TEMPERATURE (RT)**.

Note: Formalin that has been stored for more than six months, exposed to air for more than a week, or used repeatedly may result in suboptimal tissue fixation. 4% PFA must be freshly prepared for each experiment.

Dehydrate the sections

Reagents can be prepared ahead of time. Ensure all containers remain covered.

1. Prepare 200 mL 50% ethanol, 200 mL 70% ethanol, and 2X 200 mL 100% ethanol in Tissue Tek Staining Dishes.
2. Place the slides in 50% ethanol for **5 MIN** at **RT**.
3. Place the slides in 70% ethanol for **5 MIN** at **RT**.
4. Place the slides in 100% ethanol for **5 MIN** at **RT**.
5. Place slides in fresh 100% ethanol for **5 MIN** at **RT**.
Note: If needed, slides can be stored in 100% EtOH at -20°C for up to **1 WEEK**. Prolonged storage may degrade sample RNA.
6. Remove slides from ethanol, and let them dry for **5 MIN** at **RT**.

Chapter 4. Determine Pretreatment Conditions

The following protocols describe formalin-fixed, paraffin-embedded (FFPE), fixed-frozen and fresh-frozen sample pretreatment. For other sample types and preparation methods, contact support.acd@biotechne.com for the latest protocols and guidelines.

IMPORTANT! We highly recommend following these guidelines. We cannot guarantee assay results with other preparation methods.

IMPORTANT! RNA-only staining has been validated by ACD for all sample preparations described in this manual, but only FFPE samples have been fully tested for combined RNA and protein staining. Please see the following guidance for details.

Sample preparation for RNA-only staining

Pretreat FFPE sections

Target retrieval

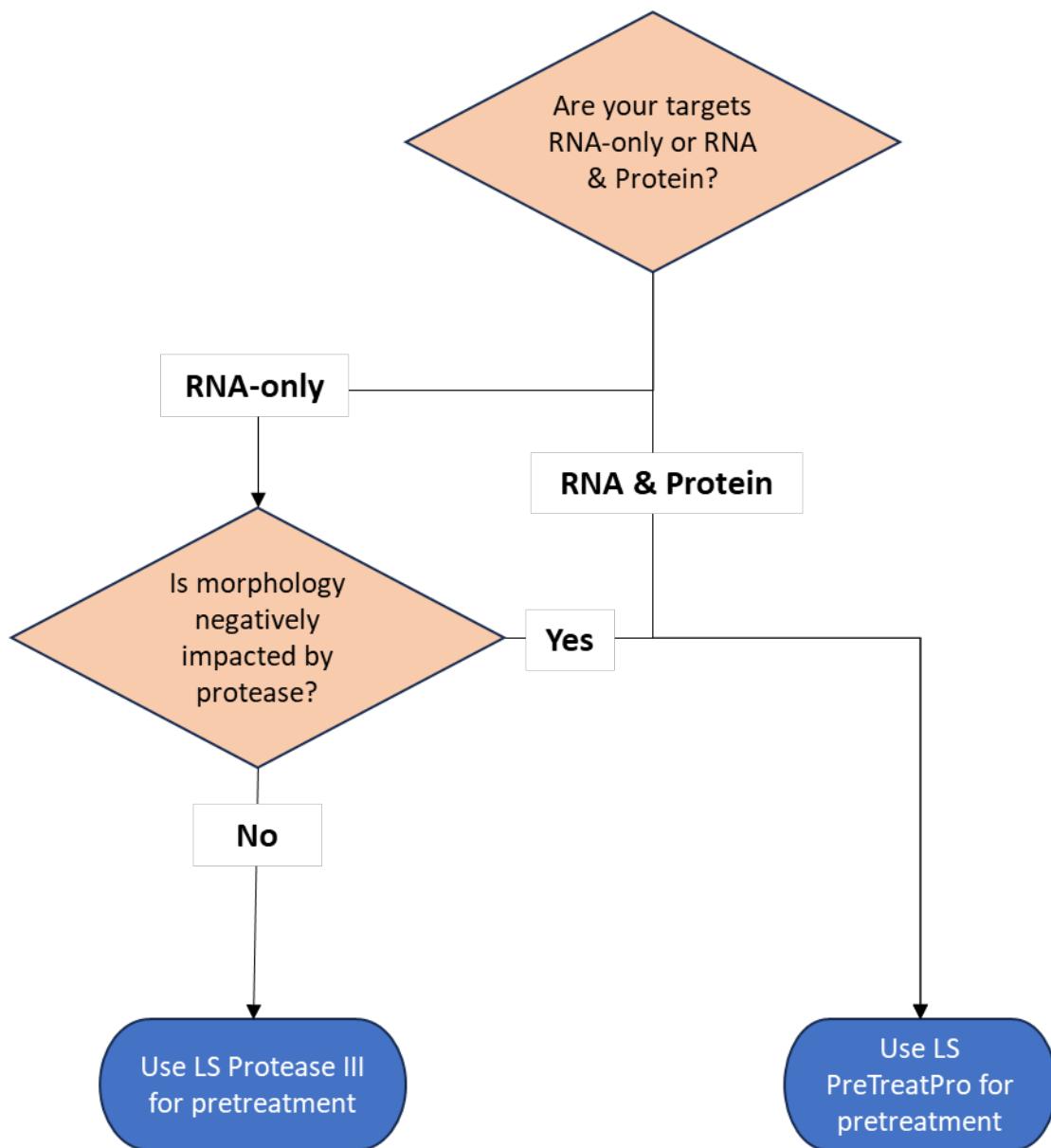
FFPE samples must be de-crosslinked with a target retrieval step. The Multiomic LS Assay uses the BOND RX's ER2 solution exclusively for this step.

Permeabilization

Two options are available:

- Protease-based permeabilization is recommended for experiments that stain only RNA. This option uses LS Protease III.
- Protease-free permeabilization uses the LS PretreatPro reagent which is free of protease enzyme. This allows co-detection of RNA and proteins that were previously incompatible with protease on the same tissue section using immunohistochemistry (IHC). Please refer to [Appendix J](#) to implement the use of LS PretreatPro reagent on the BOND RX.

To determine the correct permeabilization option, please refer to the following flowchart:



Tissue pretreatment recommendations

Use these conditions as a starting point when tissues are prepared as described in **Chapter 3**. Depending on your tissue type, vary the amount of time for the ER2 and LS Protease III until positive RNA control signal is maximized with minimal/no negative RNA control signal (see **Appendix I**. Pretreatment Guidance for FFPE Samples for a list of tissues)

Reagent	Mild	Standard
BOND ER2*	15 MIN @88C	15 MIN @95C
LS Protease III OR LS PretreatPro (see Appendix D)	15 MIN @40C	15 MIN @40C
		30 MIN @40C

*Sample types, such as certain xenografts and cell pellets, might require shorter incubation time. For these tissue types, reduce the BOND ER2 incubation time. LS Protease III incubation times can be adjusted but are rarely needed. If you have a tissue type not listed, contact ACD Support at support.acd@bio-techne.com.

Pretreat fixed-frozen sections

Target Retrieval

Fixed-frozen samples must be gently de-crosslinked with a target retrieval step. The RNAscope 2.5 LS Assay uses the BOND RX's ER2 solution exclusively for this step.

Permeabilization

Only LS Protease III has been tested for use with fixed-frozen sections. Check with ACD Support for any updates.

Tissue pretreatment recommendations

Use these conditions as a starting point when tissues are prepared as described in **Chapter 3**. Depending on your tissue type, vary the amount of time for the ER2 and/or LS Protease III until positive control RNA signal is maximized with minimal or no negative RNA control signal (see **Appendix L** for details).

Reagent	Standard
BOND ER2	5 MIN @95C*
LS Protease III†	15 MIN @40C

* You might need to create this heat treatment protocol. Please refer to **Appendix F** for further instructions.

† Sample types, such as certain xenografts and cell pellets, might require shorter incubation time. For these tissue types, reduce the BOND ER2 incubation time. LS Protease III time can also be adjusted but is rarely needed.

Pretreat fresh-frozen sections

Target Retrieval

Fresh-frozen sections do not need target retrieval.

Permeabilization

Only LS Protease IV has been tested for use with fresh-frozen sections. Check with ACD Support for any updates.

Tissue pretreatment recommendations

Use this condition as a starting point when tissues are prepared as described in **Chapter 3**. Depending on your tissue type, vary the amount of time for the **Protease IV** until positive RNA control signal is maximized with minimal/no negative RNA control signal (see **Appendix L** for details).

Reagent	Standard
LS Protease IV (ACD Part Number 322140)	30 MIN @Ambient*

* You might need to create this heat treatment protocol. Please refer to **Appendix F** for further instructions.

Sample preparation for RNA + protein staining

When performing dual staining for RNA and protein targets, detailed guidance is available for FFPE sample types. For other sample types, you can extrapolate from the RNA-only staining guidance provided in the previous section. If you need further assistance, please contact ACD Support.

Pretreat FFPE sections

Target retrieval

FFPE samples must be de-crosslinked with a target retrieval step. The Multiomic LS Assay specifically uses the BOND RX's ER2 solution for this step.

Permeabilization

Only LS PretreatPro has been tested on FFPE samples stained for both RNA and protein. Using LS Protease could negatively impact staining performance of protease sensitive antigens.

Tissue pretreatment recommendations

Use these conditions as a starting point when FFPE tissues are prepared as described in **Chapter 3**. Depending on your tissue type, vary the amount of time for the ER2 until positive RNA control signal is maximized with minimal/no negative RNA control signal (see **Appendix I**. Pretreatment Guidance for FFPE Samples for a list of tissues)

Reagent	Mild	Standard**
BOND ER2	15 MIN @88C	15 MIN @95C
LS PretreatPro*		30 MIN @40C

* Sample types, such as certain xenografts and cell pellets, might require shorter incubation time. For these tissue types, reduce the BOND ER2 incubation time. If you have a tissue type not listed, contact ACD Support at support.acd@bio-techne.com.

** Target retrieval for 20min at 100°C can be used if dye trapping or lower antibody signal is observed.

5a

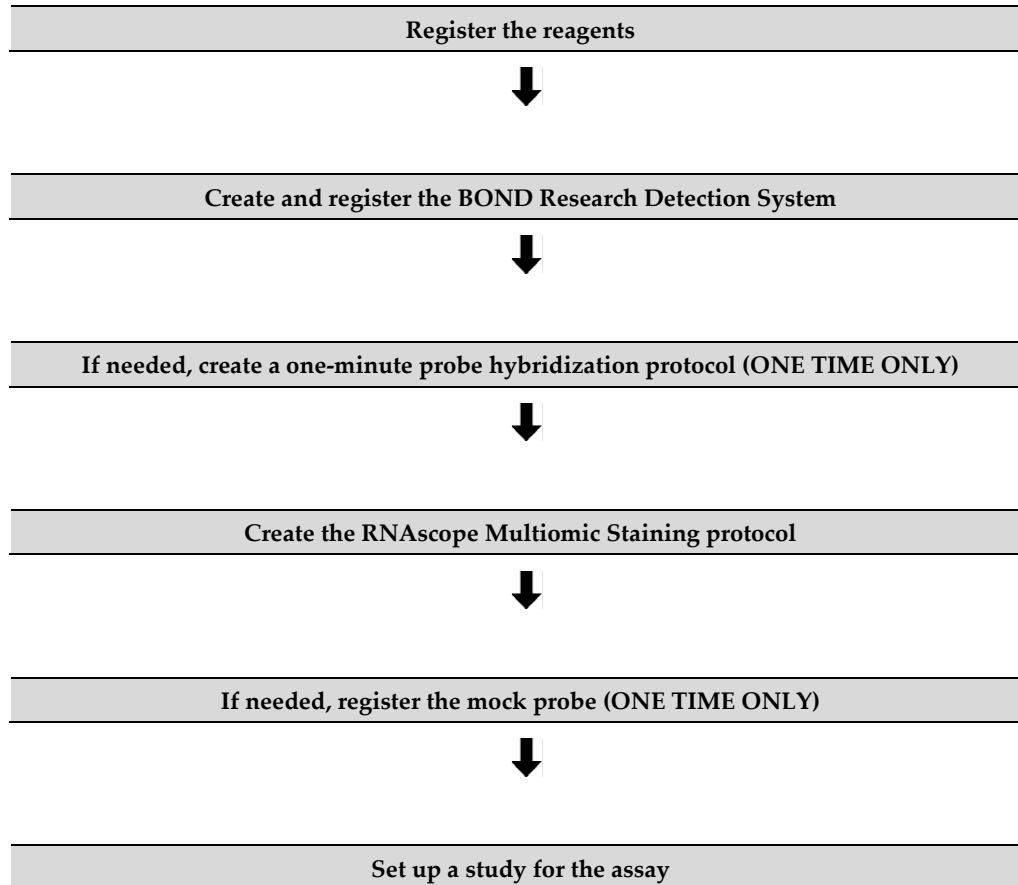
Chapter 5. Set Up a Staining Protocol Using the Mock Probe (software version 6.0 and 7.0)

Use the instructions in this chapter to set up the RNAscope Multiomic LS Assay using the mock probe workaround. This software version is required for runs that combine RNA and protein staining. To set up the assay without the mock probe for RNA-only staining, proceed to [Chapter 6](#).

IMPORTANT! We strongly recommend you run the Control Slides (Cat. No. 310045 for human or Cat. No. 310023 for mouse) using the RNAscope Multiomic LS positive and negative control probes along with your samples in every run.

IMPORTANT! Make sure you have software version 6.0 or higher before setting up the assay. The RNAscope Multiomic LS Assay is not supported by earlier software versions.

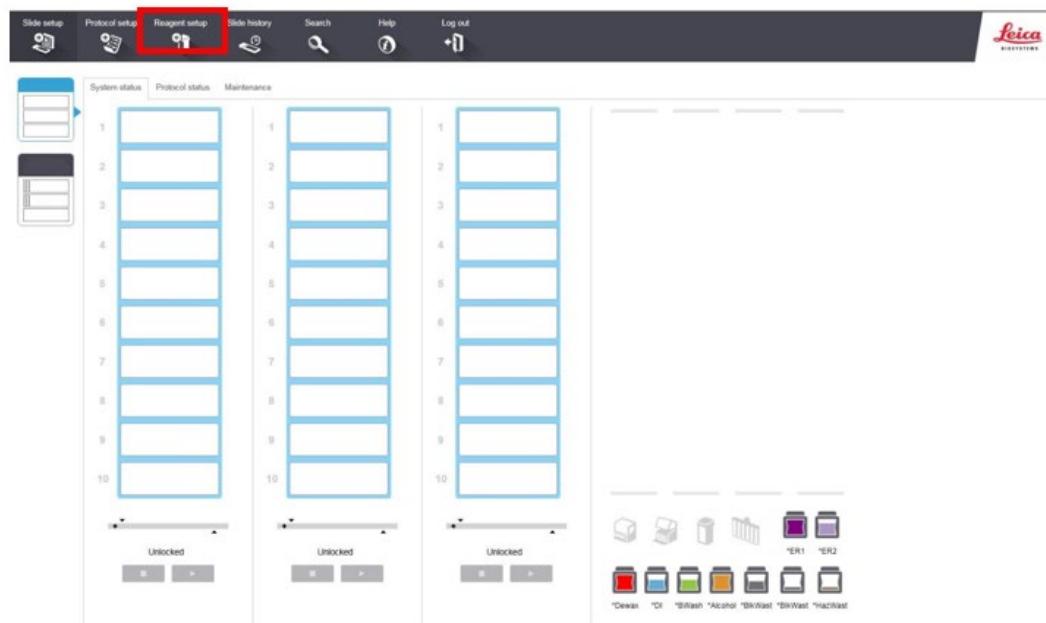
Workflow



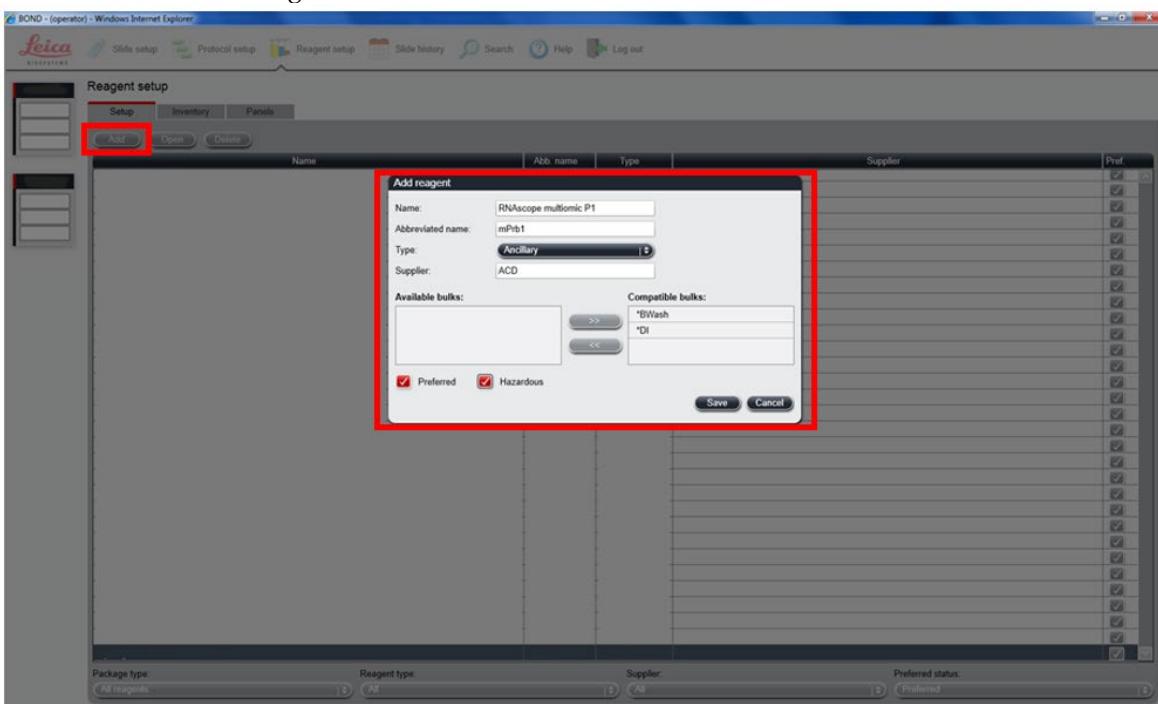
Register the reagents

This step is a “workaround” to the existing 6.0/7.0 software to accommodate the RNAscope Multiomic LS Assay. Your ACD Field Application Specialist (FAS) can help implement this procedure. In summary, a probe is created as an ancillary reagent and added to the staining protocol.

1. Select the **Reagent Setup** icon at the top of the screen.



2. Select **Add** to enter reagent information.



3. Enter a reagent name (for example, RNAscope Multiomic probe 1) in the Name text box.
4. Enter **RNAscope Multiomic P1** (for example) in the Abbreviated name text box.
5. Select **Ancillary** in the Type drop-down menu.
6. Enter **ACD** in the Supplier text box.
7. Check both the **Preferred** and **Hazardous** boxes (for the probes, RNAscope Multiomic Amp 1, and RNAscope Multiomic Amp 3 only).

8. Select **Save**.
9. Repeat Steps 2–7 for each probe and for the rest of the reagents using the container names in the following table. Reagents with an * are pre-installed in the software and do not need to be added:

Reagents	Container Name	Abbreviated Name	Volume per slide (dead volume not included)	Container Type for 60 slide kit	Container Type for 20 slide kit
RNAscope Multiomic LS AMP 1	RNAscope Multiomic LS Amp 1	MO-Amp1	300 µL	30 mL Open	7 mL Titration
RNAscope Multiomic LS AMP 2	RNAscope Multiomic LS Amp 2	MO-Amp2	300 µL	30 mL Open	7 mL Titration
RNAscope Multiomic LS AMP 3	RNAscope Multiomic LS Amp 3	MO-Amp3	300 µL	30 mL Open	7 mL Titration
RNAscope Multiomic LS HRP C1	RNAscope Multiomic LS HRP-C1	MO-HRPC1	300 µL	30 mL Open	7 mL Titration
RNAscope Multiomic LS HRP C2	RNAscope Multiomic LS HRP-C2	MO-HRPC2	300 µL	30 mL Open	7 mL Titration
RNAscope Multiomic LS HRP C3	RNAscope Multiomic LS HRP-C3	MO-HRPC3	300 µL	30 mL Open	7 mL Titration
RNAscope Multiomic LS HRP C4	RNAscope Multiomic LS HRP-C4	MO-HRPC4	300 µL	30 mL Open	7 mL Titration
RNAscope Multiomic LS HRP C5	RNAscope Multiomic LS HRP-C5	MO-HRPC5	300 µL	30 mL Open	7 mL Titration
RNAscope Multiomic LS HRP C6	RNAscope Multiomic LS HRP-C6	MO-HRPC6	300 µL	30 mL Open	7 mL Titration
RNAscope Multiomic LS HRP Blocker	RNAscope Multiomic LS HRP Blocker	HRPBK	900 µL	30 mL Open	30 mL Open
Opal/TSA Vivid-fluorophore 1 (user to dilute in TSA buffer)	Multiomic TSA-F1	MO-TSAF1	300 µL	6 mL Titration	6 mL Titration
Opal/TSA Vivid-fluorophore 2 (user to dilute in TSA buffer)	Multiomic TSA-F2	MO-TSAF2	300 µL	6 mL Titration	6 mL Titration
Opal/TSA Vivid-	Multiomic TSA-F3	MO-TSAF3	300 µL	6 mL Titration	6 mL Titration

Reagents	Container Name	Abbreviated Name	Volume per slide (dead volume not included)	Container Type for 60 slide kit	Container Type for 20 slide kit
fluorophore 3 (user to dilute in TSA buffer)					
Opal-fluorophore 4 (user to dilute in TSA buffer)	Multiomic TSA-F4	MO-TSAF4	300 µL	6 mL Titration	6 mL Titration
Opal-fluorophore 5 (user to dilute in TSA buffer)	Multiomic TSA-F5	MO-TSAF5	300 µL	6 mL Titration	6 mL Titration
Opal TSA-DIG † (if Opal 780 is used)	TSA-DIG	TSA-DIG	300 µL	6 mL Titration	6 mL Titration
Opal 780† (if Opal 780 is used)	Opal 780	Opal780	300 µL	6 mL Titration	6 mL Titration
LS RNAscope Multiomic probe mix	(user's choice but must be in container registered as Ancillary in software)		300 µL	6 mL Titration	6 mL Titration
RNAscope Multiomic LS DAPI	*DAPI (Misc)	DAPI_1	150 µL	30 mL Open	30 mL Open
1X BOND Wash (for Research Detection Kit)	*Detection Wash	Detect Wash	150 µL	30 mL Open	30 mL Open
1X BOND Wash	Mock Probe (registered as Probe RNA in software)	ACDMock	270 µL		
RNAscope Multiomic LS Hydrogen Peroxide	*Open 0 Haz		150 µL	30 mL Open	7 mL Titration
RNAscope 2.5 LS Protease III	*ACD Enzyme		300 µL	30 mL Open	7 mL Titration
RNAscope LS PretreatPro	PretreatPro	PretPro	300 µL	30 mL Open	7 mL Titration
RNAscope Multiomic LS Rinse	RNAscope Multiomic LS Rinse		600 µL	30 mL Open	30 mL Open

*Indicates this reagent is hard coded in the software by Leica Biosystems.

†These reagents are only needed when Opal Polaris 780 is used in the assay.

Create and register the BOND Research Detection System (one time only)

A BOND Research Detection System from Leica is required to setup the RNAscope Multiomic LS Fluorescent Assay. Your ACD Field Application Specialist (FAS) can help implement this procedure. Each detection system barcode is valid for up to 200 tests of use.

1. Scan the barcode on the tray of a new BOND Research Detection System.
2. To setup a new detection system for the assay, enter **ACD LS Multiplex Detection Kit** in the Name text box.

Note: Creating the detection system needs to be performed only once on each BOND RX controller.

Pstn.	UPI	Reagent	Vol. (mL)
1		DAPI	
2		Bond Wash	

3. Place two new BOND 30 mL Open containers on the Research Detection System tray.
4. Scan the first container and select the registration name **DAPI**. You can mix different lots of DAPI in the same container.

Note: If you prefer not to use DAPI on the instrument or want to perform immunohistochemistry (IHC) steps after the assay, you can use Bond wash in place of DAPI in the staining protocol.

5. Scan the second container and select the registration name **Bond wash**.
6. When one Research Detection System is finished (after 200 tests), register a new detection system by scanning the barcode on the tray and select **ACD LS Multiplex Detection Kit** from the drop-down menu on the right. Enter the new lot number.
7. Register two new 30 mL containers for *DAPI and **Bond wash** by first selecting the reagent name and then scanning the barcode on the container.
8. Select **OK**.

Create a staining protocol

Due to the software workaround for the RNAscope Multiomic LS Fluorescent Assay, unique staining protocols *must be created for each probe*. Your ACD Field Application Specialist (FAS) can help implement this procedure.

1. In the Protocol setup screen, select **Staining** under the Protocol group menu.
2. Highlight the *ACD 2.5 DAB Rev B protocol. Select **Copy**.

Protocol name	Protocol type	Description	Modified by	Mod. date	Pref.
*ACD 2.5 DAB Rev B	ISH detection	ACD 2.5 DAB Rev B protocol	Leica	5/19/2023	<input checked="" type="checkbox"/>
*ACD 2.5 Red Rev B	ISH detection	ACD 2.5 Red Rev B protocol	Leica	5/19/2023	<input checked="" type="checkbox"/>
*ACD DAB ISH Protocol	ISH detection	ACD RNAscope DAB ISH protocol	Leica	5/19/2023	<input checked="" type="checkbox"/>
*ACD LS Duplex RED/DAB	ISH detection	RNAscope Duplex LS RED & DAB Protocol	Leica	5/19/2023	<input checked="" type="checkbox"/>
*ACD LS Multiplex C1-3	ISH detection	RNAscope LS Multiplex Fluorescence Channels 1 to 3	Leica	5/19/2023	<input checked="" type="checkbox"/>
*ACD Red ISH Protocol	ISH detection	ACD RNAscope Red ISH protocol	Leica	5/19/2023	<input checked="" type="checkbox"/>
*BaseScope LS RED ISH	ISH detection	BaseScope LS RED ISH protocol	Leica	5/19/2023	<input checked="" type="checkbox"/>
*BaseScope LS RED ISH - FAN	ISH detection	BaseScope LS RED ISH protocol with cooling fan	Leica	5/19/2023	<input checked="" type="checkbox"/>
*BaseScope LS RED ISH Short - FAN	ISH detection	BaseScope LS RED ISH short protocol with cooling fan	Leica	5/19/2023	<input checked="" type="checkbox"/>
*BaseScope LSx RED ISH	ISH detection	BaseScope LSx RED ISH protocol	Leica	5/19/2023	<input checked="" type="checkbox"/>
*FISH A RUO Protocol	ISH detection	Bond RUO FISH A protocol	Leica	5/19/2023	<input checked="" type="checkbox"/>
*FISH Protocol C	ISH detection	FISH wash protocol	Leica	5/19/2023	<input checked="" type="checkbox"/>
*Flexible ISH Template	ISH detection	ISH template with open dispenses plus heated wash steps	Leica	5/19/2023	<input checked="" type="checkbox"/>
*GeoMx RNA Protocol	ISH detection	GeoMx RNA Protocol	Leica	5/24/2023	<input type="checkbox"/>
*HCR ISH v5	ISH detection	HCR Staining Protocol with Probe Removal wash steps	Leica	5/19/2023	<input type="checkbox"/>
*HCR RNA-CISH AP	ISH detection	HCR Chromogenic RNA-CISH Protocol AP	Leica	5/24/2023	<input type="checkbox"/>
*HCR RNA-CISH HRP	ISH detection	HCR Chromogenic RNA-CISH Protocol HRP	Leica	5/24/2023	<input type="checkbox"/>
*ISH Heated Wash Template	ISH detection	ISH template with multiple wash temperatures	Leica	5/19/2023	<input checked="" type="checkbox"/>
*ISH Open Dispense Template	ISH detection	ISH template with Open Ancillary and Chromogen dispenses	Leica	5/19/2023	<input checked="" type="checkbox"/>
*ISH Parallel Protocol	ISH detection	ISH Parallel Template Protocol	Leica	5/19/2023	<input type="checkbox"/>
*ISH Protocol A	ISH detection	Bond Polymer Refine RNA ISH protocol	Leica	5/19/2023	<input checked="" type="checkbox"/>
*ISH Protocol B	ISH detection	Bond Polymer Refine DNA ISH protocol	Leica	5/19/2023	<input checked="" type="checkbox"/>

3. For the following steps, refer to the next figure:
 - a. Change the protocol name for your first probe to **ACD RNAscope Multiomic Protocol P1** in the Name text box, **Multi_P1** in the Abbreviated name text box, and **ACD RNAscope Multiomic Protocol P1** in the Description text box.
 - b. Select **ACD LS Multiplex Detection Kit** from the Preferred detection system menu.
 - c. For Steps 1–2, change to the desired probe container. In the example, the probe mix is in the **Open 1** container.

Note: The next seven figures display all reagent steps using TSA-DIG and Polaris 780 to label –C6

New protocol properties

Name:	ACD RNAscope multiomic Protocol P1
Abbreviated name:	Multi-P1
Description:	ACD RNAscope multiomic Protocol P1

Staining method: Single Preliminary Final Preferred

BOND RX Import protocol

Preferred detection system: ACD Ls Mux Detection Kit

Step N°	Wash	Reagent	Supplier	Ambient	Temperature	Inc. (min)	Dispense type
1		PretreatPro		✓		0:00	150 µL
2		PretreatPro			40	30:00	150 µL
8		*RNAscope 2.5 LSx H ₂ O ₂	Advanced Cell Diagnostics	✓		10:00	150 µL
14		*Open 1	User	✓		10:00	150 µL
15		*Open 1	User		42	120:00	150 µL
27		ACD RNAscope Multiomic Amp 1			42	1:00	150 µL
28		ACD RNAscope Multiomic Amp 1			42	30:00	150 µL
37		*LS Rinse	Advanced Cell Diagnostics	✓		5:00	150 µL
38		*LS Rinse	Advanced Cell Diagnostics	✓		5:00	150 µL
43		ACD RNAscope Multiomic Amp 2			42	1:00	150 µL
44		ACD RNAscope Multiomic Amp 2			42	30:00	150 µL
53		*LS Rinse	Advanced Cell Diagnostics	✓		5:00	150 µL
54		*LS Rinse	Advanced Cell Diagnostics	✓		5:00	150 µL
59		ACD RNAscope Multiomic Amp 3			42	1:00	150 µL
60		ACD RNAscope Multiomic Amp 3			42	15:00	150 µL
61		ACD RNAscope Multiomic Amp 4			42	4:00	150 µL

Show wash steps Insert wash | Insert reagent | Delete step

Save Cancel

New protocol properties

Name:	ACD RNAscope multiomic Protocol P1
Abbreviated name:	Multi-P1
Description:	ACD RNAscope multiomic Protocol P1

Staining method: Single Preliminary Final Preferred

BOND RX Import protocol

Preferred detection system: ACD Ls Mux Detection Kit

Step N°	Wash	Reagent	Supplier	Ambient	Temperature	Inc. (min)	Dispense type
37		*LS Rinse	Advanced Cell Diagnostics	✓		5:00	150 µL
38		*LS Rinse	Advanced Cell Diagnostics	✓		5:00	150 µL
43		ACD RNAscope Multiomic Amp 2			42	1:00	150 µL
44		ACD RNAscope Multiomic Amp 2			42	30:00	150 µL
53		*LS Rinse	Advanced Cell Diagnostics	✓		5:00	150 µL
54		*LS Rinse	Advanced Cell Diagnostics	✓		5:00	150 µL
59		ACD RNAscope Multiomic Amp 3			42	1:00	150 µL
60		ACD RNAscope Multiomic Amp 3			42	15:00	150 µL
61		ACD RNAscope Multiomic Amp 4			42	4:00	150 µL

Show wash steps Insert wash | Insert reagent | Delete step

Save Cancel

New protocol properties

Name:	ACD RNAscope multiomic Protocol P1																																																																																
Abbreviated name:	Multi-P1																																																																																
Description:	ACD RNAscope multiomic Protocol P1																																																																																
Staining method:	<input checked="" type="checkbox"/> Single <input checked="" type="checkbox"/> Preliminary <input checked="" type="checkbox"/> Final <input checked="" type="checkbox"/> Preferred																																																																																
BOND RX																																																																																	
Import protocol Protocol type: ISH detection																																																																																	
Preferred detection system: <input type="button" value="ACD Ls Mux Detection Kit"/>																																																																																	
<table border="1"> <thead> <tr> <th>Step N°</th> <th>Wash</th> <th>Reagent</th> <th>Supplier</th> <th>Ambient</th> <th>Temperature</th> <th>Inc. (min)</th> <th>Dispense type</th> </tr> </thead> <tbody> <tr> <td>60</td> <td></td> <td>ACD RNAscope Multiomic Amp 3</td> <td></td> <td></td> <td>42</td> <td>15:00</td> <td>150 µL</td> </tr> <tr> <td>69</td> <td></td> <td>ACD RNAscope Multiomic HRP-C1</td> <td></td> <td></td> <td>42</td> <td>1:00</td> <td>150 µL</td> </tr> <tr> <td>70</td> <td></td> <td>ACD RNAscope Multiomic HRP-C1</td> <td></td> <td></td> <td>42</td> <td>15:00</td> <td>150 µL</td> </tr> <tr> <td>79</td> <td></td> <td>*ACD Multiplex TSA-F1</td> <td>Advanced Cell Diagnostics</td> <td><input checked="" type="checkbox"/></td> <td></td> <td>1:00</td> <td>150 µL</td> </tr> <tr> <td>80</td> <td></td> <td>*ACD Multiplex TSA-F1</td> <td>Advanced Cell Diagnostics</td> <td><input checked="" type="checkbox"/></td> <td></td> <td>30:00</td> <td>150 µL</td> </tr> <tr> <td>88</td> <td></td> <td>ACD RNAscope Multiomic HRP blocker</td> <td></td> <td></td> <td>42</td> <td>1:00</td> <td>150 µL</td> </tr> <tr> <td>89</td> <td></td> <td>ACD RNAscope Multiomic HRP blocker</td> <td></td> <td></td> <td>42</td> <td>15:00</td> <td>150 µL</td> </tr> <tr> <td>97</td> <td></td> <td>ACD RNAscope Multiomic HRP-C2</td> <td></td> <td></td> <td>42</td> <td>1:00</td> <td>150 µL</td> </tr> <tr> <td>98</td> <td></td> <td>ACD RNAscope Multiomic HRP-C2</td> <td></td> <td></td> <td>42</td> <td>15:00</td> <td>150 µL</td> </tr> </tbody> </table>		Step N°	Wash	Reagent	Supplier	Ambient	Temperature	Inc. (min)	Dispense type	60		ACD RNAscope Multiomic Amp 3			42	15:00	150 µL	69		ACD RNAscope Multiomic HRP-C1			42	1:00	150 µL	70		ACD RNAscope Multiomic HRP-C1			42	15:00	150 µL	79		*ACD Multiplex TSA-F1	Advanced Cell Diagnostics	<input checked="" type="checkbox"/>		1:00	150 µL	80		*ACD Multiplex TSA-F1	Advanced Cell Diagnostics	<input checked="" type="checkbox"/>		30:00	150 µL	88		ACD RNAscope Multiomic HRP blocker			42	1:00	150 µL	89		ACD RNAscope Multiomic HRP blocker			42	15:00	150 µL	97		ACD RNAscope Multiomic HRP-C2			42	1:00	150 µL	98		ACD RNAscope Multiomic HRP-C2			42	15:00	150 µL
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New protocol properties

Name:	ACD RNAscope multiomic Protocol P1
Abbreviated name:	Multi-P1
Description:	ACD RNAscope multiomic Protocol P1
Staining method:	<input checked="" type="checkbox"/> Single <input checked="" type="checkbox"/> Preliminary <input checked="" type="checkbox"/> Final <input checked="" type="checkbox"/> Preferred

BOND RX Import protocol Protocol type: ISH detection

Preferred detection system: ACD Ls Mux Detection Kit

Step N°	Wash	Reagent	Supplier	Ambient	Temperature	Inc. (min)	Dispense type
142		ACD RNAscope Multiomic HRP blocker	Advanced Cell Diagnostics	42	1:00	150 µL	
143		ACD RNAscope Multiomic HRP blocker		42	15:00	150 µL	
150		ACD RNAscope Multiomic HRP-C4		42	1:00	150 µL	
151		ACD RNAscope Multiomic HRP-C4		42	15:00	150 µL	
158		ACD Multiplex TSA-F4		✓	1:00	150 µL	
159		ACD Multiplex TSA-F4		✓	30:00	150 µL	
167		ACD RNAscope Multiomic HRP blocker		42	1:00	150 µL	
168		ACD RNAscope Multiomic HRP blocker		42	15:00	150 µL	

Show wash steps Insert wash | Insert reagent | Delete step

Save Cancel

New protocol properties

Name:	ACD RNAscope multiomic Protocol P1
Abbreviated name:	Multi-P1
Description:	ACD RNAscope multiomic Protocol P1
Staining method:	<input checked="" type="checkbox"/> Single <input checked="" type="checkbox"/> Preliminary <input checked="" type="checkbox"/> Final <input checked="" type="checkbox"/> Preferred

BOND RX Import protocol Protocol type: ISH detection

Preferred detection system: ACD Ls Mux Detection Kit

Step N°	Wash	Reagent	Supplier	Ambient	Temperature	Inc. (min)	Dispense type
160		ACD RNAscope Multiomic HRP blocker		42	1:00	150 µL	
175		ACD RNAscope Multiomic HRP-C5		42	1:00	150 µL	
176		ACD RNAscope Multiomic HRP-C5		42	15:00	150 µL	
183		ACD Multiplex TSA-F5		✓	1:00	150 µL	
184		ACD Multiplex TSA-F5		✓	30:00	150 µL	
192		ACD RNAscope Multiomic HRP blocker		42	1:00	150 µL	
193		ACD RNAscope Multiomic HRP blocker		42	15:00	150 µL	
200		ACD RNAscope Multiomic HRP-C6		42	1:00	150 µL	
201		ACD RNAscope Multiomic HRP-C6		42	15:00	150 µL	

Show wash steps Insert wash | Insert reagent | Delete step

Save Cancel

New protocol properties

Name:	ACD RNAscope multiomic Protocol P1
Abbreviated name:	Multi-P1
Description:	ACD RNAscope multiomic Protocol P1
Staining method:	<input checked="" type="checkbox"/> Single <input checked="" type="checkbox"/> Preliminary <input checked="" type="checkbox"/> Final <input checked="" type="checkbox"/> Preferred

BOND RX Import protocol Protocol type: ISH detection

Preferred detection system: ACD Ls Mux Detection Kit

Step N°	Wash	Reagent	Supplier	Ambient	Temperature	Inc. (min)	Dispense type
200		ACD RNAscope Multiomic HRP-C6			42	15.00	150 µL
201		ACD RNAscope Multiomic HRP-C6			42	15.00	150 µL
208		TSA-DIG		✓		1:00	150 µL
209		TSA-DIG		✓		30:00	150 µL
215		ACD RNAscope Multiomic HRP blocker			42	1:00	150 µL
216		ACD RNAscope Multiomic HRP blocker			42	15:00	150 µL
222		Polaris 780		✓		1:00	150 µL
223		Polaris 780		✓		30:00	150 µL
229		*DAPI	Other	✓		10:00	150 µL

Show wash steps Insert wash | Insert reagent | Delete step

Save Cancel

4. Click **Show wash steps** to also view the wash steps. Insert BOND Washes to match each of the protocol steps shown.
5. Compare and confirm the on-screen protocol with the protocol listed in **Appendix A**.
 - a. Refer to Appendices B-E for other 6-plex assay permutations.
6. Select **Preferred** in the top right corner of the window.

New protocol properties

Name:	ACD RNAscope multiomic Protocol P1
Abbreviated name:	Multi-P1
Description:	ACD RNAscope multiomic Protocol P1
Staining method:	<input checked="" type="checkbox"/> Single <input checked="" type="checkbox"/> Preliminary <input checked="" type="checkbox"/> Final <input checked="" type="checkbox"/> Preferred

BOND RX Import protocol Protocol type: ISH detection

Preferred detection system: ACD Ls Mux Detection Kit

Step N°	Wash	Reagent	Supplier	Ambient	Temperature	Inc. (min)	Dispense type
1		PretreatPro		✓		0:00	150 µL
2		PretreatPro			40	30:00	150 µL
8		*RNAscope 2.5 LSx H ₂ O ₂	Advanced Cell Diagnostics	✓		10:00	150 µL
14		*Open 1	User	✓		10:00	150 µL
15		*Open 1	User		42	120:00	150 µL
27		ACD RNAscope Multiomic Amp 1			42	1:00	150 µL
28		ACD RNAscope Multiomic Amp 1			42	30:00	150 µL
37		*LS Rinse	Advanced Cell Diagnostics	✓		5:00	150 µL
39		*Open 2	Advanced Cell Diagnostics			5:00	150 µL

Show wash steps Insert wash | Insert reagent | Delete step

Save Cancel

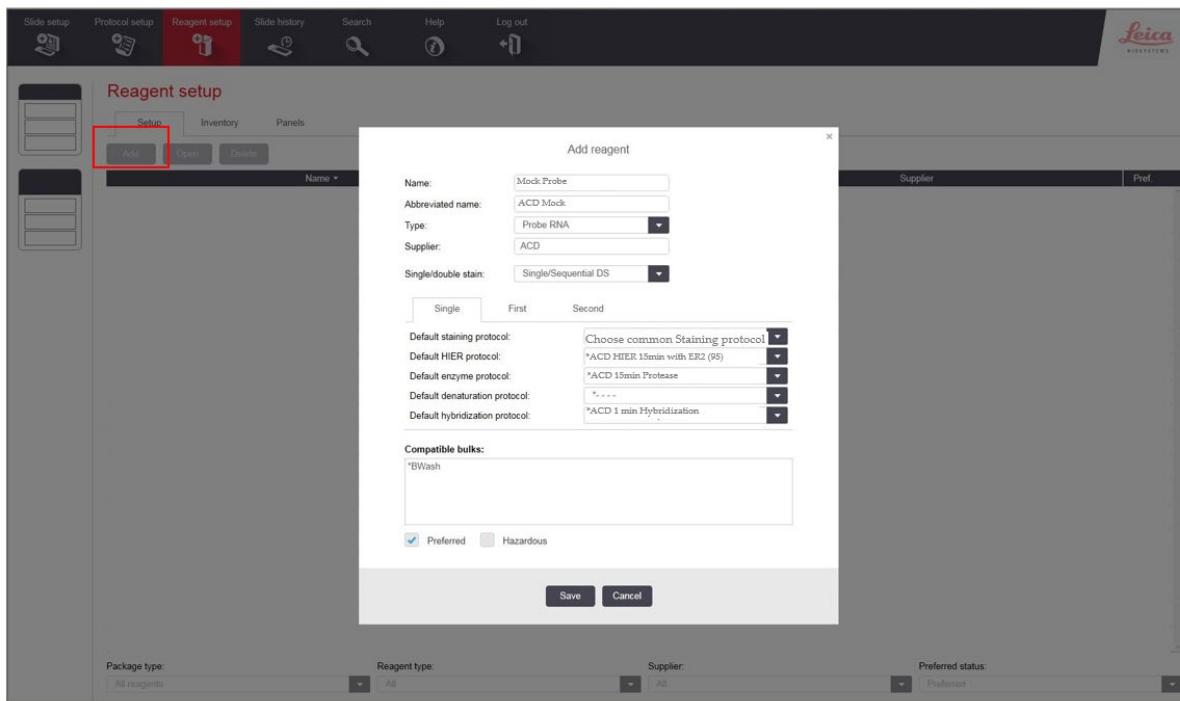
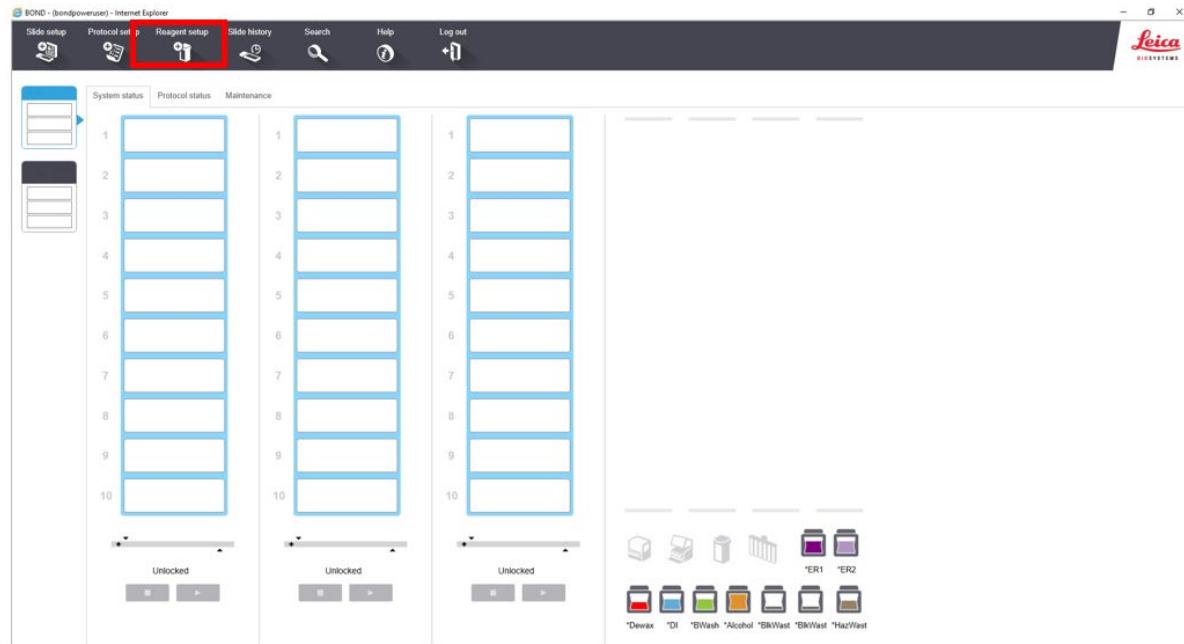
7. Select **Save**.
8. Click **Next** to proceed. Ignore any pop-ups that may appear on the screen.
9. Create a new probe protocol.

Note: You must create a new protocol for each new probe name you use.

Register the mock probe (if not already created)

Create a mock probe in the reagent set up.

1. Click the **Reagent setup** icon.



1.

2. Select **Add**.
3. Enter **Mock Probe** in the Name and **ACD Mock** in Abbreviated name text boxes.
4. Select **Probe RNA** in the Type drop-down menu. Enter **ACD** in the Supplier text box.
5. Select **ACD RNAscope Multiomic Protocol P1** (or your most frequently used protocol) as the Default staining protocol.

6. Select *ACD HIER 15min with ER2 (95) as the Default HIER protocol.
7. Select *ACD 15min Protease as the Default enzyme protocol.
8. Leave the Default denaturation protocol blank.
9. Select ACD 1 min Hybridization as the Default hybridization protocol.
10. Select Save.
11. Fill a 30 mL Open container with 1X Bond Wash from the instrument's bulk container and label it **Mock Probe**. This container will be registered as a Mock Probe in the software.

Prepare the instrument reagents

1. Fill the DAPI container with DAPI and the Bond Wash container with Leica Biosystems' 1X Bond Wash. The kit requires 150 µL DAPI or Bond Wash per slide.
2. Carefully transfer all other RNAscope Multiomic LS kit reagents *except for the TSA buffer* into empty 30 mL BOND Open containers.
3. If you haven't already done so, fill a 30 mL Open container with 1X Bond Wash and register this container as **Mock Probe** in the software.

Note: Before each run, make sure you have enough of each reagent. See the table on page 26 for the reagent volume required per slide.

IMPORTANT! Do not introduce bubbles into the solutions by shaking the containers. To mix reagents, gently invert the containers several times. If bubbles are present, leave the containers out at room temperature until the bubbles dissipate.

Note: You may use your own DAPI, or other counterstain, in place of the DAPI provided in the kit.

4. Fill the *Spectral DAPI container with DAPI and the *Bond Wash container with 1X BOND Wash. The kit requires 150 µL DAPI or 1X BOND Wash per slide.
5. Prepare the LS RNAscope Multiomic target probe mix:
 - a. Determine the volume of probe needed (volume needed for the total number of slides plus container dead-volume). If using Mock Probe workaround, you will require 300 µL of RNAscope probe mix per slide plus dead volume for the container. Make sure to add enough dead-volume to your calculation depending on the container type used:
 - 2.5 mL dead-volume when using a BOND 30 mL Open container.
 - 1 mL dead-volume when using a BOND 7 mL Open container.
 - 600 µL dead-volume when using a BOND Titration container (6 mL).
 - b. Dilute the 50X C2 and C3, C4, C5 and C6 probe stocks 1:50 into the Ready-To-Use C1 probe. For example, add 320 µL 50X C2 probe and 320 µL 50X C3 probe to a tube, then add enough C1 probe to bring the final volume to 16 mL.
 - c. Transfer the LS RNAscope Multiomic probe mix into the appropriate Bond container,
 - d. Reminder: probe containers must be registered as **Ancillary** in the software.

Note: The LS RNAscope Multiomic probe mix is stable for one year at 2–8°C.

6. Prepare the Opal fluorophore dilutions:
 - a. Determine the volume of Opal fluorophore needed, and make sure to add dead volume per container (0.6mL for 6 mL Titration container).
 - b. Dilute the Opal fluorophore stock using the TSA buffer provided in the reagent kit.
 - c. Add the diluted fluorophores to the appropriate Leica containers.

- d. For best results, assign brighter fluorophores to low expressors, develop high expressors last and low expressors first, and assign co-expressing markers to spectrally distinct fluorophores.

Reagents	Recommended dilution range
Opal 480*	1:750–1:3000 (in TSA buffer)
Opal 520*	1:750–1:3000 (in TSA buffer)
Opal 570*	1:750–1:3000 (in TSA buffer)
Opal 620*	1:750 –1:3000 (in TSA buffer)
Opal 690*	1:750–1:3000 (in TSA buffer)
Opal TSA-DIG* (if using Opal 780)	1:750–1:3000 (in TSA buffer)
Opal Polaris 780* (if using Opal 780)	1:187.5–1:750† (in Akoya or Bond diluent)

* Reconstitute all Opals (except Opal Polaris 780) with 75 µL Dimethylsulfoxide (DMSO).

Reconstitute Opal Polaris 780 with 300 µL double distilled water (ddH₂O).

† We recommend keeping the dilution factors of Opal TSA-DIG and Opal Polaris 780 at a constant ratio. For example, when using 1:1500 dilution for Opal TSA-DIG, use 1:375 dilution for Opal Polaris 780. When using 1:750 dilution for Opal TSA-DIG, use 1:187.5 dilution for Opal Polaris 780

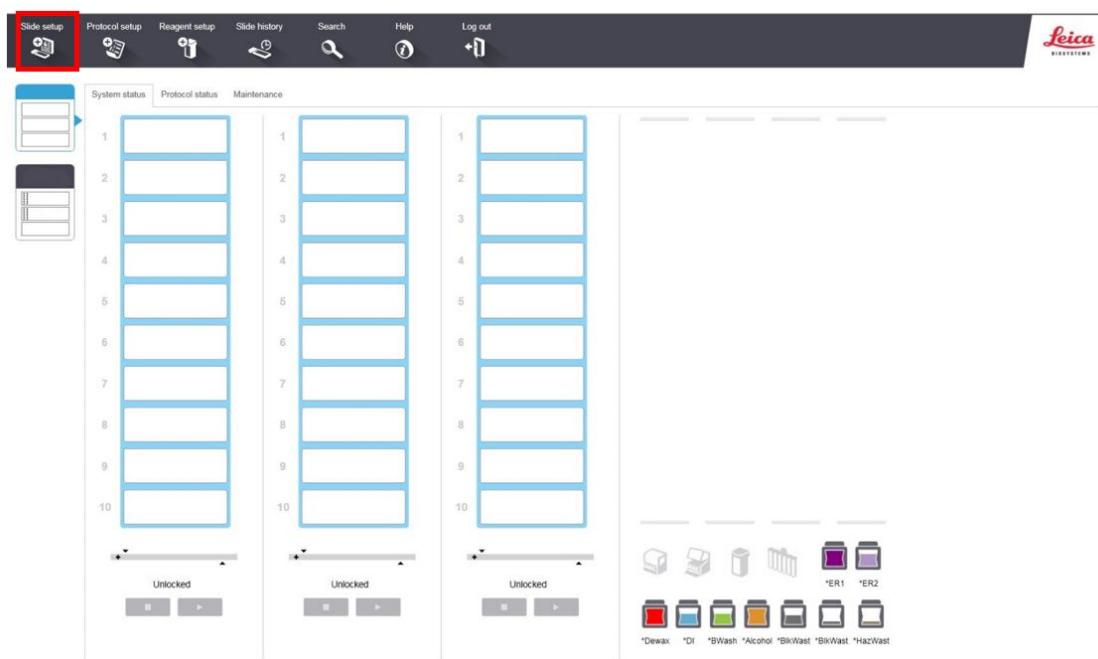
Scan containers into the software

1. Using the Barcode Scanner, scan the barcode located on the front of the BOND Open container. A window appears.
2. From the drop-down menu, select the corresponding name of the reagent as shown in the table on page 26 under **Container name**.

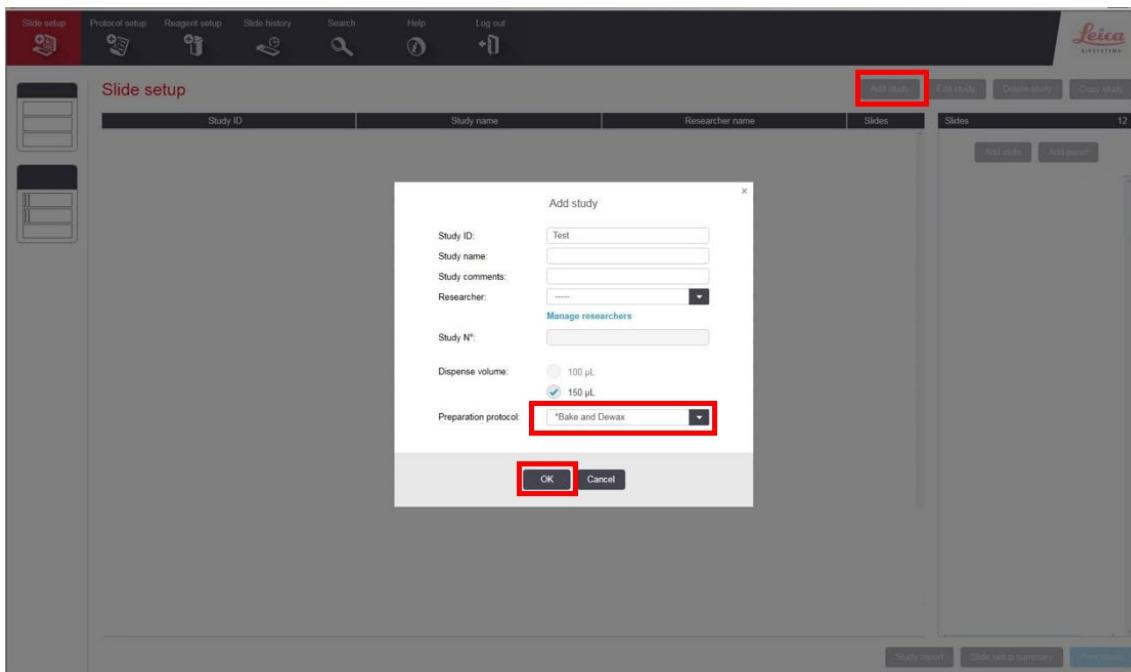
Set up a study for the assay

IMPORTANT! When performing a 6-plex assay, the system can support a maximum of one tray. Follow the protocol steps listed in **Appendices A-E**. Adding any additional steps will prevent the system from running the experiment.

1. To build a study for the RNAscope Multiomic LS Assay, select the **Slide setup** icon at the top of the screen.

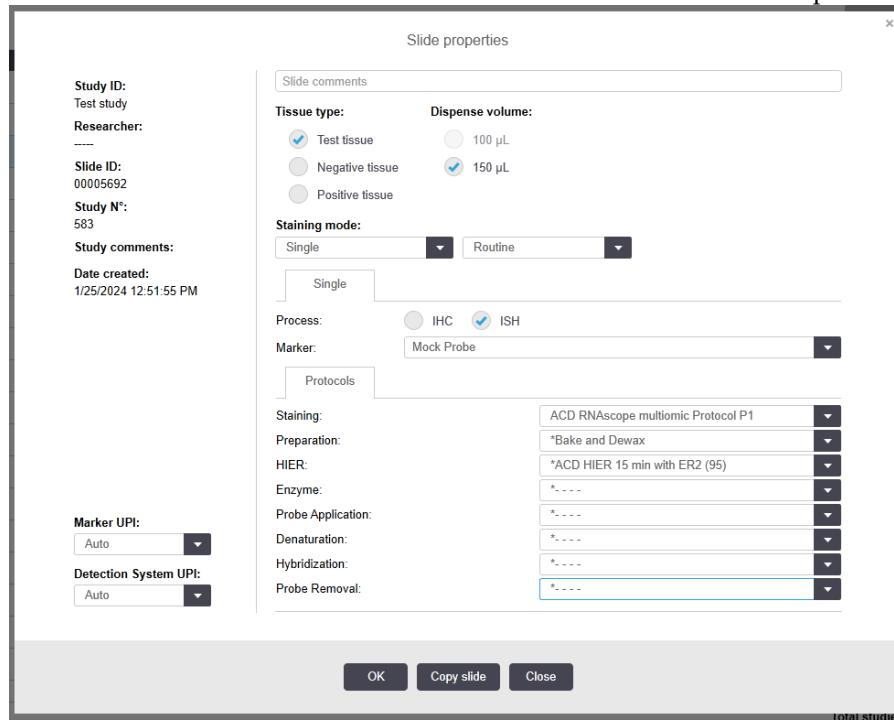


2. Select Add study.



3. Enter a name in the Study ID field (keep the Dispense volume at **150 µL** as shown).
 - a. For FFPE tissues, select ***Bake and Dewax** as the Preparation protocol.
 - b. For fixed-frozen or fresh-frozen tissues, select ***----** instead.
4. Select **OK**.
5. Select **Add slide** to assign a protocol to each slide.

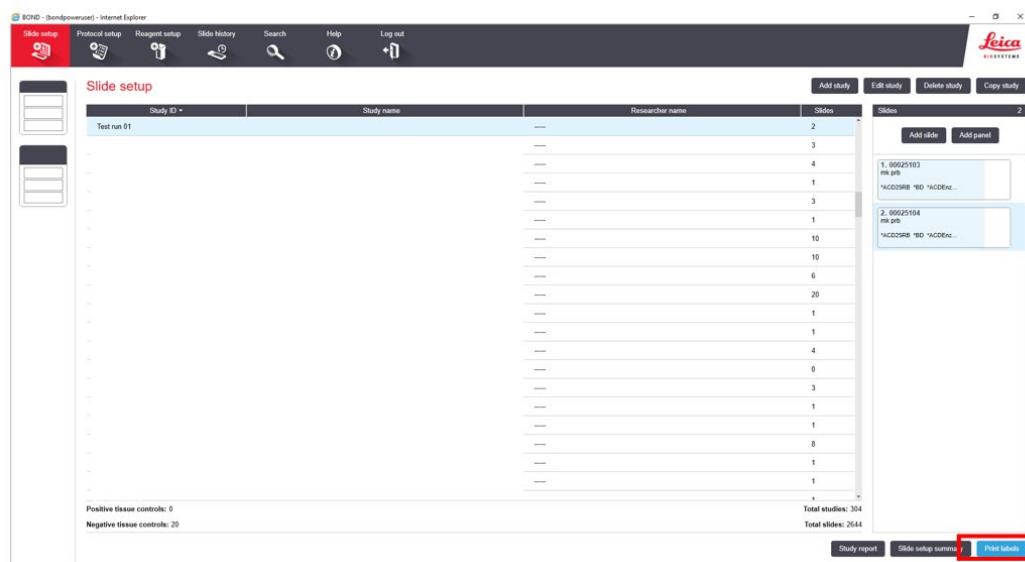
6. Enter the tissue type and probe name under the Comments field.
7. Keep **Single** as default from the Staining mode drop down menu.
8. Select **ISH** under Process and **Mock Probe (ACD)** from the Marker drop down menu.



+

9. Under the **Protocols** tab, do the following:
 - For *each* distinct probe, select an *applicable* protocol from the Staining drop down menu (for example, RNAscope Multiomic Protocol P1).
 - For standard FFPE tissues, select the protocol ***Bake and Dewax** from the Preparation drop down menu. For fixed or fresh frozen tissues, select ***----** instead.

- Select *ACD HIER 15 min with ER2 (95) as the HIER protocol or the appropriate HIER protocol for your tissue.
 - Select *ACD 15 min Protease for Enzyme, or the appropriate enzyme protocol for your tissue.
 - a. Select *---- for Probe Application
 - Select *---- for Denaturation
 - Select *---- for Hybridization.
 - Select *____ for Probe Removal
10. Select **Add slide** for each target probe and for each of the slides used in the run.
 11. After adding all the slides to the study, select **Close** to return to the Slide setup screen.
 12. Select **Print labels** to print barcodes to attach to the slides.

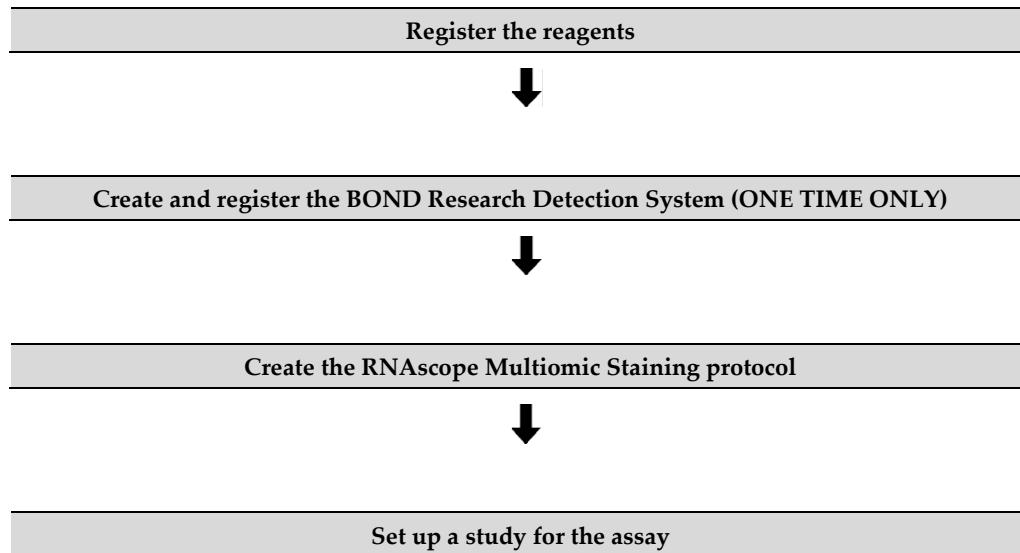


6

Chapter 6. Set Up a Staining Protocol without the Mock Probe

If you are staining RNA targets only, use the instructions in this chapter to set up the RNAscope Multiomic LS Assay without mock probe workaround recommended for LS software versions 6.0 and 7.0.

Workflow



Register the reagents

Your ACD Field Application Specialist (FAS) can help implement this procedure.

1. Select the **Reagent Setup** icon at the top of the screen.
2. Select **Add** to enter reagent information for components that are not pre-filled in the Leica software.
3. Enter a reagent name in the Name text box (see below Table)
4. Select **Ancillary** in the Type drop-down menu.
5. Enter **ACD** in the Supplier text box.
6. Check both the **Preferred** and **Hazardous** boxes (for RNAscope Multiomic Amp 1, and RNAscope Multiomic Amp 3 only).
7. Select **Save**.
8. Repeat Steps 2–7 for the rest of the reagents using the container names in the following table:

Reagents	Container Name	Abbreviated Name	Volume per slide (dead volume not included)
RNAscope Multiomic LS AMP 1	RNAscope Multiomic Amp 1	MOAmp1	300 µL
RNAscope Multiomic LS AMP 2	RNAscope Multiomic Amp 2	MOAmp2	300 µL

Reagents	Container Name	Abbreviated Name	Volume per slide (dead volume not included)
RNAscope Multiomic LS AMP 3	RNAscope Multiomic Amp 3	MOAmp3	300 µL
RNAscope Multiomic LS HRP C1	RNAscope Multiomic HRP-C1	MOHRPC1	300 µL
RNAscope Multiomic LS HRP C2	RNAscope Multiomic HRP-C2	MOHRPC2	300 µL
RNAscope Multiomic LS HRP C3	RNAscope Multiomic HRP-C3	MOHRPC3	300 µL
RNAscope Multiomic LS HRP C4	RNAscope Multiomic HRP-C4	MOHRPC4	300 µL
RNAscope Multiomic LS HRP C5	RNAscope Multiomic HRP-C5	MOHRPC5	300 µL
RNAscope Multiomic LS HRP C6	RNAscope Multiomic HRP-C6	MOHRPC6	300 µL
RNAscope Multiomic LS HRP Blocker	RNAscope Multiomic HRP Blocker	MOHRPbk	1800 µL
Opal-fluorophore 1 (user to dilute in TSA buffer)	Multiomic TSA-F1	MOTSAF1	300 µL
Opal-fluorophore 2 (user to dilute in TSA buffer)	Multiomic TSA-F2	MOTSAF2	300 µL
Opal-fluorophore 3 (user to dilute in TSA buffer)	Multiomic TSA-F3	MOTSAF3	300 µL
Opal-fluorophore 4 (user to dilute in TSA buffer)	Multiomic TSA-F4	MOTSAF4	300 µL
Opal-fluorophore 5 (user to dilute in TSA buffer)	Multiomic TSA-F5	MOTSAF5	300 µL
Opal TSA-DIG † (if Opal 780 is used)	TSA-DIG	TSA-DIG	300 µL
Opal 780‡ (if Opal 780 is used)	Opal 780	Opal780	300 µL
LS RNAscope Multiomic probe mix	(use any container but make sure that the container is registered as Probe RNA in the software)	—	370 µL for 6.0/7.0 software
RNAscope Multiomic LS DAPI	DAPI	DAPI	150 µL
BOND Wash	Bond Wash	BondWash	150 µL
RNAscope Multiomic LS Hydrogen Peroxide	*RNAscope 2.5 LSx H2O2	—	150 µL
RNAscope Multiomic LS Protease III	*RNAscope 2.5 LSx Protease	—	200 µL
RNAscope Multiomic LS Rinse	*LS Rinse	—	600 µL

*Indicates this reagent is hard coded in the software by Leica Biosystems.

†These reagents are only needed when Opal Polaris 780 is used in the assay.

Create and register the BOND Research Detection System (one time only)

A BOND Research Detection System from Leica is required to set up the RNAscope Multiomic LS Assay. Your ACD Field Application Specialist (FAS) should implement this procedure. Each detection system barcode is valid for up to 40 mL of use (equivalent to ~260 slides or four RNAscope Multiomic LS Fluorescent Reagent Kits).

1. Scan the barcode on the tray of a new BOND Research Detection System.
2. To setup a new detection system for the assay, enter **ACD LS Multiplex Detection Kit** in the Name text box.

Note: Creating the detection system needs to be performed only once on each BOND RX controller.

Pstn.	UPI	Reagent	Vol. (mL)
1	20029225	DAPI	
2	20029225	Bond Wash	

3. Place two new BOND 30 mL Open containers on the Research Detection System rack.
4. Scan the first container and select the registration name **DAPI**. You can mix different lots of DAPI in the same container.

Note: If you prefer not to use DAPI on the instrument or want to perform immunohistochemistry (IHC) steps after the assay, you may use BOND Wash in place of DAPI in the protocol.

5. Scan the second container and select the registration name **Bond Wash**.
6. When one Research Detection System is finished (up to 40 mL), register a new detection system by scanning the barcode on the tray and select **ACD LS Multiplex Detection Kit** from the drop-down menu on the right.
7. Select **Add**.

Create a staining protocol

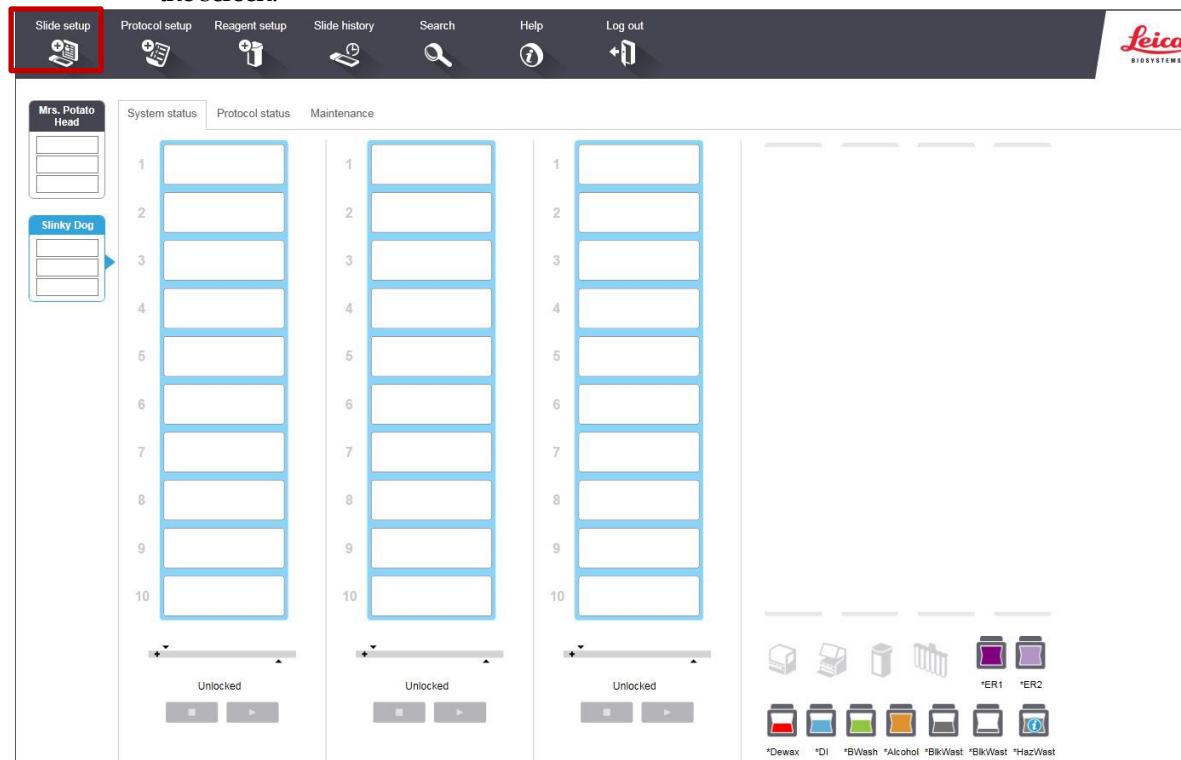
Your ACD Field Application Specialist (FAS) can help implement this procedure.

1. In the Protocol setup screen, select **Staining** under the Protocol drop down menu.

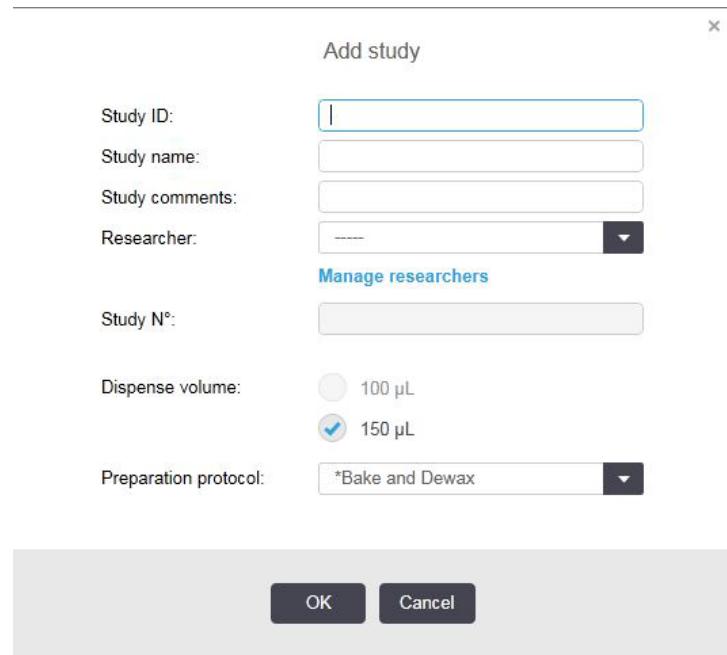
2. Highlight the *ACD 2.5 DAB Rev B protocol. Select **Copy**.
3. Change the protocol name to **ACD RNAscope Multiomic Protocol (or 6 mRNA)** in the Name text box, **Multi_P1** in the Abbreviated name text box, and **ACD RNAscope Multiomic Protocol** in the Description text box.
4. Select ACD LS Multiplex Detection Kit from the Preferred detection system menu.
5. Edit the protocol on-screen to match the protocol listed in Compare and confirm the on-screen protocol with the protocol listed in **Appendix B**.
6. Highlight and select each step to edit. Click **Show wash steps** to also view the wash steps. Insert BOND Washes to match each of the protocol steps shown.
7. Once the staining protocol matches the steps in **Appendix B**, delete steps 1–14. **ACD Multiomic Amp1** should be steps 1 and 2.
8. Select **Preferred** in the bottom right corner of the window. Select **Save**.

Set up a study for the RNAscope 6-plex LS Assay

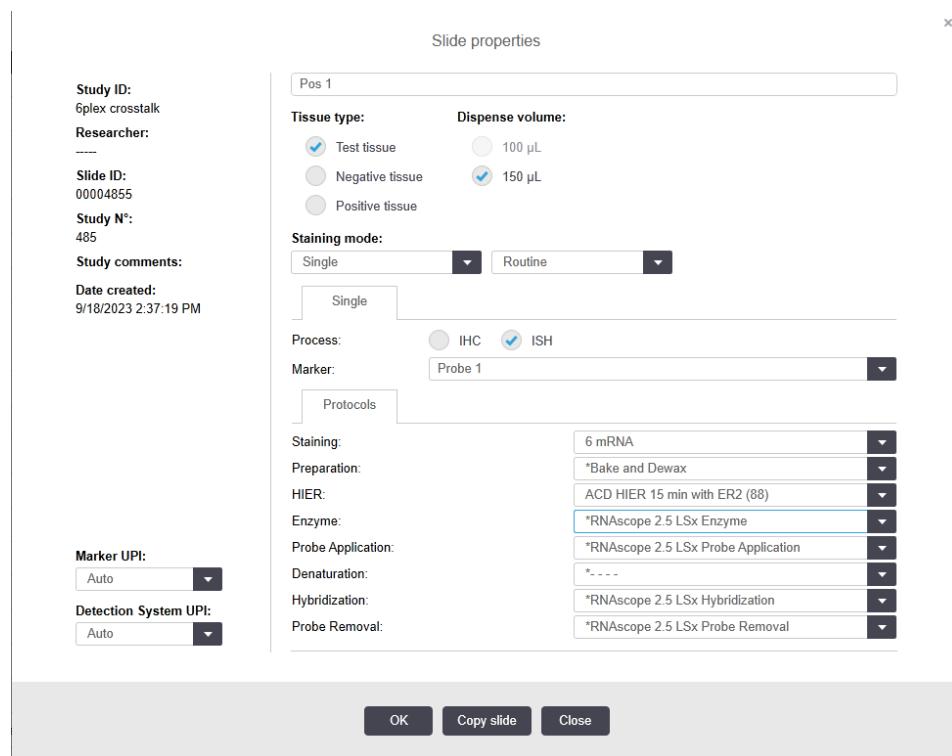
1. To build a study for the RNAscope 6-plex LS Assay, select the **Slide setup** icon at the top of the screen.



2. Select **Add study** and enter a name in the Study ID field (keep the Dispense volume at 150 µL as shown).



3. For FFPE tissues, select ***Bake and Dewax** as the Preparation protocol. For frozen tissues, select **----** instead.
4. Select **OK**.
5. Select **Add slide** to assign a protocol to each slide.
6. Enter the tissue type and probe name under the Comments field.
7. Keep **Single** as default from the Staining mode drop down menu.
8. Select **ISH** under Process and **the appropriate probe** from the Marker drop down menu.



9. Under the **Protocols** tab, do the following:

- a. Select the appropriate protocol from the Staining drop down menu (for example, ACD RNAscope Multiomic Protocol).
- b. For standard FFPE tissues, select the protocol ***Bake and Dewax** from the Preparation drop down menu. For frozen tissues, select *---- instead.
- c. Select ***RNAscope 2.5 LSx Target Retrieval (88)** as the HIER protocol or the appropriate HIER protocol for your tissue.
- d. Select ***RNAscope 2.5 LSx Enzyme**, or the appropriate enzyme protocol for your tissue. (if Pretreat Pro is being used it must be placed in a container and registered as LSx enzyme and used with an edited protocol; see Appendix K).
- e. Select ***RNAscope 2.5 LSx Probe Application** from the Probe Application dropdown menu.
- f. Select * ---- for Denaturation.
- g. Select ***RNAscope 2.5 LSx Hybridization** for Hybridization.
- h. Select ***RNAscope 2.5 LSx Probe Removal** for Probe Removal.
10. Select **Add slide** for each target probe and for each of the slides used in the run.
11. After adding all the slides to the study, select **Close** to return to the Slide setup screen.
12. Select **Print labels** to print barcodes to attach to the slides.

Study ID	Study name	Researcher name	Slides
Test run 01		—	2
		—	3
		—	4
		—	1
		—	3
		—	1
		—	10
		—	10
		—	6
		—	20
		—	1
		—	1
		—	4
		—	0
		—	3
		—	1
		—	1
		—	8
		—	1
		—	1

Total studies: 304
Total slides: 2044

Positive tissue controls: 0
Negative tissue controls: 20

Leica
ASIST SYSTEMS

Add study Edit study Delete study Copy study

Slides 2

Add slide Add panel

1. 000252403.msf.pbf *ACD25R-BD *ACDEnz...

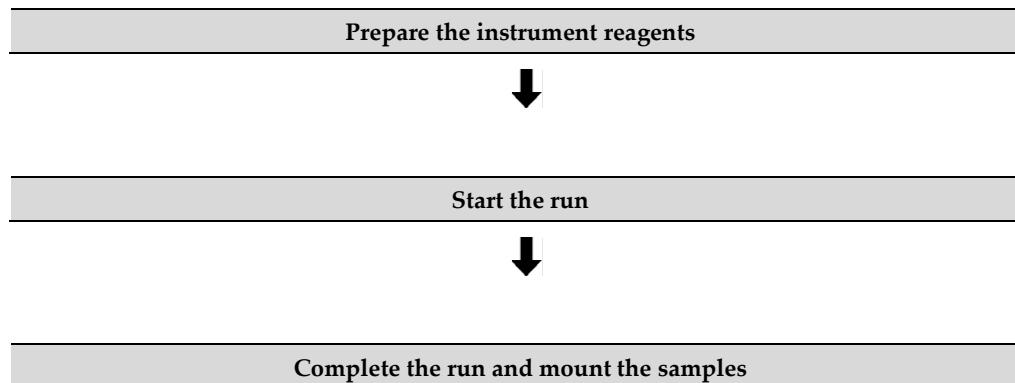
2. 000252104.msf.pbf *ACD25R-BD *ACDEnz...

Study report Slide setup summary Print labels

7

Chapter 7. Run the Multiomic LS Assay

Workflow



Prepare the instrument reagents

1. Fill the DAPI container with DAPI and the Bond Wash container with Leica Biosystems' 1X BOND Wash. The kit requires 150 µL DAPI or Bond Wash per slide.
2. Carefully transfer all other RNAscope LS kit reagents *except for the TSA buffer* into empty 30 mL BOND Open containers.

Note: Before each run, make sure you have enough of each reagent. See the table on page 40 for the reagent volume required per slide.

IMPORTANT! Do not introduce bubbles into the solutions by shaking the containers. To mix reagents, gently invert the containers several times. If bubbles are present, leave the containers out at room temperature until the bubbles dissipate.

Note: You may use your own DAPI, or other counterstain, in place of the DAPI provided in the kit.

3. Prepare the RNAscope LS target probe mix:
 - a. Determine the volume of probe needed (volume needed for the total number of slides plus container dead-volume). For software 6.0/7.0, you will require 370 uL of probe per slide plus dead volume for the container. Make sure to add enough dead-volume to your calculation depending on the container type used:
 - 2.5 mL dead-volume when using a BOND 30 mL Open container.
 - 1 mL dead-volume when using a BOND 7 mL Open container.
 - 600 µL dead-volume when using a BOND Titration container (6 mL).
 - b. Dilute the 50X C2, C3, C4, C5 and C6 probe stocks 1:50 into the Ready-To-Use C1 probe. For example, add 320 µL of each of 50X C2, 50X C3, 50X C4, 50X C5 and 50X C6 probe to a tube, then add enough C1 probe to bring the final volume to 16 mL.
 - c. Transfer the RNAscope Multiomic LS probe mix into the appropriate Bond container, probe containers must be registered as **Probe RNA** in the software.

Note: The RNAscope probe mix is stable for one year at 2–8°C.

4. If performing an RNA + Protein staining, prepare conjugated antibodies:

- Determine the volume of antibody mix needed (see Appendices C, D, and E for suggested concentration of each antibody) and add extra amount for dead volume of the container.
- Dilute primary antibody conjugates together into one tube.
- Dilute the antibody conjugates using the multiomic antibody diluent provided with the antibody.
- If using a secondary conjugate, use a separate container to dilute it.

IMPORTANT! Do not pool secondaries with primary antibody conjugates.

1. Add diluted antibody conjugates to the appropriate Leica containers.

2. Prepare the Opal fluorophore dilutions:

- Determine the volume of Opal fluorophore needed and make sure to add dead volume per container. Each slide requires 300 µL of fluorophore for staining.
- Dilute the Opal fluorophore stock using the TSA buffer provided in the reagent kit.
- Add the diluted fluorophores to the appropriate Leica containers.

Reagents	Recommended dilution range	Dye Intensity
Opal 480*	1:750–1:3000 (in TSA buffer)	Highest
Opal 520*	1:750–1:3000 (in TSA buffer)	Highest
Opal 570*	1:750–1:3000 (in TSA buffer)	Medium
Opal 620*	1:750 -1:3000 (in TSA buffer)	Medium
Opal 690*	1:750–1:3000 (in TSA buffer)	Low
Opal TSA-DIG* (if using Opal 780)	1:750–1:3000 (in TSA buffer)	Lowest
Opal Polaris 780* (if using Opal 780)	1:187.5–1:750† (in Akoya or Bond diluent)	Lowest

* Reconstitute all Opals (except Opal Polaris 780) with 75 µL Dimethylsulfoxide (DMSO).

Reconstitute Opal Polaris 780 with 300 µL double distilled water (ddH₂O).

† We recommend keeping the dilution factors of Opal TSA-DIG and Opal Polaris 780 at a constant ratio. For example, when using 1:1500 dilution for Opal TSA-DIG, use 1:375 dilution for Opal Polaris 780. When using 1:750 dilution for Opal TSA-DIG, use 1:187.5 dilution for Opal Polaris 780.

5. For best results, assign brighter fluorophores to low expressors, develop high expressors last and low expressors first, and assign co-expressing markers to spectrally distinct fluorophores.

Scan containers into the software

- Using the Barcode Scanner, scan the barcode located on the front of the BOND Open container. A window appears.
- From the drop-down menu, select the corresponding name of the reagent as shown in the table on page 40 under **Container name**. Then click **Save**.

Prepare the instrument

1. Fill the large containers located in the bottom of the instrument with the Leica BOND RX bulk reagents.
2. Dilute BOND Wash Solution 1:10.

Note: Insufficient bulk reagent volumes may lead to run failure.

IMPORTANT! Do not introduce bubbles into the solutions by shaking the containers. To mix reagents, gently invert the containers several times. If bubbles are present, leave the containers out at room temperature until the bubbles dissipate.

3. Use clean, dry covertiles for every run. Follow Leica instructions to clean used covertiles with water, bleach, and ethanol. Air dry before reuse.
4. Before starting a run, empty bulk waste containers. Discard waste according to all local, state/provincial and/or national regulations.

Start the run

1. Attach the barcodes to the slides and add the slides to the slide tray with the label sides facing up.
2. Add a covertile on top of each slide and verify placement and seating of each covertile.

Note: The rectangular-shaped neck of the covertile should fit into the groove of the slide tray.

3. Place the tray in the Leica BOND RX and press the button to load the tray onto the machine.
4. Once the slides have been scanned, select the **PLAY** (triangular) button on the screen located under the start tray to start the run. Alternatively, right-click on scanned label images, and select **Delayed Start** to start the run at a future time. Do not use Delayed Start with fixed or freshfrozen tissue.

Note: The total run time for LS RNAscope Multiomic Fluorescent Assay is 14–15 hrs depending on the number of slides.

IMPORTANT! Before leaving the instrument unattended, ensure that the instrument is running successfully.

Complete the run and mount the samples

1. After the run is complete, press the button on the front of the instrument to unload the slides.
2. Remove the covertiles.
3. Add a drop of ProLong Gold Antifade Mountant to each slide. Avoid introducing bubbles.
4. Carefully place a glass coverslip on the slides, and dry overnight in the dark.
5. Store the slides at 4°C in the dark for up to two weeks.

Chapter 8. Evaluate the Results

Evaluate the samples

Examine tissue sections under a standard fluorescent microscope at 20–40X magnification. You may also use a confocal microscope.

- Assess tissue and cell morphology.
- Assess the negative control background first. One dot to every 10 cells displaying background staining per 20X microscope field is acceptable. Set the light source and exposure time of image acquisition to acceptable background levels.
- Assess positive control signal strength. Positive control signal should be visible as punctate dots within the cell at 20–40X magnification.

Scoring Guidelines

When used for RNA detection, the assay enables a semiquantitative scoring guideline utilizing the estimated number of punctate dots present within each cell boundary. An example of how to develop such a guideline for semi-quantitative assessment of mRNA staining intensity is presented below for a gene with expression level varying between 1 to > 10 dots per cell.

Note: If your gene expression level is higher or lower than this range, you may need to scale the criteria accordingly.

mRNA scoring is divided into five categories:

Staining Score	Microscope Objective Scoring
0	No staining or less than 1 dot per 10 cells
1	1–3 dots per cell
2	4–9 dots per cell, no or very few dot clusters
3	10–15 dots per cell and/or <10% dots are in clusters
4	>15 dots per cell and/or >10% dots are in clusters

Control examples

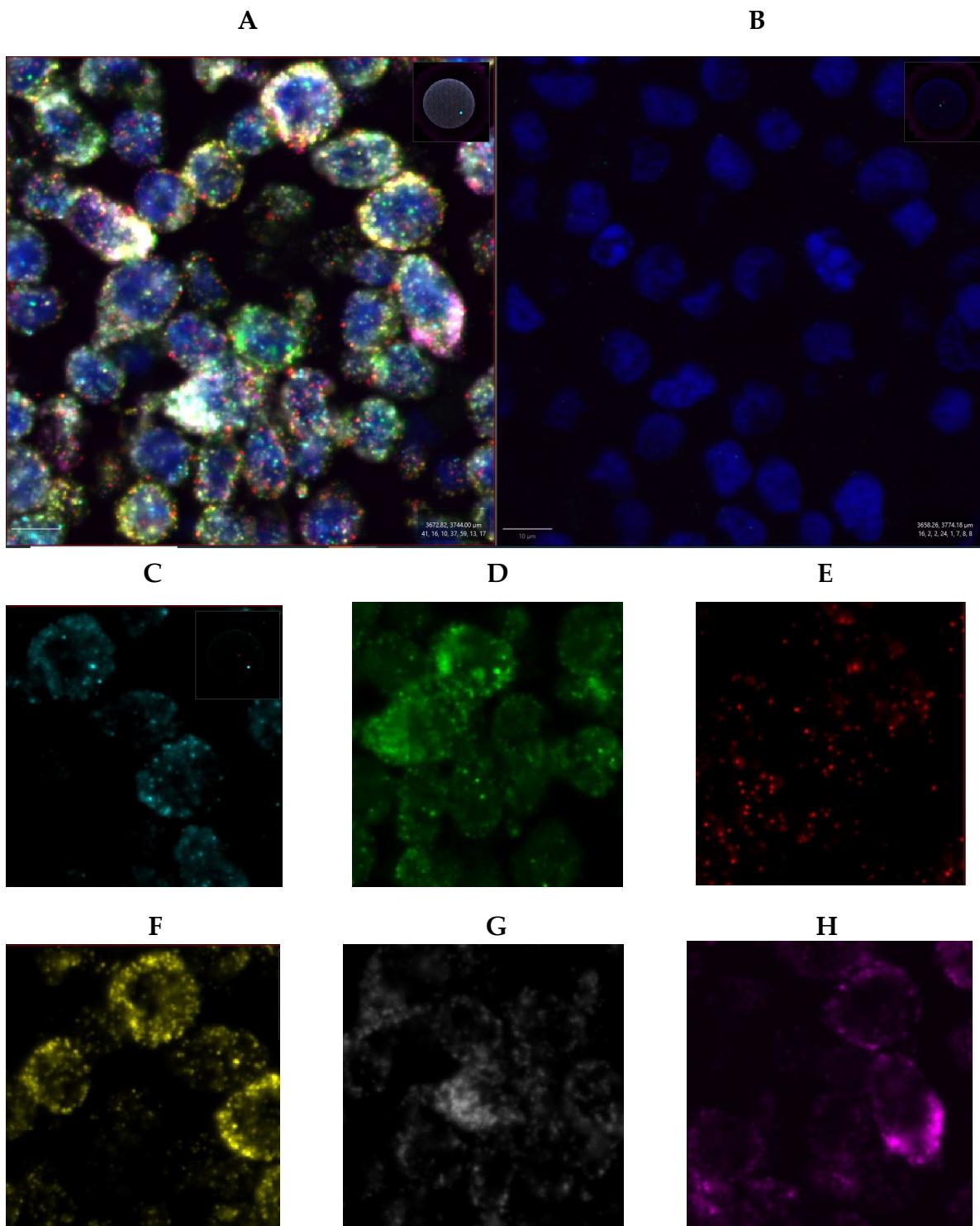


Figure 2: mRNA staining examples from FFPE HeLa cell pellet using RNAscope Multiomic LS Fluorescent Assay at 40X magnification. **A)** Positive control slide showing 6 RNA targets: *POLR2A* (teal), *ACTB* (green), *HPRT1* (red), *UBC* (yellow), *PPIB* (white), *TUBB* (pink). **B)** Negative control slide with *dapB*. **C-H)** Individual channels at higher magnification.

If the assay is successful, the staining should look like the following image:

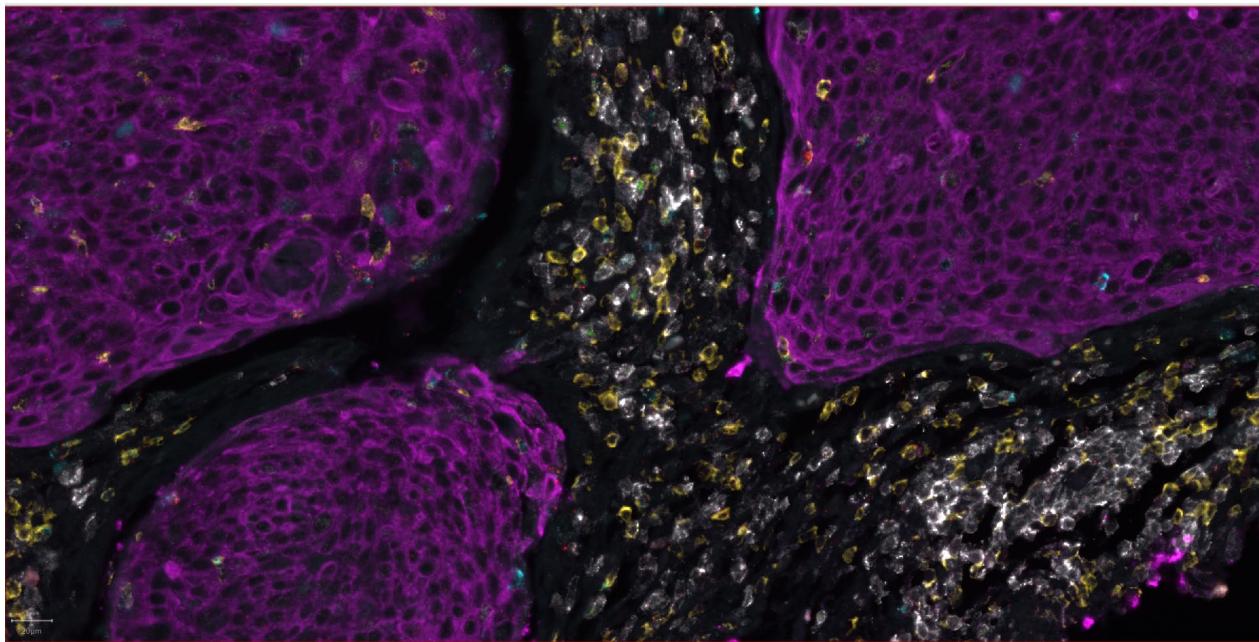


Figure 3. RNAscope Multiomic LS Fluorescent Assay detection of 2 RNAs and 3 proteins. *IFNg* (red) *GZMB* (blue) CD4 (yellow) CD8a (white) PanCK (pink) in Cervical cancer FFPE tissue at 20X magnification.

Troubleshooting

If you obtain less than satisfactory results, troubleshoot your assay by following these simple guidelines:

- Always use optimal fluorescent filter settings and imaging tools.
 - If signal intensity is too low for your imaging tools, increase the fluorophore concentration.
 - Use optimized fluorescence filter sets to reduce signal bleed-through. If you observe fluorescence bleed-through, reduce the fluorophore concentration of the bleeding-through channel and/or reduce the exposure time during image acquisition to avoid over-exposure.
 - If your RNA ISH signal cannot be distinguished from autofluorescence in tissues with high autofluorescence, increase the fluorophore concentration.
 - If you observe the presence of background staining, limit the sensitivity of image acquisition or reduce the corresponding Opal fluorophore concentration. Always acquire images using the setting in which background is under-detected.
 - If the signal-to-noise ratio is low due to high background, optimize pretreatment conditions. Contact ACD support for recommendations.
 - The RNAscope RNAscope Multiomic LS Fluorescence Assay uses only the Leica Biosystem BOND Research Detection System. Do not use BOND Polymer Refine DAB/Red Detection kits or any other chromogen kits.
 - Do not shake the contents in the dispensers as this will form bubbles and may lead to weak or no staining. If bubbles are present, leave the containers out at room temperature until the bubbles dissipate.
 - **Target retrieval for 20min at 100°C can be used if dye trapping or lower antibody signal is observed.**
 - For troubleshooting information, please contact technical support at support.acd@bio-techne.com.

A

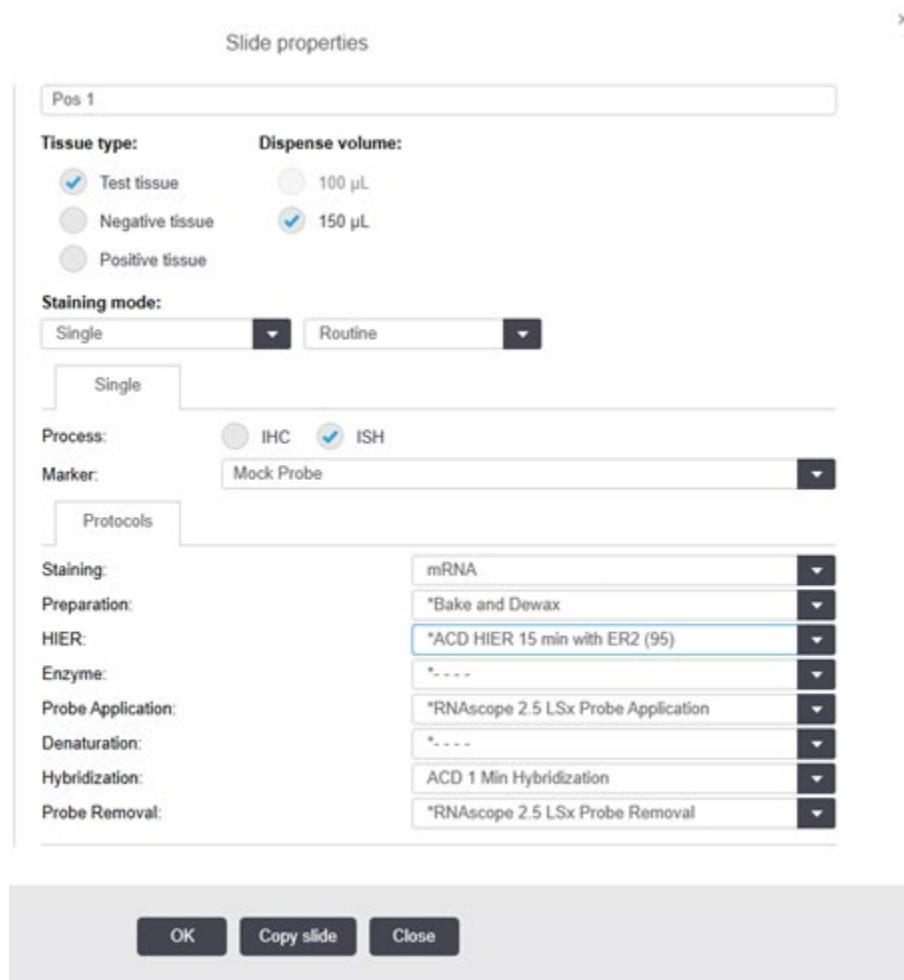
Appendix A. Protease-Free RNA 6-plex Staining Protocol (using the Mock Probe workaround)

You can perform a 6-plex RNA-only run using the workflow described in Chapter 6, which does not use the Mock Probe Reagent.

IMPORTANT! Heated *Bond Wash solution steps come from the bulk reagents and are heated by the instrument. You cannot delete these steps. You may delete other wash steps.

IMPORTANT! The BOND RX can only execute this workflow on one tray for the RNAscope Multiomic assay (six channels) due to its length, follow the protocol steps listed in the following table. Adding any additional steps will prevent the system from running even one trays.

Slide Setup:



Step No.	Reagent	Step Type	Incubation Time	Temperature†
1	RNAscope LS PretreatPro	Reagent	0 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
2	RNAscope LS PretreatPro	Reagent	30 MIN	40°C
3	*Bond Wash Solution	Wash	0 MIN	Ambient
4	*Bond Wash Solution	Wash	0 MIN	Ambient
5	*Bond Wash Solution	Wash	0 MIN	Ambient
6	*Bond Wash Solution	Wash	1 MIN	Ambient
7	*Bond Wash Solution	Wash	1 MIN	Ambient
8	RNAscope Multiomic LS Hydrogen Peroxide	Reagent	10 MIN	Ambient
9	*Bond Wash Solution	Wash	0 MIN	Ambient
10	*Bond Wash Solution	Wash	0 MIN	Ambient
11	*Bond Wash Solution	Wash	0 MIN	Ambient
12	*Bond Wash Solution	Wash	0 MIN	Ambient
13	*Bond Wash Solution	Wash	0 MIN	Ambient
14	Probe 1	Reagent	0 MIN	Ambient
15	Probe 1	Reagent	120 MIN	42°C
16	*Bond Wash Solution	Wash	0 MIN	42°C
17	*Bond Wash Solution	Wash	1 MIN	42°C
18	*Bond Wash Solution	Wash	5 MIN	42°C
19	*Bond Wash Solution	Wash	0 MIN	Ambient
20	*Bond Wash Solution	Wash	0 MIN	Ambient
21	*Bond Wash Solution	Wash	0 MIN	Ambient
22	*Bond Wash Solution	Wash	0 MIN	Ambient
23	*Bond Wash Solution	Wash	0 MIN	Ambient
24	*Bond Wash Solution	Wash	1 MIN	Ambient
25	*Bond Wash Solution	Wash	1 MIN	Ambient
26	*Bond Wash Solution	Wash	0 MIN	Ambient
27	RNAscope Multiomic LS Amp 1	Reagent	1 MIN	42°C
28	RNAscope Multiomic LS Amp 1	Reagent	30 MIN	42°C
29	*Bond Wash Solution	Wash	0 MIN	Ambient
30	*Bond Wash Solution	Wash	0 MIN	Ambient
31	*Bond Wash Solution	Wash	0 MIN	Ambient
32	*Bond Wash Solution	Wash	3 MIN	Ambient
33	*Bond Wash Solution	Wash	3 MIN	Ambient
34	*Bond Wash Solution	Wash	0 MIN	Ambient
35	*Bond Wash Solution	Wash	0 MIN	Ambient
36	*Bond Wash Solution	Wash	0 MIN	Ambient
37	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
38	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
39	*Bond Wash Solution	Wash	0 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
40	*Bond Wash Solution	Wash	0 MIN	Ambient
41	*Bond Wash Solution	Open Wash	0 MIN	Ambient
42	*Bond Wash Solution	Wash	0 MIN	Ambient
43	RNAscope Multiomic LS Amp 2	Reagent	1 MIN	42°C
44	RNAscope Multiomic LS Amp 2	Reagent	30 MIN	42°C
45	*Bond Wash Solution	Wash	0 MIN	Ambient
46	*Bond Wash Solution	Wash	0 MIN	Ambient
47	*Bond Wash Solution	Wash	0 MIN	Ambient
48	*Bond Wash Solution	Wash	3 MIN	Ambient
49	*Bond Wash Solution	Wash	3 MIN	Ambient
50	*Bond Wash Solution	Wash	0 MIN	Ambient
51	*Bond Wash Solution	Wash	0 MIN	Ambient
52	*Bond Wash Solution	Wash	0 MIN	Ambient
53	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
54	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
55	*Bond Wash Solution	Wash	0 MIN	Ambient
56	*Bond Wash Solution	Wash	1 MIN	Ambient
57	*Bond Wash Solution	Open Wash	1 MIN	Ambient
58	*Bond Wash Solution	Wash	1 MIN	Ambient
59	RNAscope Multiomic LS Amp 3	Reagent	1 MIN	42°C
60	RNAscope Multiomic LS Amp 3	Reagent	15 MIN	42°C
61	*Bond Wash Solution	Wash	0 MIN	Ambient
62	*Bond Wash Solution	Wash	0 MIN	Ambient
63	*Bond Wash Solution	Wash	0 MIN	Ambient
64	*Bond Wash Solution	Wash	1 MIN	Ambient
65	*Bond Wash Solution	Wash	1 MIN	Ambient
66	*Bond Wash Solution	Wash	1 MIN	Ambient
67	*Bond Wash Solution	Open Wash	1 MIN	Ambient
68	*Bond Wash Solution	Wash	1 MIN	Ambient
69	RNAscope Multiomic LS HRP C1	Reagent	1 MIN	42°C
70	RNAscope Multiomic LS HRP C1	Reagent	15 MIN	42°C
71	*Bond Wash Solution	Wash	0 MIN	Ambient
72	*Bond Wash Solution	Wash	0 MIN	Ambient
73	*Bond Wash Solution	Wash	0 MIN	Ambient
74	*Bond Wash Solution	Wash	1 MIN	Ambient
75	*Bond Wash Solution	Wash	1 MIN	Ambient
76	*Bond Wash Solution	Wash	1 MIN	Ambient
77	*Bond Wash Solution	Wash	1 MIN	Ambient
78	*Bond Wash Solution	Wash	1 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
79	RNAscope Multiomic TSA-F1	Reagent	1 MIN	Ambient
80	RNAscope Multiomic TSA-F1	Reagent	30 MIN	Ambient
81	*Bond Wash Solution	Wash	0 MIN	Ambient
82	*Bond Wash Solution	Wash	0 MIN	Ambient
83	*Bond Wash Solution	Wash	0 MIN	Ambient
84	*Bond Wash Solution	Wash	1 MIN	Ambient
85	*Bond Wash Solution	Wash	1 MIN	Ambient
86	*Bond Wash Solution	Wash	1 MIN	Ambient
87	*Bond Wash Solution	Wash	1 MIN	Ambient
88	RNAscope Multiomic LS HRP Blocker	Reagent	1 MIN	42°C
89	RNAscope Multiomic LS HRP Blocker	Reagent	15 MIN	42°C
90	*Bond Wash Solution	Wash	0 MIN	Ambient
91	*Bond Wash Solution	Wash	0 MIN	Ambient
92	*Bond Wash Solution	Wash	0 MIN	Ambient
93	*Bond Wash Solution	Wash	1 MIN	Ambient
94	*Bond Wash Solution	Wash	1 MIN	Ambient
95	*Bond Wash Solution	Wash	1 MIN	Ambient
96	*Bond Wash Solution	Wash	1 MIN	Ambient
97	RNAscope Multiomic HRP C2	Reagent	1 MIN	42°C
98	RNAscope Multiomic HRP C2	Reagent	15 MIN	42°C
99	*Bond Wash Solution	Wash	0 MIN	Ambient
100	*Bond Wash Solution	Wash	0 MIN	Ambient
101	*Bond Wash Solution	Wash	0 MIN	Ambient
102	*Bond Wash Solution	Wash	1 MIN	Ambient
103	*Bond Wash Solution	Wash	1 MIN	Ambient
104	*Bond Wash Solution	Wash	1 MIN	Ambient
105	*Bond Wash Solution	Wash	1 MIN	Ambient
106	RNAscope Multiomic TSA-F2	Reagent	1 MIN	Ambient
107	RNAscope Multiomic TSA-F2	Reagent	30 MIN	Ambient
108	*Bond Wash Solution	Wash	0 MIN	Ambient
109	*Bond Wash Solution	Wash	0 MIN	Ambient
110	*Bond Wash Solution	Wash	0 MIN	Ambient
111	*Bond Wash Solution	Wash	1 MIN	Ambient
112	*Bond Wash Solution	Wash	1 MIN	Ambient
113	*Bond Wash Solution	Wash	1 MIN	Ambient
114	*Bond Wash Solution	Wash	1 MIN	Ambient
115	RNAscope Multiomic LS HRP Blocker	Reagent	1 MIN	42°C

Step No.	Reagent	Step Type	Incubation Time	Temperature†
116	RNAscope Multiomic LS HRP Blocker	Reagent	15 MIN	42°C
117	*Bond Wash Solution	Wash	0 MIN	Ambient
118	*Bond Wash Solution	Wash	0 MIN	Ambient
119	*Bond Wash Solution	Wash	0 MIN	Ambient
120	*Bond Wash Solution	Wash	1 MIN	Ambient
121	*Bond Wash Solution	Wash	1 MIN	Ambient
122	*Bond Wash Solution	Wash	1 MIN	Ambient
123	*Bond Wash Solution	Wash	1 MIN	Ambient
124	RNAscope Multiomic LS HRP C3	Reagent	1 MIN	42°C
125	RNAscope Multiomic LS HRP C3	Reagent	15 MIN	42°C
126	*Bond Wash Solution	Wash	0 MIN	Ambient
127	*Bond Wash Solution	Wash	0 MIN	Ambient
128	*Bond Wash Solution	Wash	0 MIN	Ambient
129	*Bond Wash Solution	Wash	1 MIN	Ambient
130	*Bond Wash Solution	Wash	1 MIN	Ambient
131	*Bond Wash Solution	Wash	1 MIN	Ambient
132	*Bond Wash Solution	Wash	1 MIN	Ambient
133	RNAscope Multiomic TSA-F3	Reagent	1 MIN	Ambient
134	RNAscope Multiomic TSA-F3	Reagent	30 MIN	Ambient
135	*Bond Wash Solution	Wash	0 MIN	Ambient
136	*Bond Wash Solution	Wash	0 MIN	Ambient
137	*Bond Wash Solution	Wash	0 MIN	Ambient
138	*Bond Wash Solution	Wash	1 MIN	Ambient
139	*Bond Wash Solution	Wash	1 MIN	Ambient
140	*Bond Wash Solution	Wash	1 MIN	Ambient
141	*Bond Wash Solution	Wash	1 MIN	Ambient
142	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
143	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
144	*Bond Wash Solution	Wash	0 MIN	Ambient
145	*Bond Wash Solution	Wash	0 MIN	Ambient
146	*Bond Wash Solution	Wash	0 MIN	Ambient
147	*Bond Wash Solution	Wash	1 MIN	Ambient
148	*Bond Wash Solution	Wash	1 MIN	Ambient
149	*Bond Wash Solution	Wash	1 MIN	Ambient
150	RNAscope Multiomic LS HRP C4	Reagent	1 MIN	42°C
151	RNAscope Multiomic LS HRP C4	Reagent	15 MIN	42°C
152	*Bond Wash Solution	Wash	0 MIN	Ambient
153	*Bond Wash Solution	Wash	0 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
154	*Bond Wash Solution	Wash	0 MIN	Ambient
155	*Bond Wash Solution	Wash	1 MIN	Ambient
156	*Bond Wash Solution	Wash	1 MIN	Ambient
157	*Bond Wash Solution	Wash	1 MIN	Ambient
158	RNAscope Multiomic TSA-F4	Reagent	1 MIN	Ambient
159	RNAscope Multiomic TSA-F4	Reagent	30 MIN	Ambient
160	*Bond Wash Solution	Wash	0 MIN	Ambient
161	*Bond Wash Solution	Wash	0 MIN	Ambient
162	*Bond Wash Solution	Wash	0 MIN	Ambient
163	*Bond Wash Solution	Wash	1 MIN	Ambient
164	*Bond Wash Solution	Wash	1 MIN	Ambient
165	*Bond Wash Solution	Wash	1 MIN	Ambient
166	*Bond Wash Solution	Wash	1 MIN	Ambient
167	RNAscope Multiomic LS HRP Blocker	Reagent	1 MIN	42°C
168	RNAscope Multiomic LS HRP Blocker	Reagent	15 MIN	42°C
169	*Bond Wash Solution	Wash	0 MIN	Ambient
170	*Bond Wash Solution	Wash	0 MIN	Ambient
171	*Bond Wash Solution	Wash	0 MIN	Ambient
172	*Bond Wash Solution	Wash	1 MIN	Ambient
173	*Bond Wash Solution	Wash	1 MIN	Ambient
174	*Bond Wash Solution	Wash	1 MIN	Ambient
175	RNAscope Multiomic LS HRP C5	Reagent	1 MIN	42°C
176	RNAscope Multiomic LS HRP C5	Reagent	15 MIN	42°C
177	*Bond Wash Solution	Wash	0 MIN	Ambient
178	*Bond Wash Solution	Wash	0 MIN	Ambient
179	*Bond Wash Solution	Wash	0 MIN	Ambient
180	*Bond Wash Solution	Wash	1 MIN	Ambient
181	*Bond Wash Solution	Wash	1 MIN	Ambient
182	*Bond Wash Solution	Wash	1 MIN	Ambient
183	RNAscope Multiomic TSA-F5	Reagent	1 MIN	Ambient
184	RNAscope Multiomic TSA-F5	Reagent	30 MIN	Ambient
185	*Bond Wash Solution	Wash	0 MIN	Ambient
186	*Bond Wash Solution	Wash	0 MIN	Ambient
187	*Bond Wash Solution	Wash	0 MIN	Ambient
188	*Bond Wash Solution	Wash	1 MIN	Ambient
189	*Bond Wash Solution	Wash	1 MIN	Ambient
190	*Bond Wash Solution	Wash	1 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
191	*Bond Wash Solution	Wash	1 MIN	Ambient
192	RNAscope Multiomic LS HRP Blocker	Reagent	1 MIN	42°C
193	RNAscope Multiomic LS HRP Blocker	Reagent	15 MIN	42°C
194	*Bond Wash Solution	Wash	0 MIN	Ambient
195	*Bond Wash Solution	Wash	0 MIN	Ambient
196	*Bond Wash Solution	Wash	0 MIN	Ambient
197	*Bond Wash Solution	Wash	1 MIN	Ambient
198	*Bond Wash Solution	Wash	1 MIN	Ambient
199	*Bond Wash Solution	Wash	1 MIN	Ambient
200	RNAscope Multiomic LS HRP C6	Reagent	1 MIN	42°C
201	RNAscope Multiomic LS HRP C6	Reagent	15 MIN	42°C
202	*Bond Wash Solution	Wash	0 MIN	Ambient
203	*Bond Wash Solution	Wash	0 MIN	Ambient
204	*Bond Wash Solution	Wash	0 MIN	Ambient
205	*Bond Wash Solution	Wash	1 MIN	Ambient
206	*Bond Wash Solution	Wash	1 MIN	Ambient
207	*Bond Wash Solution	Wash	1 MIN	Ambient
208	Opal TSA-DIG†	Reagent	1 MIN	Ambient
209	Opal TSA-DIG†	Reagent	30 MIN	Ambient
210	*Bond Wash Solution	Wash	0 MIN	Ambient
211	*Bond Wash Solution	Wash	0 MIN	Ambient
212	*Bond Wash Solution	Wash	0 MIN	Ambient
213	*Bond Wash Solution	Wash	1 MIN	Ambient
214	*Bond Wash Solution	Wash	1 MIN	Ambient
215	RNAscope Multiomic LS HRP Blocker	Reagent	1 MIN	42°C
216	RNAscope Multiomic LS HRP Blocker	Reagent	15 MIN	42°C
217	*Bond Wash Solution	Wash	0 MIN	Ambient
218	*Bond Wash Solution	Wash	0 MIN	Ambient
219	*Bond Wash Solution	Wash	0 MIN	Ambient
220	*Bond Wash Solution	Wash	1 MIN	Ambient
221	*Bond Wash Solution	Wash	1 MIN	Ambient
222	Polaris 780	Reagent	1 MIN	Ambient
223	Polaris 780	Reagent	30 MIN	Ambient
224	*Bond Wash Solution	Wash	0 MIN	Ambient
225	*Bond Wash Solution	Wash	0 MIN	Ambient
226	*Bond Wash Solution	Wash	0 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
227	*Bond Wash Solution	Wash	1 MIN	Ambient
228	*Bond Wash Solution	Wash	1 MIN	Ambient
229	RNAscope Multiomic LS DAPI	Reagent	10 MIN	Ambient
230	*Bond Wash Solution	Wash	0 MIN	Ambient
231	*Bond Wash Solution	Wash	0 MIN	Ambient
232	*Bond Wash Solution	Wash	0 MIN	Ambient

* Indicates reagent is hard coded in the software by Leica Biosystems.

† If Opal 780 is used

‡ The standard protocol uses DAPI. Use BOND Wash instead of DAPI, if you are using DAPI offline or performing IHC steps afterwards on your samples.

Appendix B. mRNA only 6-plex Protocol (using the Mock Probe workaround)

Heated bond washes 3-5 come from the bulk reagents and are heated by the instrument. You cannot delete these steps. You may delete other wash steps.

IMPORTANT! The BOND RX can only execute this workflow on one tray for the RNAscope Multiomic assay (six channels) due to its length, follow the protocol steps listed in the following table. Adding any additional steps will prevent the system from running even one the tray.

Step No.	Reagent	Step Type	Incubation Time	Temperature†
1	Probe 1	Reagent	0 MIN	Ambient
2	Probe 1	Reagent	120 MIN	42°C
3	*Bond Wash Solution	Wash	0 MIN	42°C
4	*Bond Wash Solution	Wash	1 MIN	42°C
5	*Bond Wash Solution	Wash	5 MIN	42°C
6	*Bond Wash Solution	Wash	0 MIN	Ambient
7	*Bond Wash Solution	Wash	0 MIN	Ambient
8	*Bond Wash Solution	Wash	0 MIN	Ambient
9	*Bond Wash Solution	Wash	0 MIN	Ambient
10	*Bond Wash Solution	Wash	0 MIN	Ambient
11	*Bond Wash Solution	Wash	1 MIN	Ambient
12	*Bond Wash Solution	Wash	1 MIN	Ambient
13	*Bond Wash Solution	Wash	0 MIN	Ambient
14	RNAscope Multiomic LS Amp 1	Reagent	1 MIN	42°C
15	RNAscope Multiomic LS Amp 1	Reagent	30 MIN	42°C
16	*Bond Wash Solution	Wash	0 MIN	Ambient
17	*Bond Wash Solution	Wash	0 MIN	Ambient
18	*Bond Wash Solution	Wash	0 MIN	Ambient
19	*Bond Wash Solution	Wash	3 MIN	Ambient
20	*Bond Wash Solution	Wash	3 MIN	Ambient
21	*Bond Wash Solution	Wash	0 MIN	Ambient
22	*Bond Wash Solution	Wash	0 MIN	Ambient
23	*Bond Wash Solution	Wash	0 MIN	Ambient
24	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
25	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
26	*Bond Wash Solution	Wash	0 MIN	Ambient
27	*Bond Wash Solution	Wash	0 MIN	Ambient
28	*Bond Wash Solution	Open Wash	0 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
29	*Bond Wash Solution	Wash	0 MIN	Ambient
30	RNAscope Multiomic LS Amp 2	Reagent	1 MIN	42°C
31	RNAscope Multiomic LS Amp 2	Reagent	30 MIN	42°C
32	*Bond Wash Solution	Wash	0 MIN	Ambient
33	*Bond Wash Solution	Wash	0 MIN	Ambient
34	*Bond Wash Solution	Wash	0 MIN	Ambient
35	*Bond Wash Solution	Wash	3 MIN	Ambient
36	*Bond Wash Solution	Wash	3 MIN	Ambient
37	*Bond Wash Solution	Wash	0 MIN	Ambient
38	*Bond Wash Solution	Wash	0 MIN	Ambient
39	*Bond Wash Solution	Wash	0 MIN	Ambient
40	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
41	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
42	*Bond Wash Solution	Wash	0 MIN	Ambient
43	*Bond Wash Solution	Wash	1 MIN	Ambient
44	*Bond Wash Solution	Open Wash	1 MIN	Ambient
45	*Bond Wash Solution	Wash	1 MIN	Ambient
46	RNAscope Multiomic LS Amp 3	Reagent	1 MIN	42°C
47	RNAscope Multiomic LS Amp 3	Reagent	15 MIN	42°C
48	*Bond Wash Solution	Wash	0 MIN	Ambient
49	*Bond Wash Solution	Wash	0 MIN	Ambient
50	*Bond Wash Solution	Wash	0 MIN	Ambient
51	*Bond Wash Solution	Wash	1 MIN	Ambient
52	*Bond Wash Solution	Wash	1 MIN	Ambient
53	*Bond Wash Solution	Wash	1 MIN	Ambient
54	*Bond Wash Solution	Open Wash	1 MIN	Ambient
55	*Bond Wash Solution	Wash	1 MIN	Ambient
56	RNAscope Multiomic LS HRP C1	Reagent	1 MIN	42°C
57	RNAscope Multiomic LS HRP C1	Reagent	15 MIN	42°C
58	*Bond Wash Solution	Wash	0 MIN	Ambient
59	*Bond Wash Solution	Wash	0 MIN	Ambient
60	*Bond Wash Solution	Wash	0 MIN	Ambient
61	*Bond Wash Solution	Wash	1 MIN	Ambient
62	*Bond Wash Solution	Wash	1 MIN	Ambient
63	*Bond Wash Solution	Wash	1 MIN	Ambient
64	*Bond Wash Solution	Wash	1 MIN	Ambient
65	*Bond Wash Solution	Wash	1 MIN	Ambient
66	RNAscope Multiomic TSA-F1	Reagent	1 MIN	Ambient
67	RNAscope Multiomic TSA-F1	Reagent	30 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
68	*Bond Wash Solution	Wash	0 MIN	Ambient
69	*Bond Wash Solution	Wash	0 MIN	Ambient
70	*Bond Wash Solution	Wash	0 MIN	Ambient
71	*Bond Wash Solution	Wash	1 MIN	Ambient
72	*Bond Wash Solution	Wash	1 MIN	Ambient
73	*Bond Wash Solution	Wash	1 MIN	Ambient
74	*Bond Wash Solution	Wash	1 MIN	Ambient
75	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
76	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
77	*Bond Wash Solution	Wash	0 MIN	Ambient
78	*Bond Wash Solution	Wash	0 MIN	Ambient
79	*Bond Wash Solution	Wash	0 MIN	Ambient
80	*Bond Wash Solution	Wash	1 MIN	Ambient
81	*Bond Wash Solution	Wash	1 MIN	Ambient
82	*Bond Wash Solution	Wash	1 MIN	Ambient
83	*Bond Wash Solution	Wash	1 MIN	Ambient
84	RNAscope Multiomic LS HRP C2	Reagent	1 MIN	42°C
85	RNAscope Multiomic LS HRP C2	Reagent	15 MIN	42°C
86	*Bond Wash Solution	Wash	0 MIN	Ambient
87	*Bond Wash Solution	Wash	0 MIN	Ambient
88	*Bond Wash Solution	Wash	0 MIN	Ambient
89	*Bond Wash Solution	Wash	1 MIN	Ambient
90	*Bond Wash Solution	Wash	1 MIN	Ambient
91	*Bond Wash Solution	Wash	1 MIN	Ambient
92	*Bond Wash Solution	Wash	1 MIN	Ambient
93	RNAscope Multiomic TSA-F2	Reagent	1 MIN	Ambient
94	RNAscope Multiomic TSA-F2	Reagent	30 MIN	Ambient
95	*Bond Wash Solution	Wash	0 MIN	Ambient
96	*Bond Wash Solution	Wash	0 MIN	Ambient
97	*Bond Wash Solution	Wash	0 MIN	Ambient
98	*Bond Wash Solution	Wash	1 MIN	Ambient
99	*Bond Wash Solution	Wash	1 MIN	Ambient
100	*Bond Wash Solution	Wash	1 MIN	Ambient
101	*Bond Wash Solution	Wash	1 MIN	Ambient
102	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
103	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
104	*Bond Wash Solution	Wash	0 MIN	Ambient
105	*Bond Wash Solution	Wash	0 MIN	Ambient
106	*Bond Wash Solution	Wash	0 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
107	*Bond Wash Solution	Wash	1 MIN	Ambient
108	*Bond Wash Solution	Wash	1 MIN	Ambient
109	*Bond Wash Solution	Wash	1 MIN	Ambient
110	*Bond Wash Solution	Wash	1 MIN	Ambient
111	RNAscope Multiomic LS HRP C3	Reagent	1 MIN	42°C
112	RNAscope Multiomic LS HRP C3	Reagent	15 MIN	42°C
113	*Bond Wash Solution	Wash	0 MIN	Ambient
114	*Bond Wash Solution	Wash	0 MIN	Ambient
115	*Bond Wash Solution	Wash	0 MIN	Ambient
116	*Bond Wash Solution	Wash	1 MIN	Ambient
117	*Bond Wash Solution	Wash	1 MIN	Ambient
118	*Bond Wash Solution	Wash	1 MIN	Ambient
119	*Bond Wash Solution	Wash	1 MIN	Ambient
120	RNAscope Multiomic TSA-F3	Reagent	1 MIN	Ambient
121	RNAscope Multiomic TSA-F3	Reagent	30 MIN	Ambient
122	*Bond Wash Solution	Wash	0 MIN	Ambient
123	*Bond Wash Solution	Wash	0 MIN	Ambient
124	*Bond Wash Solution	Wash	0 MIN	Ambient
125	*Bond Wash Solution	Wash	1 MIN	Ambient
126	*Bond Wash Solution	Wash	1 MIN	Ambient
127	*Bond Wash Solution	Wash	1 MIN	Ambient
128	*Bond Wash Solution	Wash	1 MIN	Ambient
129	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
130	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
131	*Bond Wash Solution	Wash	0 MIN	Ambient
132	*Bond Wash Solution	Wash	0 MIN	Ambient
133	*Bond Wash Solution	Wash	0 MIN	Ambient
134	*Bond Wash Solution	Wash	1 MIN	Ambient
135	*Bond Wash Solution	Wash	1 MIN	Ambient
136	*Bond Wash Solution	Wash	1 MIN	Ambient
137	RNAscope Multiomic LS HRP C4	Reagent	1 MIN	42°C
138	RNAscope Multiomic LS HRP C4	Reagent	15 MIN	42°C
139	*Bond Wash Solution	Wash	0 MIN	Ambient
140	*Bond Wash Solution	Wash	0 MIN	Ambient
141	*Bond Wash Solution	Wash	0 MIN	Ambient
142	*Bond Wash Solution	Wash	1 MIN	Ambient
143	*Bond Wash Solution	Wash	1 MIN	Ambient
144	*Bond Wash Solution	Wash	1 MIN	Ambient
145	RNAscope Multiomic TSA-F4	Reagent	1 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
146	RNAscope Multiomic TSA-F4	Reagent	30 MIN	Ambient
147	*Bond Wash Solution	Wash	0 MIN	Ambient
148	*Bond Wash Solution	Wash	0 MIN	Ambient
149	*Bond Wash Solution	Wash	0 MIN	Ambient
150	*Bond Wash Solution	Wash	1 MIN	Ambient
151	*Bond Wash Solution	Wash	1 MIN	Ambient
152	*Bond Wash Solution	Wash	1 MIN	Ambient
153	*Bond Wash Solution	Wash	1 MIN	Ambient
154	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
155	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
156	*Bond Wash Solution	Wash	0 MIN	Ambient
157	*Bond Wash Solution	Wash	0 MIN	Ambient
158	*Bond Wash Solution	Wash	0 MIN	Ambient
159	*Bond Wash Solution	Wash	1 MIN	Ambient
160	*Bond Wash Solution	Wash	1 MIN	Ambient
161	*Bond Wash Solution	Wash	1 MIN	Ambient
162	RNAscope Multiomic LS HRP C5	Reagent	1 MIN	42°C
163	RNAscope Multiomic LS HRP C5	Reagent	15 MIN	42°C
164	*Bond Wash Solution	Wash	0 MIN	Ambient
165	*Bond Wash Solution	Wash	0 MIN	Ambient
166	*Bond Wash Solution	Wash	0 MIN	Ambient
167	*Bond Wash Solution	Wash	1 MIN	Ambient
168	*Bond Wash Solution	Wash	1 MIN	Ambient
169	*Bond Wash Solution	Wash	1 MIN	Ambient
170	RNAscope Multiomic TSA-F5	Reagent	1 MIN	Ambient
171	RNAscope Multiomic TSA-F5	Reagent	30 MIN	Ambient
172	*Bond Wash Solution	Wash	0 MIN	Ambient
173	*Bond Wash Solution	Wash	0 MIN	Ambient
174	*Bond Wash Solution	Wash	0 MIN	Ambient
175	*Bond Wash Solution	Wash	1 MIN	Ambient
176	*Bond Wash Solution	Wash	1 MIN	Ambient
177	*Bond Wash Solution	Wash	1 MIN	Ambient
178	*Bond Wash Solution	Wash	1 MIN	Ambient
179	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
180	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
181	*Bond Wash Solution	Wash	0 MIN	Ambient
182	*Bond Wash Solution	Wash	0 MIN	Ambient
183	*Bond Wash Solution	Wash	0 MIN	Ambient
184	*Bond Wash Solution	Wash	1 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
185	*Bond Wash Solution	Wash	1 MIN	Ambient
186	*Bond Wash Solution	Wash	1 MIN	Ambient
187	RNAscope Multiomic LS HRP C6	Reagent	1 MIN	42°C
188	RNAscope Multiomic LS HRP C6	Reagent	15 MIN	42°C
189	*Bond Wash Solution	Wash	0 MIN	Ambient
190	*Bond Wash Solution	Wash	0 MIN	Ambient
191	*Bond Wash Solution	Wash	0 MIN	Ambient
192	*Bond Wash Solution	Wash	1 MIN	Ambient
193	*Bond Wash Solution	Wash	1 MIN	Ambient
194	*Bond Wash Solution	Wash	1 MIN	Ambient
195	TSA-DIG	Reagent	1 MIN	Ambient
196	TSA-DIG	Reagent	30 MIN	Ambient
197	*Bond Wash Solution	Wash	0 MIN	Ambient
198	*Bond Wash Solution	Wash	0 MIN	Ambient
199	*Bond Wash Solution	Wash	0 MIN	Ambient
200	*Bond Wash Solution	Wash	1 MIN	Ambient
201	*Bond Wash Solution	Wash	1 MIN	Ambient
202	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
203	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
204	*Bond Wash Solution	Wash	0 MIN	Ambient
205	*Bond Wash Solution	Wash	0 MIN	Ambient
206	*Bond Wash Solution	Wash	0 MIN	Ambient
207	*Bond Wash Solution	Wash	1 MIN	Ambient
208	*Bond Wash Solution	Wash	1 MIN	Ambient
209	Polaris 780	Reagent	1 MIN	Ambient
210	Polaris 780	Reagent	30 MIN	Ambient
211	*Bond Wash Solution	Wash	0 MIN	Ambient
212	*Bond Wash Solution	Wash	0 MIN	Ambient
213	*Bond Wash Solution	Wash	0 MIN	Ambient
214	*Bond Wash Solution	Wash	1 MIN	Ambient
215	*Bond Wash Solution	Wash	1 MIN	Ambient
216	RNAscope Multiomic LS DAPI	Reagent	10 MIN	Ambient
217	*Bond Wash Solution	Wash	0 MIN	Ambient
218	*Bond Wash Solution	Wash	0 MIN	Ambient
219	*Bond Wash Solution	Wash	0 MIN	Ambient

* Indicates reagent is hard coded in the software by Leica Biosystems.

† The standard protocol uses DAPI. Use BOND Wash instead of DAPI, if you are using DAPI offline or performing IHC steps afterwards on your samples.



Appendix C. mRNA and conjugated primary antibody detection

Reagent	Container name	Details (concentration, dilution)
Salmon Sperm DNA	Open1	500ug/mL, in multiomic antibody diluent
10% NBF	10% NBF	None
RNA pooled Probe	Open 2	1:50, in RNAscope probe diluent
Antibody mix	CoDetection Antibody 1	Multiomic antibody diluent (See Appendix H for concentration)

Slide setup:

Slide properties

Pos 1

Tissue type: Dispense volume:

Test tissue 100 µL
 Negative tissue 150 µL
 Positive tissue

Staining mode:

Single Routine

Process: IHC ISH

Marker: Mock Probe

Protocols

Staining:	mRNA + Primary Conjugates
Preparation:	*Bake and Dewax
HIER:	*ACD HIER 15 min with ER2 (95)
Enzyme:	*----
Probe Application:	*RNAscope 2.5 LSx Probe Application
Denaturation:	*----
Hybridization:	ACD 1 Min Hybridization
Probe Removal:	*RNAscope 2.5 LSx Probe Removal

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Step No.	Reagent	Step Type	Incubation Time	Temperature†
1	Open1	Reagent	0 MIN	Ambient
2	Open1	Reagent	60 MIN	Ambient
3	*Bond Wash Solution	Wash	0 MIN	Ambient
4	*Bond Wash Solution	Wash	0 MIN	Ambient
5	*Bond Wash Solution	Wash	0 MIN	Ambient
6	*Bond Wash Solution	Wash	0 MIN	Ambient
7	Co-Detection Antibody 1	Reagent	0 MIN	Ambient
8	Co-Detection Antibody 1	Reagent	60 MIN	Ambient
9	*Bond Wash Solution	Wash	0 MIN	Ambient
10	*Bond Wash Solution	Wash	0 MIN	Ambient
11	*Bond Wash Solution	Wash	1 MIN	Ambient
12	*Bond Wash Solution	Wash	3MIN	Ambient
13	*Bond Wash Solution	Wash	3MIN	Ambient
14	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
15	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
16	*Bond Wash Solution	Wash	0 MIN	Ambient
17	*Bond Wash Solution	Wash	0 MIN	Ambient
18	*Bond Wash Solution	Open Wash	0 MIN	Ambient
19	*Bond Wash Solution	Wash	0 MIN	Ambient
20	10% NBF	Reagent	30 MIN	Ambient
21	*Bond Wash Solution	Wash	0 MIN	Ambient
22	*Bond Wash Solution	Wash	0 MIN	Ambient
23	*Bond Wash Solution	Wash	0 MIN	Ambient
24	*Bond Wash Solution	Wash	3 MIN	Ambient
25	*Bond Wash Solution	Wash	3 MIN	Ambient
26	*Bond Wash Solution	Wash	0 MIN	Ambient
27	*Bond Wash Solution	Wash	0 MIN	Ambient
28	*Bond Wash Solution	Wash	0 MIN	Ambient
29	PretreatPro	Reagent	0 MIN	Ambient
30	PretreatPro	Reagent	30 MIN	40°C
31	*Bond Wash Solution	Wash	0 MIN	Ambient
32	*Bond Wash Solution	Wash	0 MIN	Ambient
33	*Bond Wash Solution	Wash	0 MIN	Ambient
34	*Bond Wash Solution	Wash	1 MIN	Ambient
35	*Bond Wash Solution	Wash	1 MIN	Ambient
36	Hydrogen Peroxide	Reagent	10 MIN	Ambient
37	*Bond Wash Solution	Wash	0 MIN	Ambient
38	*Bond Wash Solution	Wash	0 MIN	Ambient
39	*Bond Wash Solution	Wash	0 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
40	*Bond Wash Solution	Wash	0 MIN	Ambient
41	*Bond Wash Solution	Wash	0 MIN	Ambient
42	Open 2	Reagent	0 MIN	Ambient
43	Open 2	Reagent	120 MIN	42°C
44	*Bond Wash Solution	Wash	0 MIN	42°C
45	*Bond Wash Solution	Wash	1 MIN	42°C
46	*Bond Wash Solution	Wash	5 MIN	42°C
47	*Bond Wash Solution	Wash	0 MIN	Ambient
48	*Bond Wash Solution	Wash	0 MIN	Ambient
49	*Bond Wash Solution	Wash	0 MIN	Ambient
50	*Bond Wash Solution	Wash	0 MIN	Ambient
51	*Bond Wash Solution	Wash	0 MIN	Ambient
52	*Bond Wash Solution	Wash	1 MIN	Ambient
53	*Bond Wash Solution	Wash	1 MIN	Ambient
54	*Bond Wash Solution	Wash	0 MIN	Ambient
55	RNAscope Multiomic Amp 1	Reagent	1 MIN	42°C
56	RNAscope Multiomic Amp 1	Reagent	30 MIN	42°C
57	*Bond Wash Solution	Wash	0 MIN	Ambient
58	*Bond Wash Solution	Wash	0 MIN	Ambient
59	*Bond Wash Solution	Wash	0 MIN	Ambient
60	*Bond Wash Solution	Wash	3 MIN	Ambient
61	*Bond Wash Solution	Wash	3 MIN	Ambient
62	*Bond Wash Solution	Wash	0 MIN	Ambient
63	*Bond Wash Solution	Wash	0 MIN	Ambient
64	*Bond Wash Solution	Wash	0 MIN	Ambient
65	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
66	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
67	*Bond Wash Solution	Wash	0 MIN	Ambient
68	*Bond Wash Solution	Wash	0 MIN	Ambient
69	*Bond Wash Solution	Open Wash	0 MIN	Ambient
70	*Bond Wash Solution	Wash	0 MIN	Ambient
71	RNAscope Multiomic Amp 2	Reagent	1 MIN	42°C
72	RNAscope Multiomic Amp 2	Reagent	30 MIN	42°C
73	*Bond Wash Solution	Wash	0 MIN	Ambient
74	*Bond Wash Solution	Wash	0 MIN	Ambient
75	*Bond Wash Solution	Wash	0 MIN	Ambient
76	*Bond Wash Solution	Wash	3 MIN	Ambient
77	*Bond Wash Solution	Wash	3 MIN	Ambient
78	*Bond Wash Solution	Wash	0 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
79	*Bond Wash Solution	Wash	0 MIN	Ambient
80	*Bond Wash Solution	Wash	0 MIN	Ambient
81	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
82	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
83	*Bond Wash Solution	Wash	0 MIN	Ambient
84	*Bond Wash Solution	Wash	1 MIN	Ambient
85	*Bond Wash Solution	Open Wash	1 MIN	Ambient
86	*Bond Wash Solution	Wash	1 MIN	Ambient
87	RNAscope Multiomic Amp 3	Reagent	1 MIN	42°C
88	RNAscope Multiomic Amp 3	Reagent	15 MIN	42°C
89	*Bond Wash Solution	Wash	0 MIN	Ambient
90	*Bond Wash Solution	Wash	0 MIN	Ambient
91	*Bond Wash Solution	Wash	0 MIN	Ambient
92	*Bond Wash Solution	Wash	1 MIN	Ambient
93	*Bond Wash Solution	Wash	1 MIN	Ambient
94	*Bond Wash Solution	Wash	1 MIN	Ambient
95	*Bond Wash Solution	Open Wash	1 MIN	Ambient
96	*Bond Wash Solution	Wash	1 MIN	Ambient
97	RNAscope Multiomic HRP-C1	Reagent	1 MIN	42°C
98	RNAscope Multiomic HRP-C1	Reagent	15 MIN	42°C
99	*Bond Wash Solution	Wash	0 MIN	Ambient
100	*Bond Wash Solution	Wash	0 MIN	Ambient
101	*Bond Wash Solution	Wash	0 MIN	Ambient
102	*Bond Wash Solution	Wash	1 MIN	Ambient
103	*Bond Wash Solution	Wash	1 MIN	Ambient
104	*Bond Wash Solution	Wash	1 MIN	Ambient
105	*Bond Wash Solution	Wash	1 MIN	Ambient
106	*Bond Wash Solution	Wash	1 MIN	Ambient
107	RNAscope Multiomic TSA-F1	Reagent	1 MIN	Ambient
108	RNAscope Multiomic TSA-F1	Reagent	30 MIN	Ambient
109	*Bond Wash Solution	Wash	0 MIN	Ambient
110	*Bond Wash Solution	Wash	0 MIN	Ambient
111	*Bond Wash Solution	Wash	0 MIN	Ambient
112	*Bond Wash Solution	Wash	1 MIN	Ambient
113	*Bond Wash Solution	Wash	1 MIN	Ambient
114	*Bond Wash Solution	Wash	1 MIN	Ambient
115	*Bond Wash Solution	Wash	1 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
116	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
117	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
118	*Bond Wash Solution	Wash	0 MIN	Ambient
119	*Bond Wash Solution	Wash	0 MIN	Ambient
120	*Bond Wash Solution	Wash	0 MIN	Ambient
121	*Bond Wash Solution	Wash	1 MIN	Ambient
122	*Bond Wash Solution	Wash	1 MIN	Ambient
123	*Bond Wash Solution	Wash	1 MIN	Ambient
124	*Bond Wash Solution	Wash	1 MIN	Ambient
125	RNAscope Multiomic LS HRP C2	Reagent	1 MIN	42°C
126	RNAscope Multiomic LS HRP C2	Reagent	15 MIN	42°C
127	*Bond Wash Solution	Wash	0 MIN	Ambient
128	*Bond Wash Solution	Wash	0 MIN	Ambient
129	*Bond Wash Solution	Wash	0 MIN	Ambient
130	*Bond Wash Solution	Wash	1 MIN	Ambient
131	*Bond Wash Solution	Wash	1 MIN	Ambient
132	*Bond Wash Solution	Wash	1 MIN	Ambient
133	*Bond Wash Solution	Wash	1 MIN	Ambient
134	RNAscope Multiomic TSA-F2	Reagent	1 MIN	Ambient
135	RNAscope Multiomic TSA-F2	Reagent	30 MIN	Ambient
136	*Bond Wash Solution	Wash	0 MIN	Ambient
137	*Bond Wash Solution	Wash	0 MIN	Ambient
138	*Bond Wash Solution	Wash	0 MIN	Ambient
139	*Bond Wash Solution	Wash	1 MIN	Ambient
140	*Bond Wash Solution	Wash	1 MIN	Ambient
141	*Bond Wash Solution	Wash	1 MIN	Ambient
142	*Bond Wash Solution	Wash	1 MIN	Ambient
143	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
144	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
145	*Bond Wash Solution	Wash	0 MIN	Ambient
146	*Bond Wash Solution	Wash	0 MIN	Ambient
147	*Bond Wash Solution	Wash	0 MIN	Ambient
148	*Bond Wash Solution	Wash	1 MIN	Ambient
149	*Bond Wash Solution	Wash	1 MIN	Ambient
150	*Bond Wash Solution	Wash	1 MIN	Ambient
151	*Bond Wash Solution	Wash	1 MIN	Ambient
152	RNAscope Multiomic LS HRP C3	Reagent	1 MIN	42°C
153	RNAscope Multiomic LS HRP C3	Reagent	15 MIN	42°C
154	*Bond Wash Solution	Wash	0 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
155	*Bond Wash Solution	Wash	0 MIN	Ambient
156	*Bond Wash Solution	Wash	0 MIN	Ambient
157	*Bond Wash Solution	Wash	1 MIN	Ambient
158	*Bond Wash Solution	Wash	1 MIN	Ambient
159	*Bond Wash Solution	Wash	1 MIN	Ambient
160	*Bond Wash Solution	Wash	1 MIN	Ambient
161	RNAscope Multiomic TSA-F3	Reagent	1 MIN	Ambient
162	RNAscope Multiomic TSA-F3	Reagent	30 MIN	Ambient
163	*Bond Wash Solution	Wash	0 MIN	Ambient
164	*Bond Wash Solution	Wash	0 MIN	Ambient
165	*Bond Wash Solution	Wash	0 MIN	Ambient
166	*Bond Wash Solution	Wash	1 MIN	Ambient
167	*Bond Wash Solution	Wash	1 MIN	Ambient
168	*Bond Wash Solution	Wash	1 MIN	Ambient
169	*Bond Wash Solution	Wash	1 MIN	Ambient
170	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
171	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
172	*Bond Wash Solution	Wash	0 MIN	Ambient
173	*Bond Wash Solution	Wash	0 MIN	Ambient
174	*Bond Wash Solution	Wash	0 MIN	Ambient
175	*Bond Wash Solution	Wash	1 MIN	Ambient
176	*Bond Wash Solution	Wash	1 MIN	Ambient
177	*Bond Wash Solution	Wash	1 MIN	Ambient
178	RNAscope Multiomic LS HRP C4	Reagent	1 MIN	42°C
179	RNAscope Multiomic LS HRP C4	Reagent	15 MIN	42°C
180	*Bond Wash Solution	Wash	0 MIN	Ambient
181	*Bond Wash Solution	Wash	0 MIN	Ambient
182	*Bond Wash Solution	Wash	0 MIN	Ambient
183	*Bond Wash Solution	Wash	1 MIN	Ambient
184	*Bond Wash Solution	Wash	1 MIN	Ambient
185	*Bond Wash Solution	Wash	1 MIN	Ambient
186	RNAscope Multiomic TSA-F4	Reagent	1 MIN	Ambient
187	RNAscope Multiomic TSA-F4	Reagent	30 MIN	Ambient
188	*Bond Wash Solution	Wash	0 MIN	Ambient
189	*Bond Wash Solution	Wash	0 MIN	Ambient
190	*Bond Wash Solution	Wash	0 MIN	Ambient
191	*Bond Wash Solution	Wash	1 MIN	Ambient
192	*Bond Wash Solution	Wash	1 MIN	Ambient
193	*Bond Wash Solution	Wash	1 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
194	*Bond Wash Solution	Wash	1 MIN	Ambient
195	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
196	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
197	*Bond Wash Solution	Wash	0 MIN	Ambient
198	*Bond Wash Solution	Wash	0 MIN	Ambient
199	*Bond Wash Solution	Wash	0 MIN	Ambient
200	*Bond Wash Solution	Wash	1 MIN	Ambient
201	*Bond Wash Solution	Wash	1 MIN	Ambient
202	*Bond Wash Solution	Wash	1 MIN	Ambient
203	RNAscope Multiomic LS HRP C5	Reagent	1 MIN	42°C
204	RNAscope Multiomic LS HRP C5	Reagent	15 MIN	42°C
205	*Bond Wash Solution	Wash	0 MIN	Ambient
206	*Bond Wash Solution	Wash	0 MIN	Ambient
207	*Bond Wash Solution	Wash	0 MIN	Ambient
208	*Bond Wash Solution	Wash	1 MIN	Ambient
209	*Bond Wash Solution	Wash	1 MIN	Ambient
210	*Bond Wash Solution	Wash	1 MIN	Ambient
211	RNAscope Multiomic TSA-F5	Reagent	1 MIN	Ambient
212	RNAscope Multiomic TSA-F5	Reagent	30 MIN	Ambient
213	*Bond Wash Solution	Wash	0 MIN	Ambient
214	*Bond Wash Solution	Wash	0 MIN	Ambient
215	*Bond Wash Solution	Wash	0 MIN	Ambient
216	*Bond Wash Solution	Wash	1 MIN	Ambient
217	*Bond Wash Solution	Wash	1 MIN	Ambient
218	*Bond Wash Solution	Wash	1 MIN	Ambient
219	*Bond Wash Solution	Wash	1 MIN	Ambient
220	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
221	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
222	*Bond Wash Solution	Wash	0 MIN	Ambient
223	*Bond Wash Solution	Wash	0 MIN	Ambient
224	*Bond Wash Solution	Wash	0 MIN	Ambient
225	*Bond Wash Solution	Wash	1 MIN	Ambient
226	*Bond Wash Solution	Wash	1 MIN	Ambient
227	*Bond Wash Solution	Wash	1 MIN	Ambient
228	RNAscope Multiomic LS HRP C6	Reagent	1 MIN	42°C
229	RNAscope Multiomic LS HRP C6	Reagent	15 MIN	42°C
230	*Bond Wash Solution	Wash	0 MIN	Ambient
231	*Bond Wash Solution	Wash	0 MIN	Ambient
232	*Bond Wash Solution	Wash	0 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
233	*Bond Wash Solution	Wash	1 MIN	Ambient
234	*Bond Wash Solution	Wash	1 MIN	Ambient
235	*Bond Wash Solution	Wash	1 MIN	Ambient
236	TSA-DIG	Reagent	1 MIN	Ambient
237	TSA-DIG	Reagent	30 MIN	Ambient
238	*Bond Wash Solution	Wash	0 MIN	Ambient
239	*Bond Wash Solution	Wash	0 MIN	Ambient
240	*Bond Wash Solution	Wash	0 MIN	Ambient
241	*Bond Wash Solution	Wash	1 MIN	Ambient
242	*Bond Wash Solution	Wash	1 MIN	Ambient
243	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
244	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
245	*Bond Wash Solution	Wash	0 MIN	Ambient
246	*Bond Wash Solution	Wash	0 MIN	Ambient
247	*Bond Wash Solution	Wash	0 MIN	Ambient
248	*Bond Wash Solution	Wash	1 MIN	Ambient
249	*Bond Wash Solution	Wash	1 MIN	Ambient
250	Polaris 780	Reagent	1 MIN	Ambient
251	Polaris 780	Reagent	30 MIN	Ambient
252	*Bond Wash Solution	Wash	0 MIN	Ambient
253	*Bond Wash Solution	Wash	0 MIN	Ambient
254	*Bond Wash Solution	Wash	0 MIN	Ambient
255	*Bond Wash Solution	Wash	1 MIN	Ambient
256	*Bond Wash Solution	Wash	1 MIN	Ambient
257	DAPI	Reagent	10 MIN	Ambient
258	*Bond Wash Solution	Wash	0 MIN	Ambient
259	*Bond Wash Solution	Wash	0 MIN	Ambient
260	*Bond Wash Solution	Wash	0 MIN	Ambient

D

Appendix D. mRNA + secondary antibody + Primary conjugated antibodies

	Reagent	Container name	Details (concentration, dilution)
1	Salmon Sperm DNA	Open1	500ug/ml, in multiomic antibody diluent
2	10% NBF	10% NBF	None
3	RNA Probe	Open 2	1:50, in RNAscope probe diluent
4	Primary Raw Antibody mix	CoDetection Antibody 1	Multiomic antibody diluent (user defined)
5	Secondary Conjugated Antibody mix	CoDetection Antibody 2	Multiomic antibody diluent (See Appendix H for concentration)
6	Primary Conjugated Antibody mix	CoDetection Antibody 3	Multiomic antibody diluent (See Appendix H for concentration)
7	Antibody Blocker (mouse and/or Rabbit IgG)	Antibody Blocker	5ug/ml, in multiomic antibody diluent

IMPORTANT: When using the RNAscope conjugated secondary antibodies, they may bind to the RNAscope primary conjugated antibodies. To avoid cross-detection, addition of antibody blocker, 5ug/ml of each mouse and rabbit IgG (Mouse IgG2A Isotype Control (R&D Systems MAB003) and Normal Rabbit IgG Control (R&D Systems MAB1050) in multiomic antibody diluent for 30min at RT between the incubation with secondary conjugated antibody and incubation with conjugated primary antibody is required. This will help eliminate this cross-reactivity. The IgG blocker may be added in the “Antibody Blocker” in steps 19 and 20 below.

Slide setup:

Slide properties

Pos 1

Tissue type:	Dispense volume:
<input checked="" type="radio"/> Test tissue	<input type="radio"/> 100 µL
<input type="radio"/> Negative tissue	<input checked="" type="radio"/> 150 µL
<input type="radio"/> Positive tissue	
Staining mode:	
Single	Routine
Single	
Process:	<input type="radio"/> IHC <input checked="" type="radio"/> ISH
Marker:	Mock Probe
Protocols	
Staining:	Primary Antibody + mRNA + Secondary Antibody
Preparation:	*Bake and Dewax
HIER:	*ACD HIER 15 min with ER2 (95)
Enzyme:	*---
Probe Application:	*RNAscope 2.5 LSx Probe Application
Denaturation:	*---
Hybridization:	ACD 1 Min Hybridization
Probe Removal:	*RNAscope 2.5 LSx Probe Removal

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Step No.	Reagent	Step Type	Incubation Time	Temperature†
1	Open1	Reagent	0 MIN	Ambient
2	Open1	Reagent	60 MIN	Ambient
3	*Bond Wash Solution	Wash	0 MIN	Ambient
4	*Bond Wash Solution	Wash	0 MIN	Ambient
5	*Bond Wash Solution	Wash	0 MIN	Ambient
6	*Bond Wash Solution	Wash	0 MIN	Ambient
7	Co-Detection Antibody 1	Reagent	0 MIN	Ambient
8	Co-Detection Antibody 1	Reagent	60 MIN	Ambient
9	*Bond Wash Solution	Wash	0 MIN	Ambient
10	*Bond Wash Solution	Wash	0 MIN	Ambient
11	*Bond Wash Solution	Wash	1 MIN	Ambient
12	*Bond Wash Solution	Wash	1 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
13	Co-Detection Antibody 2	Reagent	0 MIN	Ambient
14	Co-Detection Antibody 2	Reagent	30 MIN	Ambient
15	*Bond Wash Solution	Wash	0 MIN	Ambient
16	*Bond Wash Solution	Wash	0 MIN	Ambient
17	*Bond Wash Solution	Wash	1 MIN	Ambient
18	*Bond Wash Solution	Wash	1MIN	Ambient
19	Antibody Blocker	Reagent	0 MIN	Ambient
20	Antibody Blocker	Reagent	30 MIN	Ambient
21	*Bond Wash Solution	Wash	0 MIN	Ambient
22	*Bond Wash Solution	Wash	0 MIN	Ambient
23	*Bond Wash Solution	Wash	1 MIN	Ambient
24	*Bond Wash Solution	Wash	3MIN	Ambient
25	*Bond Wash Solution	Wash	3MIN	Ambient
26	Co-Detection Antibody 3	Reagent	0 MIN	Ambient
27	Co-Detection Antibody 3	Reagent	60 MIN	Ambient
28	*Bond Wash Solution	Wash	0 MIN	Ambient
29	*Bond Wash Solution	Wash	0 MIN	Ambient
30	*Bond Wash Solution	Wash	1 MIN	Ambient
31	*Bond Wash Solution	Wash	3MIN	Ambient
32	*Bond Wash Solution	Wash	3MIN	Ambient
33	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
34	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
35	*Bond Wash Solution	Wash	0 MIN	Ambient
36	*Bond Wash Solution	Wash	0 MIN	Ambient
37	*Bond Wash Solution	Open Wash	0 MIN	Ambient
38	*Bond Wash Solution	Wash	0 MIN	Ambient
39	10% NBF	Reagent	30 MIN	Ambient
40	*Bond Wash Solution	Wash	0 MIN	Ambient
41	*Bond Wash Solution	Wash	0 MIN	Ambient
42	*Bond Wash Solution	Wash	0 MIN	Ambient
43	*Bond Wash Solution	Wash	3 MIN	Ambient
44	*Bond Wash Solution	Wash	3 MIN	Ambient
45	*Bond Wash Solution	Wash	0 MIN	Ambient
46	*Bond Wash Solution	Wash	0 MIN	Ambient
47	*Bond Wash Solution	Wash	0 MIN	Ambient
48	PretreatPro	Reagent	0 MIN	Ambient
49	PretreatPro	Reagent	30 MIN	40°C
50	*Bond Wash Solution	Wash	0 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
51	*Bond Wash Solution	Wash	0 MIN	Ambient
52	*Bond Wash Solution	Wash	0 MIN	Ambient
53	*Bond Wash Solution	Wash	1 MIN	Ambient
54	*Bond Wash Solution	Wash	1 MIN	Ambient
55	Hydrogen Peroxide	Reagent	10 MIN	Ambient
56	*Bond Wash Solution	Wash	0 MIN	Ambient
57	*Bond Wash Solution	Wash	0 MIN	Ambient
58	*Bond Wash Solution	Wash	0 MIN	Ambient
59	*Bond Wash Solution	Wash	0 MIN	Ambient
60	*Bond Wash Solution	Wash	0 MIN	Ambient
61	Open 2	Reagent	0 MIN	Ambient
62	Open 2	Reagent	120 MIN	42°C
63	*Bond Wash Solution	Wash	0 MIN	42°C
64	*Bond Wash Solution	Wash	1 MIN	42°C
65	*Bond Wash Solution	Wash	5 MIN	42°C
66	*Bond Wash Solution	Wash	0 MIN	Ambient
67	*Bond Wash Solution	Wash	0 MIN	Ambient
68	*Bond Wash Solution	Wash	0 MIN	Ambient
69	*Bond Wash Solution	Wash	0 MIN	Ambient
70	*Bond Wash Solution	Wash	0 MIN	Ambient
71	*Bond Wash Solution	Wash	1 MIN	Ambient
72	*Bond Wash Solution	Wash	1 MIN	Ambient
73	*Bond Wash Solution	Wash	0 MIN	Ambient
74	RNAscope Multiomic Amp 1	Reagent	1 MIN	42°C
75	RNAscope Multiomic Amp 1	Reagent	30 MIN	42°C
76	*Bond Wash Solution	Wash	0 MIN	Ambient
77	*Bond Wash Solution	Wash	0 MIN	Ambient
78	*Bond Wash Solution	Wash	0 MIN	Ambient
79	*Bond Wash Solution	Wash	3 MIN	Ambient
80	*Bond Wash Solution	Wash	3 MIN	Ambient
81	*Bond Wash Solution	Wash	0 MIN	Ambient
82	*Bond Wash Solution	Wash	0 MIN	Ambient
83	*Bond Wash Solution	Wash	0 MIN	Ambient
84	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
85	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
86	*Bond Wash Solution	Wash	0 MIN	Ambient
87	*Bond Wash Solution	Wash	0 MIN	Ambient
88	*Bond Wash Solution	Open Wash	0 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
89	*Bond Wash Solution	Wash	0 MIN	Ambient
90	RNAscope Multiomic Amp 2	Reagent	1 MIN	42°C
91	RNAscope Multiomic Amp 2	Reagent	30 MIN	42°C
92	*Bond Wash Solution	Wash	0 MIN	Ambient
93	*Bond Wash Solution	Wash	0 MIN	Ambient
94	*Bond Wash Solution	Wash	0 MIN	Ambient
95	*Bond Wash Solution	Wash	3 MIN	Ambient
96	*Bond Wash Solution	Wash	3 MIN	Ambient
97	*Bond Wash Solution	Wash	0 MIN	Ambient
98	*Bond Wash Solution	Wash	0 MIN	Ambient
99	*Bond Wash Solution	Wash	0 MIN	Ambient
100	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
101	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
102	*Bond Wash Solution	Wash	0 MIN	Ambient
103	*Bond Wash Solution	Wash	1 MIN	Ambient
104	*Bond Wash Solution	Open Wash	1 MIN	Ambient
105	*Bond Wash Solution	Wash	1 MIN	Ambient
106	RNAscope Multiomic Amp 3	Reagent	1 MIN	42°C
107	RNAscope Multiomic Amp 3	Reagent	15 MIN	42°C
108	*Bond Wash Solution	Wash	0 MIN	Ambient
109	*Bond Wash Solution	Wash	0 MIN	Ambient
110	*Bond Wash Solution	Wash	0 MIN	Ambient
111	*Bond Wash Solution	Wash	1 MIN	Ambient
112	*Bond Wash Solution	Wash	1 MIN	Ambient
113	*Bond Wash Solution	Wash	1 MIN	Ambient
114	*Bond Wash Solution	Open Wash	1 MIN	Ambient
115	*Bond Wash Solution	Wash	1 MIN	Ambient
116	RNAscope Multiomic HRP-C1	Reagent	1 MIN	42°C
117	RNAscope Multiomic HRP-C1	Reagent	15 MIN	42°C
118	*Bond Wash Solution	Wash	0 MIN	Ambient
119	*Bond Wash Solution	Wash	0 MIN	Ambient
120	*Bond Wash Solution	Wash	0 MIN	Ambient
121	*Bond Wash Solution	Wash	1 MIN	Ambient
122	*Bond Wash Solution	Wash	1 MIN	Ambient
123	*Bond Wash Solution	Wash	1 MIN	Ambient
124	*Bond Wash Solution	Wash	1 MIN	Ambient
125	*Bond Wash Solution	Wash	1 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
126	RNAscope Multiomic TSA-F1	Reagent	1 MIN	Ambient
127	RNAscope Multiomic TSA-F1	Reagent	30 MIN	Ambient
128	*Bond Wash Solution	Wash	0 MIN	Ambient
129	*Bond Wash Solution	Wash	0 MIN	Ambient
130	*Bond Wash Solution	Wash	0 MIN	Ambient
131	*Bond Wash Solution	Wash	1 MIN	Ambient
132	*Bond Wash Solution	Wash	1 MIN	Ambient
133	*Bond Wash Solution	Wash	1 MIN	Ambient
134	*Bond Wash Solution	Wash	1 MIN	Ambient
135	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
136	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
137	*Bond Wash Solution	Wash	0 MIN	Ambient
138	*Bond Wash Solution	Wash	0 MIN	Ambient
139	*Bond Wash Solution	Wash	0 MIN	Ambient
140	*Bond Wash Solution	Wash	1 MIN	Ambient
141	*Bond Wash Solution	Wash	1 MIN	Ambient
142	*Bond Wash Solution	Wash	1 MIN	Ambient
143	*Bond Wash Solution	Wash	1 MIN	Ambient
144	RNAscope Multiomic LS HRP C2	Reagent	1 MIN	42°C
145	RNAscope Multiomic LS HRP C2	Reagent	15 MIN	42°C
146	*Bond Wash Solution	Wash	0 MIN	Ambient
147	*Bond Wash Solution	Wash	0 MIN	Ambient
148	*Bond Wash Solution	Wash	0 MIN	Ambient
149	*Bond Wash Solution	Wash	1 MIN	Ambient
150	*Bond Wash Solution	Wash	1 MIN	Ambient
151	*Bond Wash Solution	Wash	1 MIN	Ambient
152	*Bond Wash Solution	Wash	1 MIN	Ambient
153	RNAscope Multiomic TSA-F2	Reagent	1 MIN	Ambient
154	RNAscope Multiomic TSA-F2	Reagent	30 MIN	Ambient
155	*Bond Wash Solution	Wash	0 MIN	Ambient
156	*Bond Wash Solution	Wash	0 MIN	Ambient
157	*Bond Wash Solution	Wash	0 MIN	Ambient
158	*Bond Wash Solution	Wash	1 MIN	Ambient
159	*Bond Wash Solution	Wash	1 MIN	Ambient
160	*Bond Wash Solution	Wash	1 MIN	Ambient
161	*Bond Wash Solution	Wash	1 MIN	Ambient
162	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
163	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
164	*Bond Wash Solution	Wash	0 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
165	*Bond Wash Solution	Wash	0 MIN	Ambient
166	*Bond Wash Solution	Wash	0 MIN	Ambient
167	*Bond Wash Solution	Wash	1 MIN	Ambient
168	*Bond Wash Solution	Wash	1 MIN	Ambient
169	*Bond Wash Solution	Wash	1 MIN	Ambient
170	*Bond Wash Solution	Wash	1 MIN	Ambient
171	RNAscope Multiomic LS HRP C3	Reagent	1 MIN	42°C
172	RNAscope Multiomic LS HRP C3	Reagent	15 MIN	42°C
173	*Bond Wash Solution	Wash	0 MIN	Ambient
174	*Bond Wash Solution	Wash	0 MIN	Ambient
175	*Bond Wash Solution	Wash	0 MIN	Ambient
176	*Bond Wash Solution	Wash	1 MIN	Ambient
177	*Bond Wash Solution	Wash	1 MIN	Ambient
178	*Bond Wash Solution	Wash	1 MIN	Ambient
179	*Bond Wash Solution	Wash	1 MIN	Ambient
180	RNAscope Multiomic TSA-F3	Reagent	1 MIN	Ambient
181	RNAscope Multiomic TSA-F3	Reagent	30 MIN	Ambient
182	*Bond Wash Solution	Wash	0 MIN	Ambient
183	*Bond Wash Solution	Wash	0 MIN	Ambient
184	*Bond Wash Solution	Wash	0 MIN	Ambient
185	*Bond Wash Solution	Wash	1 MIN	Ambient
186	*Bond Wash Solution	Wash	1 MIN	Ambient
187	*Bond Wash Solution	Wash	1 MIN	Ambient
188	*Bond Wash Solution	Wash	1 MIN	Ambient
189	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
190	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
191	*Bond Wash Solution	Wash	0 MIN	Ambient
192	*Bond Wash Solution	Wash	0 MIN	Ambient
193	*Bond Wash Solution	Wash	0 MIN	Ambient
194	*Bond Wash Solution	Wash	1 MIN	Ambient
195	*Bond Wash Solution	Wash	1 MIN	Ambient
196	*Bond Wash Solution	Wash	1 MIN	Ambient
197	RNAscope Multiomic LS HRP C4	Reagent	1 MIN	42°C
198	RNAscope Multiomic LS HRP C4	Reagent	15 MIN	42°C
199	*Bond Wash Solution	Wash	0 MIN	Ambient
200	*Bond Wash Solution	Wash	0 MIN	Ambient
201	*Bond Wash Solution	Wash	0 MIN	Ambient
202	*Bond Wash Solution	Wash	1 MIN	Ambient
203	*Bond Wash Solution	Wash	1 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
204	*Bond Wash Solution	Wash	1 MIN	Ambient
205	RNAscope Multiomic TSA-F4	Reagent	1 MIN	Ambient
206	RNAscope Multiomic TSA-F4	Reagent	30 MIN	Ambient
207	*Bond Wash Solution	Wash	0 MIN	Ambient
208	*Bond Wash Solution	Wash	0 MIN	Ambient
209	*Bond Wash Solution	Wash	0 MIN	Ambient
210	*Bond Wash Solution	Wash	1 MIN	Ambient
211	*Bond Wash Solution	Wash	1 MIN	Ambient
212	*Bond Wash Solution	Wash	1 MIN	Ambient
213	*Bond Wash Solution	Wash	1 MIN	Ambient
214	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
215	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
216	*Bond Wash Solution	Wash	0 MIN	Ambient
217	*Bond Wash Solution	Wash	0 MIN	Ambient
218	*Bond Wash Solution	Wash	0 MIN	Ambient
219	*Bond Wash Solution	Wash	1 MIN	Ambient
220	*Bond Wash Solution	Wash	1 MIN	Ambient
221	*Bond Wash Solution	Wash	1 MIN	Ambient
222	RNAscope Multiomic LS HRP C5	Reagent	1 MIN	42°C
223	RNAscope Multiomic LS HRP C5	Reagent	15 MIN	42°C
224	*Bond Wash Solution	Wash	0 MIN	Ambient
225	*Bond Wash Solution	Wash	0 MIN	Ambient
226	*Bond Wash Solution	Wash	0 MIN	Ambient
227	*Bond Wash Solution	Wash	1 MIN	Ambient
228	*Bond Wash Solution	Wash	1 MIN	Ambient
229	*Bond Wash Solution	Wash	1 MIN	Ambient
230	RNAscope Multiomic TSA-F5	Reagent	1 MIN	Ambient
231	RNAscope Multiomic TSA-F5	Reagent	30 MIN	Ambient
232	*Bond Wash Solution	Wash	0 MIN	Ambient
233	*Bond Wash Solution	Wash	0 MIN	Ambient
234	*Bond Wash Solution	Wash	0 MIN	Ambient
235	*Bond Wash Solution	Wash	1 MIN	Ambient
236	*Bond Wash Solution	Wash	1 MIN	Ambient
237	*Bond Wash Solution	Wash	1 MIN	Ambient
238	*Bond Wash Solution	Wash	1 MIN	Ambient
239	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
240	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
241	*Bond Wash Solution	Wash	0 MIN	Ambient
242	*Bond Wash Solution	Wash	0 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
243	*Bond Wash Solution	Wash	0 MIN	Ambient
244	*Bond Wash Solution	Wash	1 MIN	Ambient
245	*Bond Wash Solution	Wash	1 MIN	Ambient
246	*Bond Wash Solution	Wash	1 MIN	Ambient
247	RNAscope Multiomic LS HRP C6	Reagent	1 MIN	42°C
248	RNAscope Multiomic LS HRP C6	Reagent	15 MIN	42°C
249	*Bond Wash Solution	Wash	0 MIN	Ambient
250	*Bond Wash Solution	Wash	0 MIN	Ambient
251	*Bond Wash Solution	Wash	0 MIN	Ambient
252	*Bond Wash Solution	Wash	1 MIN	Ambient
253	*Bond Wash Solution	Wash	1 MIN	Ambient
254	*Bond Wash Solution	Wash	1 MIN	Ambient
255	TSA-DIG	Reagent	1 MIN	Ambient
256	TSA-DIG	Reagent	30 MIN	Ambient
257	*Bond Wash Solution	Wash	0 MIN	Ambient
258	*Bond Wash Solution	Wash	0 MIN	Ambient
259	*Bond Wash Solution	Wash	0 MIN	Ambient
260	*Bond Wash Solution	Wash	1 MIN	Ambient
261	*Bond Wash Solution	Wash	1 MIN	Ambient
262	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
263	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
264	*Bond Wash Solution	Wash	0 MIN	Ambient
265	*Bond Wash Solution	Wash	0 MIN	Ambient
266	*Bond Wash Solution	Wash	0 MIN	Ambient
267	*Bond Wash Solution	Wash	1 MIN	Ambient
268	*Bond Wash Solution	Wash	1 MIN	Ambient
269	Polaris 780	Reagent	1 MIN	Ambient
270	Polaris 780	Reagent	30 MIN	Ambient
271	*Bond Wash Solution	Wash	0 MIN	Ambient
272	*Bond Wash Solution	Wash	0 MIN	Ambient
273	*Bond Wash Solution	Wash	0 MIN	Ambient
274	*Bond Wash Solution	Wash	1 MIN	Ambient
275	*Bond Wash Solution	Wash	1 MIN	Ambient
276	DAPI	Reagent	10 MIN	Ambient
277	*Bond Wash Solution	Wash	0 MIN	Ambient
278	*Bond Wash Solution	Wash	0 MIN	Ambient
279	*Bond Wash Solution	Wash	0 MIN	Ambient

E

Appendix E. 2 secondary antibodies + 4 Primary conjugated antibodies

	Reagent	Container name	Details (concentration, dilution)
1	Salmon Sperm DNA	Open1	500 ug/mL, in multiomic antibody diluent
2	10% NBF	10% NBF	None
3	Primary Raw Antibody mix	CoDetection Antibody 1	Multiomic antibody diluent
4	Secondary Conjugated Antibody mix	CoDetection Antibody 2	Multiomic antibody diluent (See Appendix H for concentration)
5	Primary Conjugated Antibody mix	CoDetection Antibody 3	Multiomic antibody diluent (See Appendix H for concentration)
6	Antibody Blocker (mouse and/or Rabbit IgG)	Antibody Blocker	5ug/ml, in multiomic antibody diluent

Slide Setup:

Slide properties

Pos 1

Tissue type: Test tissue Negative tissue Positive tissue Dispense volume: 100 µL 150 µL

Staining mode: Single Routine

Process: IHC ISH

Marker: Mock Probe

Protocols

Staining:	Primary Antibody + Secondary Antibody
Preparation:	*Bake and Dewax
HIER:	*ACD HIER 15 min with ER2 (95)
Enzyme:	*---
Probe Application:	*RNAscope 2.5 LSx Probe Application
Denaturation:	*---
Hybridization:	ACD 1 Min Hybridization
Probe Removal:	*RNAscope 2.5 LSx Probe Removal

Buttons: OK, Copy slide, Close

Step No.	Reagent	Step Type	Incubation Time	Temperature†
1	Open1	Reagent	0 MIN	Ambient
2	Open1	Reagent	60 MIN	Ambient
3	*Bond Wash Solution	Wash	0 MIN	Ambient
4	*Bond Wash Solution	Wash	0 MIN	Ambient
5	*Bond Wash Solution	Wash	0 MIN	Ambient
6	*Bond Wash Solution	Wash	0 MIN	Ambient
7	Co-Detection Antibody 1	Reagent	0 MIN	Ambient
8	Co-Detection Antibody 1	Reagent	60 MIN	Ambient
9	*Bond Wash Solution	Wash	0 MIN	Ambient
10	*Bond Wash Solution	Wash	0 MIN	Ambient
11	*Bond Wash Solution	Wash	1 MIN	Ambient
12	*Bond Wash Solution	Wash	1 MIN	Ambient
13	Co-Detection Antibody 2	Reagent	0 MIN	Ambient
14	Co-Detection Antibody 2	Reagent	30 MIN	Ambient
15	*Bond Wash Solution	Wash	0 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
16	*Bond Wash Solution	Wash	0 MIN	Ambient
17	*Bond Wash Solution	Wash	1 MIN	Ambient
18	*Bond Wash Solution	Wash	1MIN	Ambient
19	Antibody Blocker	Reagent	0 MIN	Ambient
20	Antibody Blocker	Reagent	30 MIN	Ambient
21	*Bond Wash Solution	Wash	0 MIN	Ambient
22	*Bond Wash Solution	Wash	0 MIN	Ambient
23	*Bond Wash Solution	Wash	1 MIN	Ambient
24	*Bond Wash Solution	Wash	3MIN	Ambient
25	*Bond Wash Solution	Wash	3MIN	Ambient
26	Co-Detection Antibody 3	Reagent	0 MIN	Ambient
27	Co-Detection Antibody 3	Reagent	60 MIN	Ambient
28	*Bond Wash Solution	Wash	0 MIN	Ambient
29	*Bond Wash Solution	Wash	0 MIN	Ambient
30	*Bond Wash Solution	Wash	1 MIN	Ambient
31	*Bond Wash Solution	Wash	3MIN	Ambient
32	*Bond Wash Solution	Wash	3MIN	Ambient
33	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
34	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
35	*Bond Wash Solution	Wash	0 MIN	Ambient
36	*Bond Wash Solution	Wash	0 MIN	Ambient
37	*Bond Wash Solution	Open Wash	0 MIN	Ambient
38	*Bond Wash Solution	Wash	0 MIN	Ambient
39	10% NBF	Reagent	30 MIN	Ambient
40	*Bond Wash Solution	Wash	0 MIN	Ambient
41	*Bond Wash Solution	Wash	0 MIN	Ambient
42	*Bond Wash Solution	Wash	0 MIN	Ambient
43	*Bond Wash Solution	Wash	3 MIN	Ambient
44	*Bond Wash Solution	Wash	3 MIN	Ambient
45	*Bond Wash Solution	Wash	0 MIN	Ambient
46	*Bond Wash Solution	Wash	0 MIN	Ambient
47	*Bond Wash Solution	Wash	0 MIN	Ambient
48	PretreatPro	Reagent	0 MIN	Ambient
49	PretreatPro	Reagent	30 MIN	40°C
50	*Bond Wash Solution	Wash	0 MIN	Ambient
51	*Bond Wash Solution	Wash	0 MIN	Ambient
52	*Bond Wash Solution	Wash	0 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
53	*Bond Wash Solution	Wash	1 MIN	Ambient
54	*Bond Wash Solution	Wash	1 MIN	Ambient
55	Hydrogen Peroxide	Reagent	10 MIN	Ambient
56	*Bond Wash Solution	Wash	0 MIN	Ambient
57	*Bond Wash Solution	Wash	0 MIN	Ambient
58	*Bond Wash Solution	Wash	0 MIN	Ambient
59	*Bond Wash Solution	Wash	0 MIN	Ambient
60	*Bond Wash Solution	Wash	0 MIN	Ambient
61	RNAscope Multiomic Amp 1	Reagent	1 MIN	42°C
62	RNAscope Multiomic Amp 1	Reagent	30 MIN	42°C
63	*Bond Wash Solution	Wash	0 MIN	Ambient
64	*Bond Wash Solution	Wash	0 MIN	Ambient
65	*Bond Wash Solution	Wash	0 MIN	Ambient
66	*Bond Wash Solution	Wash	3 MIN	Ambient
67	*Bond Wash Solution	Wash	3 MIN	Ambient
68	*Bond Wash Solution	Wash	0 MIN	Ambient
69	*Bond Wash Solution	Wash	0 MIN	Ambient
70	*Bond Wash Solution	Wash	0 MIN	Ambient
71	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
72	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
73	*Bond Wash Solution	Wash	0 MIN	Ambient
74	*Bond Wash Solution	Wash	0 MIN	Ambient
75	*Bond Wash Solution	Open Wash	0 MIN	Ambient
76	*Bond Wash Solution	Wash	0 MIN	Ambient
77	RNAscope Multiomic Amp 2	Reagent	1 MIN	42°C
78	RNAscope Multiomic Amp 2	Reagent	30 MIN	42°C
79	*Bond Wash Solution	Wash	0 MIN	Ambient
80	*Bond Wash Solution	Wash	0 MIN	Ambient
81	*Bond Wash Solution	Wash	0 MIN	Ambient
82	*Bond Wash Solution	Wash	3 MIN	Ambient
83	*Bond Wash Solution	Wash	3 MIN	Ambient
84	*Bond Wash Solution	Wash	0 MIN	Ambient
85	*Bond Wash Solution	Wash	0 MIN	Ambient
86	*Bond Wash Solution	Wash	0 MIN	Ambient
87	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
88	RNAscope Multiomic LS Rinse	Reagent	5 MIN	Ambient
89	*Bond Wash Solution	Wash	0 MIN	Ambient
90	*Bond Wash Solution	Wash	1 MIN	Ambient
91	*Bond Wash Solution	Open Wash	1 MIN	Ambient
92	*Bond Wash Solution	Wash	1 MIN	Ambient
93	RNAscope Multiomic Amp 3	Reagent	1 MIN	42°C
94	RNAscope Multiomic Amp 3	Reagent	15 MIN	42°C
95	*Bond Wash Solution	Wash	0 MIN	Ambient
96	*Bond Wash Solution	Wash	0 MIN	Ambient
97	*Bond Wash Solution	Wash	0 MIN	Ambient
98	*Bond Wash Solution	Wash	1 MIN	Ambient
99	*Bond Wash Solution	Wash	1 MIN	Ambient
100	*Bond Wash Solution	Wash	1 MIN	Ambient
101	*Bond Wash Solution	Open Wash	1 MIN	Ambient
102	*Bond Wash Solution	Wash	1 MIN	Ambient
103	RNAscope Multiomic HRP-C1	Reagent	1 MIN	42°C
104	RNAscope Multiomic HRP-C1	Reagent	15 MIN	42°C
105	*Bond Wash Solution	Wash	0 MIN	Ambient
106	*Bond Wash Solution	Wash	0 MIN	Ambient
107	*Bond Wash Solution	Wash	0 MIN	Ambient
108	*Bond Wash Solution	Wash	1 MIN	Ambient
109	*Bond Wash Solution	Wash	1 MIN	Ambient
110	*Bond Wash Solution	Wash	1 MIN	Ambient
111	*Bond Wash Solution	Wash	1 MIN	Ambient
112	*Bond Wash Solution	Wash	1 MIN	Ambient
113	RNAscope Multiomic TSA-F1	Reagent	1 MIN	Ambient
114	RNAscope Multiomic TSA-F1	Reagent	30 MIN	Ambient
115	*Bond Wash Solution	Wash	0 MIN	Ambient
116	*Bond Wash Solution	Wash	0 MIN	Ambient
117	*Bond Wash Solution	Wash	0 MIN	Ambient
118	*Bond Wash Solution	Wash	1 MIN	Ambient
119	*Bond Wash Solution	Wash	1 MIN	Ambient
120	*Bond Wash Solution	Wash	1 MIN	Ambient
121	*Bond Wash Solution	Wash	1 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
122	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
123	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
124	*Bond Wash Solution	Wash	0 MIN	Ambient
125	*Bond Wash Solution	Wash	0 MIN	Ambient
126	*Bond Wash Solution	Wash	0 MIN	Ambient
127	*Bond Wash Solution	Wash	1 MIN	Ambient
128	*Bond Wash Solution	Wash	1 MIN	Ambient
129	*Bond Wash Solution	Wash	1 MIN	Ambient
130	*Bond Wash Solution	Wash	1 MIN	Ambient
131	RNAscope Multiomic LS HRP C2	Reagent	1 MIN	42°C
132	RNAscope Multiomic LS HRP C2	Reagent	15 MIN	42°C
133	*Bond Wash Solution	Wash	0 MIN	Ambient
134	*Bond Wash Solution	Wash	0 MIN	Ambient
135	*Bond Wash Solution	Wash	0 MIN	Ambient
136	*Bond Wash Solution	Wash	1 MIN	Ambient
137	*Bond Wash Solution	Wash	1 MIN	Ambient
138	*Bond Wash Solution	Wash	1 MIN	Ambient
139	*Bond Wash Solution	Wash	1 MIN	Ambient
140	RNAscope Multiomic TSA-F2	Reagent	1 MIN	Ambient
141	RNAscope Multiomic TSA-F2	Reagent	30 MIN	Ambient
142	*Bond Wash Solution	Wash	0 MIN	Ambient
143	*Bond Wash Solution	Wash	0 MIN	Ambient
144	*Bond Wash Solution	Wash	0 MIN	Ambient
145	*Bond Wash Solution	Wash	1 MIN	Ambient
146	*Bond Wash Solution	Wash	1 MIN	Ambient
147	*Bond Wash Solution	Wash	1 MIN	Ambient
148	*Bond Wash Solution	Wash	1 MIN	Ambient
149	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
150	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
151	*Bond Wash Solution	Wash	0 MIN	Ambient
152	*Bond Wash Solution	Wash	0 MIN	Ambient
153	*Bond Wash Solution	Wash	0 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
154	*Bond Wash Solution	Wash	1 MIN	Ambient
155	*Bond Wash Solution	Wash	1 MIN	Ambient
156	*Bond Wash Solution	Wash	1 MIN	Ambient
157	*Bond Wash Solution	Wash	1 MIN	Ambient
158	RNAscope Multiomic LS HRP C3	Reagent	1 MIN	42°C
159	RNAscope Multiomic LS HRP C3	Reagent	15 MIN	42°C
160	*Bond Wash Solution	Wash	0 MIN	Ambient
161	*Bond Wash Solution	Wash	0 MIN	Ambient
162	*Bond Wash Solution	Wash	0 MIN	Ambient
163	*Bond Wash Solution	Wash	1 MIN	Ambient
164	*Bond Wash Solution	Wash	1 MIN	Ambient
165	*Bond Wash Solution	Wash	1 MIN	Ambient
166	*Bond Wash Solution	Wash	1 MIN	Ambient
167	RNAscope Multiomic TSA-F3	Reagent	1 MIN	Ambient
168	RNAscope Multiomic TSA-F3	Reagent	30 MIN	Ambient
169	*Bond Wash Solution	Wash	0 MIN	Ambient
170	*Bond Wash Solution	Wash	0 MIN	Ambient
171	*Bond Wash Solution	Wash	0 MIN	Ambient
172	*Bond Wash Solution	Wash	1 MIN	Ambient
173	*Bond Wash Solution	Wash	1 MIN	Ambient
174	*Bond Wash Solution	Wash	1 MIN	Ambient
175	*Bond Wash Solution	Wash	1 MIN	Ambient
176	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
177	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
178	*Bond Wash Solution	Wash	0 MIN	Ambient
179	*Bond Wash Solution	Wash	0 MIN	Ambient
180	*Bond Wash Solution	Wash	0 MIN	Ambient
181	*Bond Wash Solution	Wash	1 MIN	Ambient
182	*Bond Wash Solution	Wash	1 MIN	Ambient
183	*Bond Wash Solution	Wash	1 MIN	Ambient
184	RNAscope Multiomic LS HRP C4	Reagent	1 MIN	42°C
185	RNAscope Multiomic LS HRP C4	Reagent	15 MIN	42°C

Step No.	Reagent	Step Type	Incubation Time	Temperature†
186	*Bond Wash Solution	Wash	0 MIN	Ambient
187	*Bond Wash Solution	Wash	0 MIN	Ambient
188	*Bond Wash Solution	Wash	0 MIN	Ambient
189	*Bond Wash Solution	Wash	1 MIN	Ambient
190	*Bond Wash Solution	Wash	1 MIN	Ambient
191	*Bond Wash Solution	Wash	1 MIN	Ambient
192	RNAscope Multiomic TSA-F4	Reagent	1 MIN	Ambient
193	RNAscope Multiomic TSA-F4	Reagent	30 MIN	Ambient
194	*Bond Wash Solution	Wash	0 MIN	Ambient
195	*Bond Wash Solution	Wash	0 MIN	Ambient
196	*Bond Wash Solution	Wash	0 MIN	Ambient
197	*Bond Wash Solution	Wash	1 MIN	Ambient
198	*Bond Wash Solution	Wash	1 MIN	Ambient
199	*Bond Wash Solution	Wash	1 MIN	Ambient
200	*Bond Wash Solution	Wash	1 MIN	Ambient
201	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
202	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
203	*Bond Wash Solution	Wash	0 MIN	Ambient
204	*Bond Wash Solution	Wash	0 MIN	Ambient
205	*Bond Wash Solution	Wash	0 MIN	Ambient
206	*Bond Wash Solution	Wash	1 MIN	Ambient
207	*Bond Wash Solution	Wash	1 MIN	Ambient
208	*Bond Wash Solution	Wash	1 MIN	Ambient
209	RNAscope Multiomic LS HRP C5	Reagent	1 MIN	42°C
210	RNAscope Multiomic LS HRP C5	Reagent	15 MIN	42°C
211	*Bond Wash Solution	Wash	0 MIN	Ambient
212	*Bond Wash Solution	Wash	0 MIN	Ambient
213	*Bond Wash Solution	Wash	0 MIN	Ambient
214	*Bond Wash Solution	Wash	1 MIN	Ambient
215	*Bond Wash Solution	Wash	1 MIN	Ambient
216	*Bond Wash Solution	Wash	1 MIN	Ambient
217	RNAscope Multiomic TSA-F5	Reagent	1 MIN	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature†
218	RNAscope Multiomic TSA-F5	Reagent	30 MIN	Ambient
219	*Bond Wash Solution	Wash	0 MIN	Ambient
220	*Bond Wash Solution	Wash	0 MIN	Ambient
221	*Bond Wash Solution	Wash	0 MIN	Ambient
222	*Bond Wash Solution	Wash	1 MIN	Ambient
223	*Bond Wash Solution	Wash	1 MIN	Ambient
224	*Bond Wash Solution	Wash	1 MIN	Ambient
225	*Bond Wash Solution	Wash	1 MIN	Ambient
226	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
227	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C
228	*Bond Wash Solution	Wash	0 MIN	Ambient
229	*Bond Wash Solution	Wash	0 MIN	Ambient
230	*Bond Wash Solution	Wash	0 MIN	Ambient
231	*Bond Wash Solution	Wash	1 MIN	Ambient
232	*Bond Wash Solution	Wash	1 MIN	Ambient
233	*Bond Wash Solution	Wash	1 MIN	Ambient
234	RNAscope Multiomic LS HRP C6	Reagent	1 MIN	42°C
235	RNAscope Multiomic LS HRP C6	Reagent	15 MIN	42°C
236	*Bond Wash Solution	Wash	0 MIN	Ambient
237	*Bond Wash Solution	Wash	0 MIN	Ambient
238	*Bond Wash Solution	Wash	0 MIN	Ambient
239	*Bond Wash Solution	Wash	1 MIN	Ambient
240	*Bond Wash Solution	Wash	1 MIN	Ambient
241	*Bond Wash Solution	Wash	1 MIN	Ambient
242	TSA-DIG	Reagent	1 MIN	Ambient
243	TSA-DIG	Reagent	30 MIN	Ambient
244	*Bond Wash Solution	Wash	0 MIN	Ambient
245	*Bond Wash Solution	Wash	0 MIN	Ambient
246	*Bond Wash Solution	Wash	0 MIN	Ambient
247	*Bond Wash Solution	Wash	1 MIN	Ambient
248	*Bond Wash Solution	Wash	1 MIN	Ambient
249	RNAscope Multiomic HRP Blocker	Reagent	1 MIN	42°C
250	RNAscope Multiomic HRP Blocker	Reagent	15 MIN	42°C

Step No.	Reagent	Step Type	Incubation Time	Temperature†
251	*Bond Wash Solution	Wash	0 MIN	Ambient
252	*Bond Wash Solution	Wash	0 MIN	Ambient
253	*Bond Wash Solution	Wash	0 MIN	Ambient
254	*Bond Wash Solution	Wash	1 MIN	Ambient
255	*Bond Wash Solution	Wash	1 MIN	Ambient
256	Polaris 780	Reagent	1 MIN	Ambient
257	Polaris 780	Reagent	30 MIN	Ambient
258	*Bond Wash Solution	Wash	0 MIN	Ambient
259	*Bond Wash Solution	Wash	0 MIN	Ambient
260	*Bond Wash Solution	Wash	0 MIN	Ambient
261	*Bond Wash Solution	Wash	1 MIN	Ambient
262	*Bond Wash Solution	Wash	1 MIN	Ambient
263	DAPI	Reagent	10 MIN	Ambient
264	*Bond Wash Solution	Wash	0 MIN	Ambient
265	*Bond Wash Solution	Wash	0 MIN	Ambient
266	*Bond Wash Solution	Wash	0 MIN	Ambient

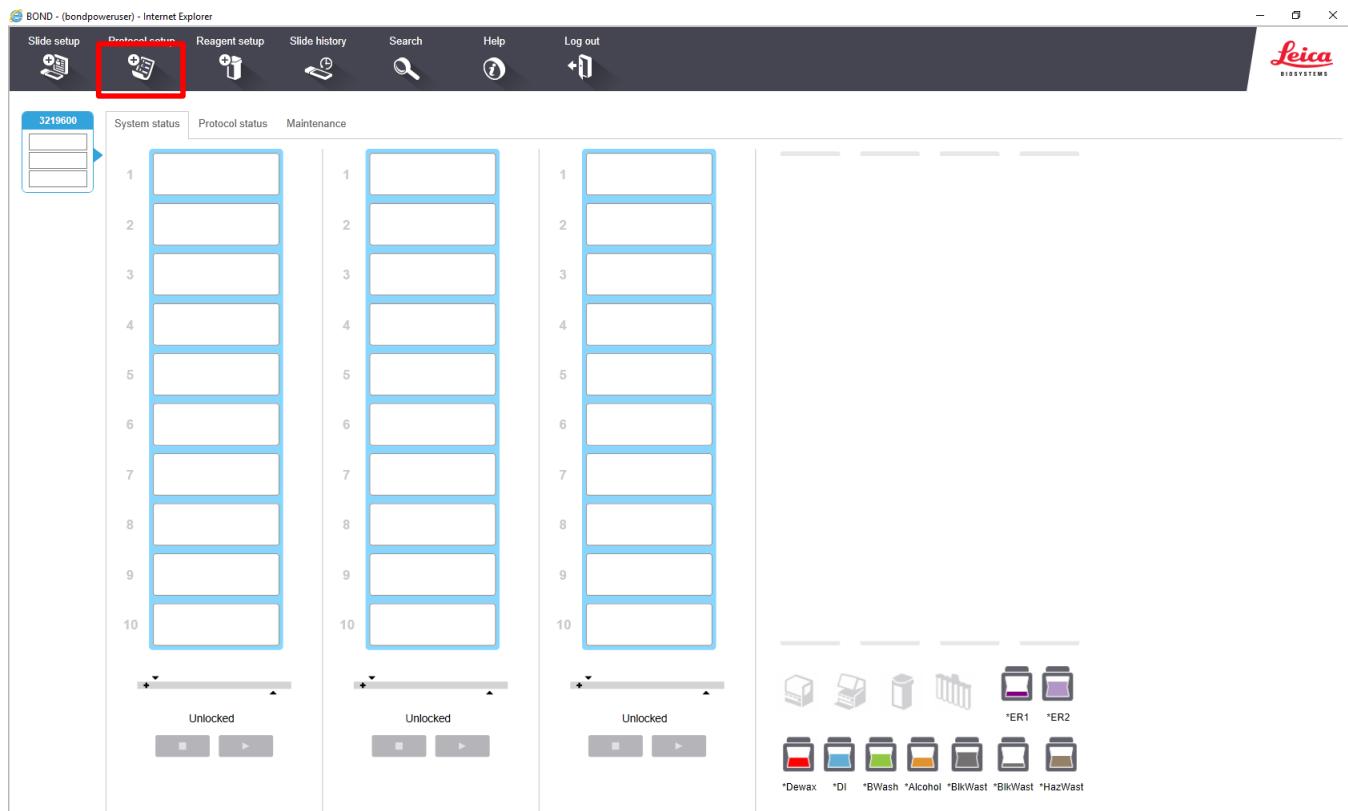
F

Appendix F. Edit the Epitope Retrieval Protocol

The following example shows how to edit the Epitope Retrieval procedure from within the software.

Create a prestaining protocol

1. Open the Leica BOND software and click on the **Protocol setup icon** as shown.



2. Select **Prestaining** under the Protocol group menu and **Heat pretreatment** under the Protocol type menu to access the heat pretreatment protocols.

Protocol name	Protocol type	Description	Modified by	Mod. date	Prof.
*AccuCyte CTC HIER 8 mins	Heat pretreatment	AccuCyte CTC HIER 8 mins	Leica	9/1/2020	✓
*ACD HIER 15 min with ER2 (95)	Heat pretreatment	ACD RNAscope heat pretreatment	Leica	9/1/2020	✓
*E10 and H2 20 HIER	Heat pretreatment	10 min enzyme & 20 min heat retrieval with ER2 variant 1	Leica	9/1/2020	
*E10 and H2 20 HIER2	Heat pretreatment	10 min enzyme & 20 min heat retrieval with ER2 variant 2	Leica	9/1/2020	
*HIER 10 min with ER1	Heat pretreatment	10 min Heat Retrieval using ER1	Leica	9/1/2020	✓
*HIER 10 min with ER2	Heat pretreatment	10 min Heat Retrieval using ER2	Leica	9/1/2020	✓
*HIER 20 min with ER1	Heat pretreatment	20 min Heat Retrieval using ER1	Leica	9/1/2020	✓
*HIER 20 min with ER1 (95)	Heat pretreatment	20 min Heat Retrieval using ER1 at 95	Leica	9/1/2020	
*HIER 20 min with ER2	Heat pretreatment	20 min Heat Retrieval using ER2	Leica	9/1/2020	✓
*HIER 25 min with ER1 (97)	Heat pretreatment	25 min Heat Retrieval using ER1	Leica	9/1/2020	✓
*HIER 30 min with ER1	Heat pretreatment	30 min Heat Retrieval using ER1	Leica	9/1/2020	✓
*HIER 30 min with ER2	Heat pretreatment	30 min Heat Retrieval using ER2	Leica	9/1/2020	✓
*HIER 40 min with ER1	Heat pretreatment	40 min Heat Retrieval using ER1	Leica	9/1/2020	✓
*HIER 40 min with ER1 (98)	Heat pretreatment	40 min Heat Retrieval using ER1 at 98	Leica	9/1/2020	
*HIER 40 min with ER2	Heat pretreatment	40 min Heat Retrieval using ER2	Leica	9/1/2020	✓
*HIER 5 min with ER1	Heat pretreatment	5 min Heat Retrieval using ER1	Leica	9/1/2020	✓
*RNAscope 2.5 LSx Target Retrieval (88)	Heat pretreatment	RNAscope 2.5 LSx heat retrieval 88C	Leica	9/1/2020	✓
*RNAscope 2.5 LSx Target Retrieval (95)	Heat pretreatment	RNAscope 2.5 LSx heat pretreatment 95C	Leica	9/1/2020	✓
*ViewRNA HIER 10 min, ER1 (95)	Heat pretreatment	10 min heat pretreatment, ER1 @ 95C for Affymetrix ViewRNA eZ Assay	Leica	9/1/2020	✓
*ViewRNA HIER 10 min, ER2 (90)	Heat pretreatment	10 min heat pretreatment, ER2 @ 90C for Affymetrix ViewRNA eZ Assay	Leica	9/1/2020	✓
ACD HIER 10 min with ER2 (88)	Heat pretreatment	ACD RNAscope heat pretreatment 10 min at 88	bondpoweruser	10/18/2021	✓

3. Highlight the ***ACD HIER 15 min with ER2 (95)** protocol. Select **Copy**.

Note: ER2 = Epitope Retrieval 2.

- Rename the protocol as **ACD HIER 15 min with ER2 (88)**.
- Rename the Abbreviated name as **ER2-88**.
- Rename the Description to **ACD RNAscope heat pretreatment 88**.
- Highlight the third ***BOND ER Solution 2** step (see above) and change temperature to **88°C**.

New protocol properties

Name:	ACD HIER 15 min with ER2 (88)
Abbreviated name:	ACD88
Description:	ACD RNAscope heat pretreatment 88

Preferred

BOND RX	Import protocol	Protocol type: Heat pretreatment						
Step N°	Wash	Reagent	Supplier	Ambient	Temperature	Inc. (min)	Dispense type	
1		*Bond ER Solution 2	Leica Microsystems	✓	0:00	150 µL		
2		*Bond ER Solution 2	Leica Microsystems	✓	0:00	150 µL		
3		*Bond ER Solution 2	Leica Microsystems	✓	88	15:00	Intermediate	
4		*Bond ER Solution 2	Leica Microsystems	✓	0:00	150 µL		

Show wash steps

Save Cancel

8. For RNAscope, ER 2 temperature varies between 95°C and 88°C depending on the tissue type used. Please see Appendix A for a list of tissues.
9. Select **Save** to create a protocol for ER2 pretreatment at 88°C.
10. If needed, repeat Steps 1–8 to create new heating protocols for different incubation times (for example, ACD 25minER2).



Appendix G. Edit the Protease Protocol

The following example shows how to edit the Protease procedure from within the software.

1. Select **Enzyme Pretreatment** under the Protocol type menu (bottom left).
2. Highlight the *ACD 15min Protease protocol. Select **Copy**.

Protocol name	Protocol type	Description	Modified by	Mod. date	Pref.
*ACD 15 min Protease	Enzyme pretreatment	ACD RNAscope enzyme pretreatment	Leica	9/1/2020	<input checked="" type="checkbox"/>
*BaseScope LSx Enzyme	Enzyme pretreatment	15 min Enzyme Pretreatment using BaseScope LSx Protease	Leica	5/5/2021	
*E10 and H2 20	Enzyme pretreatment	10 min enzyme & 20 min heat retrieval with ER2	Leica	9/1/2020	
*Enzyme 1 (20)	Enzyme pretreatment	Enzyme 1 pretreatment (20 min) for ViewRNA eZ-L Assay	Leica	9/1/2020	
*Enzyme 1 for 10 min	Enzyme pretreatment	10 min Enzyme Pretreatment using Enzyme 1	Leica	9/1/2020	<input checked="" type="checkbox"/>
*Enzyme 1 for 15 min	Enzyme pretreatment	15 min Enzyme Pretreatment using Enzyme 1	Leica	9/1/2020	<input checked="" type="checkbox"/>
*Enzyme 1 for 5 min	Enzyme pretreatment	5 min Enzyme Pretreatment using Enzyme 1	Leica	9/1/2020	<input checked="" type="checkbox"/>
*Enzyme 2 (20)	Enzyme pretreatment	Enzyme 2 pretreatment (20 min) for ViewRNA eZ-L Assay	Leica	9/1/2020	
*Enzyme 2 for 10 min	Enzyme pretreatment	10 min Enzyme Pretreatment using Enzyme 2	Leica	9/1/2020	<input checked="" type="checkbox"/>
*Enzyme 2 for 15 min	Enzyme pretreatment	15 min Enzyme Pretreatment using Enzyme 2	Leica	9/1/2020	<input checked="" type="checkbox"/>
*Enzyme 3 (20)	Enzyme pretreatment	Enzyme 3 pretreatment (20 min) for ViewRNA eZ-L Assay	Leica	9/1/2020	
*Enzyme 3 for 10 min	Enzyme pretreatment	10 min Enzyme Pretreatment using Enzyme 3	Leica	9/1/2020	<input checked="" type="checkbox"/>
*Enzyme 3 for 15 min	Enzyme pretreatment	15 min Enzyme Pretreatment using Enzyme 3	Leica	9/1/2020	<input checked="" type="checkbox"/>
*Enzyme 5 for 25 min	Enzyme pretreatment	25 min Enzyme Pretreatment using Enzyme 5	Leica	9/1/2020	<input checked="" type="checkbox"/>
*Protease 20 min and fix	Enzyme pretreatment	Protease 20 min and fix	Leica	9/1/2020	<input checked="" type="checkbox"/>
*RNAscope 2.5 LSx Enzyme	Enzyme pretreatment	15 min Enzyme Pretreatment using RNAscope 2.5 LSx Protease	Leica	9/1/2020	<input checked="" type="checkbox"/>
*ViewRNA Enzyme 1 (20)	Enzyme pretreatment	20 min enzyme pretreatment for Affymetrix ViewRNA eZ Assay	Leica	9/1/2020	<input checked="" type="checkbox"/>
*ViewRNA Enzyme 2 (20)	Enzyme pretreatment	20 min enzyme pretreatment for Affymetrix ViewRNA eZ Assay	Leica	9/1/2020	<input checked="" type="checkbox"/>
*ViewRNA Enzyme 3 (20)	Enzyme pretreatment	20 min enzyme pretreatment for Affymetrix ViewRNA eZ Assay	Leica	9/1/2020	<input checked="" type="checkbox"/>
2hr 54C preheat	Enzyme pretreatment	ACD RNAscope enzyme pretreatment 2hr 54C	bondpoveruser	2/21/2023	<input checked="" type="checkbox"/>
30min 40C preheat	Enzyme pretreatment	ACD RNAscope enzyme pretreatment 30min 40c	bondpoveruser	7/31/2023	<input checked="" type="checkbox"/>

Protocol group: Protocol type: Staining status: Protocol origin: Preferred status:

3. Rename the protocol to **ACD 25min Protease**.
4. Rename the Abbreviated name to **25minPro**.
5. Rename the Description to **ACD RNAscope 25min enzyme**.
2. Highlight the second *ACD Enzyme step. Keep the temperature at 40°C and set the enzyme incubation time to the desired time (for example, 25min).

New protocol properties

Name:	ACD 25 min Protease
Abbreviated name:	25mPro
Description:	ACD RNAscope 25min enzyme pretreatment

Preferred

BOND RX Import protocol Protocol type: Enzyme pretreatment

Step N°	Wash	Reagent	Supplier	Ambient	Temperature	Inc. (min)	Dispense type
2		*ACD Enzyme	Advanced Cell Diagnostics		40	0:00	150 µL
3		*ACD Enzyme	Advanced Cell Diagnostics		40	25:00	50 µL
7		*Open 0 Haz	User	✓		10:00	150 µL

Show wash steps

Save Cancel

3. Select **Save**.
4. If needed, repeat Steps 1–7 to create a new protease protocol for different sample types (for example, ACD 10min Protease or ACD 15min Protease at ambient temperature).

H

Appendix H. Conjugated antibody concentration

The following guide shows how to dilute the conjugated antibodies with the Multiomic antibody diluent provided.

1. Determine the number of slides needed for the run.
2. Calculate total volume for primary and secondary conjugated antibodies. Make sure to add enough dead-volume to your calculation depending on the container type used. Use 6 mL BOND Titration containers for up to 10 slides.
3. 600 µL dead-volume when using a BOND Titration container (6mL)
4. Use the following table for suggested concentration of each conjugated antibody.
5. Dilute secondary conjugated antibody in a separate tube.
6. Pool all four primary conjugated antibodies together in same tube.
7. Add the diluted antibodies to the appropriate containers.
8. Assign fluorophores using the following recommendations for best results.

RNAscope antibody	Cat No.	Channel	Dilution factor	Opal Dye	Dye Dilution
RNAscope™ Ab Hs CD4-C3	322949	C3	75x	480	1:3000
RNAscope™ Ab Hs CD8-C4	322951	C4	75x	690	1:5000
RNAscope™ Ab Hs PanCK-C5	322952	C5	75x	780	1:500 (TSA-DIG) + 1:125 (Opal 780 Reagent†)
RNAscope™ Ab Hs FoxP3-C6	322953	C6	75x	520	1:5000
RNAscope™ Ab NeuN-C3	AB0018-C3	C3	75x	480	1:3000
RNAscope™ Ab GFAP-C4	AB0028-C4	C4	75x	520	1:3000
RNAscope™ Ab IBA-1-C5	AB0038-C5	C5	75x	690	1:5000
RNAscope™ anti-rabbit-C1	322954	C1	25x	620	1:10000
RNAscope™ anti-mouse-C2	322956	C2	25x	570	1:10000

Appendix I. Pretreatment Guidance for FFPE Samples – for RNA targets only

Follow the recommended pretreatment conditions based on your tissue type for:

- Any new or previously untested FFPE tissue types
- Samples prepared differently than the sample preparation protocol found in [Chapter 3](#).
- For specific guidance on other sample preparations contact ACD Support at support.acd@bio-techne.com

Tissue-specific pretreatment conditions

Refer to the following table for tissue specific FFPE pretreatment conditions. For information about species or tissue type not listed here, contact support at support.acd@bio-techne.com.

Species	Tissue Type	Pathology	Pretreatment Condition	Species	Tissue Type	Pathology	Pretreatment Condition
Mouse/Rat	Intestine	Normal	Standard	Human	Head	Cancer	Standard
	Intestine	Tumor	Standard		Neck	Cancer	Standard
	Embryo	Normal	Standard		Liver	Cancer	Standard
	Brain	Normal	Standard		Liver	Normal	Standard
	Spleen	Normal	Standard		Heart	Normal	Standard
	Eye/Retina	Normal	Extended		GI tract	Normal	Standard
	Liver	Normal	Standard		Kidney	Normal	Standard
	Kidney	Normal	Standard		Skin	Normal	Standard
Human	Breast	Tumor	Standard		Lymphoma	Cancer	Standard
	Colon	Tumor	Standard		Thymus	Normal	Mild/Standard
	Colon	Normal	Standard		Melanoma	Tumor	Standard
	Lung	Tumor	Standard		Nevus	Benign	Standard
	Lung	Normal	Standard		Placenta	Normal	Standard
	Prostate	Tumor	Standard		Skin (TMA*)	Normal	Standard
	Prostate	Normal	Standard		Breast (TMA*)	Normal	Standard
	Lymph node	Tumor	Standard		Melanoma (TMA*)	Normal	Standard
	Lymph node	Normal	Mild		Nevus (TMA)	Benign	Standard
	Tonsil	Normal	Mild/Standard		Stomach (TMA)	Normal	Standard
	Pancreas	Normal	Standard		Stomach (TMA)	Tumor	Standard
	Cervical	Cancer	Standard		Cell pellets, fixed with 10% NBF	—	Mild
	Cervical	Normal	Standard		HeLa or 3T3 cells, fixed with 10% Formaldehyde /PBS/ACD Control	—	Mild
	Cervical dysplasia	Abnormal	Standard		Xenograft tissue	—	Mild
	Brain	Tumor	Standard				
	Brain	Normal	Standard				

*Tissue Microarray

Species	Tissue Type	Pathology	Pretreatment Condition	Species	Tissue Type	Pathology	Pretreatment Condition
Cyno monkey	Spleen	Normal	Mild	Dog	Spleen	Normal	Mild
	Lymph Node	Normal	Mild		Lymph Node	Mild	Mild
	Tonsil	Normal	Mild		Tonsil	N.A.	N.A.
	Thymus	Normal	Mild		Thymus	Mild	Mild
	Retina	Normal	Mild		Retina	Mild	Mild
	Prostate Gland	Normal	Standard/Mild		Prostate Gland	Mild	Mild
	Epididymis	Normal	Mild/Standard		Epididymis	Mild	Mild
	Testis	Normal	Mild/Standard		Testis	Mild/Standard	Mild/Standard
	Ovary	Normal	Mild/Standard		Ovary	Mild/Standard	Mild/Standard
	Duodenum	Normal	Mild/Standard		Duodenum	Normal	Mild
	Jejunum	Normal	Mild/Standard		Jejunum	Normal	Mild
	Colon	Normal	Standard		Colon	Normal	Mild
	Adrenal Gland	Normal	Mild/Standard		Adrenal Gland	Normal	Standard/Mild
Rat	Spleen	Normal	Mild				
	Lymph Node	Normal	Mild				
	Tonsil	Normal	N.A.				
	Thymus	Normal	Mild				
	Retina	Normal	Mild				
	Prostate Gland	Normal	Standard/Mild				
	Epididymis	Normal	Standard				
	Testis	Normal	Standard				
	Ovary	Normal	Standard				
	Duodenum	Normal	Standard/Mild				
	Jejunum	Normal	Standard				
	Colon	Normal	Standard				
	Adrenal Gland	Normal	N.A.				

Appendix J. How to use LS PretreatPro – for RNA targets only

Prepare the instrument reagents

This workflow should be followed if you are performing a staining run with only RNA targets and want to utilize a protease-free permeabilization approach (see page 21 for decision tree). This workflow uses LS PretreatPro in place of traditional reagents.

Traditional protease workflow	New protease free workflow
LS Protease III	LS PretreatPro

1. Place RNAscope 2.5 LS PretreatPro into a BOND Open container and register it as ***ACD Enzyme**.

Note: Visually identify the container as LS PretreatPro to avoid unintended use.

2. Place the ACD Amp Pro into a BOND Open container and register it as reagent created.

Note: Place only one *ACD Enzyme reagent on the instrument at a time.

3. When registering containers select the corresponding name of the reagent from the drop-down menu as showing in the following table under **Container Name**:

Reagents	Container Name
RNAscope 2.5 LS Hydrogen Peroxide	*Open 0 Haz
RNAscope 2.5 LS PretreatPro	*ACD Enzyme
RNAscope Multiomic LS AMP 1	Amp 1
RNAscope Multiomic LS AMP 2	Amp 2
RNAscope Multiomic LS AMP 3	Amp 3
RNAscope Multiomic LS HRP C1	MOHRPC1
RNAscope Multiomic LS HRP C2	MOHRPC2
RNAscope Multiomic LS HRP C3	MOHRPC3
RNAscope Multiomic LS HRP C4	MOHRPC4
ACD Multiomic TSA-F1	MOTSAF1
ACD Multiomic TSA-F2	MOTSAF2
ACD Multiomic TSA-F3	MOTSAF3
ACD Multiomic TSA-F4	MOTSAF4
RNAscope 2.5 LS Rinse	RNAscope Multiomic LS Rinse
RNAscope 2.5 LS Target Probe	Variable (probe 1 2.5)

* Indicates reagent is hard coded in software by Leica Biosystems.

Note: When paired with correlated Enzyme treatment protocol, container names *Open 0 Haz and *RNAscope 2.5 LSx Hydrogen Peroxide are equivalent and *ACD Enzyme and *RNAscope 2.5 LSx Enzyme are equivalent.

5. Enter the RNAscope 2.5 LS Reagent Kit lot number and the expiration date in their respective fields. Select **OK**.

IMPORTANT! Do not introduce bubbles into the solutions by shaking the containers. To mix reagents, gently invert the containers several times. If bubbles are present, leave the containers out at room temperature until the bubbles dissipate.

Update the protocol

1. Filter **Protocol Group** by **Prestaining**, and filter for **Protocol type** by **Enzyme Pretreatment**. Find existing protocol ***ACD 15min Protease** that includes the two reagents ***ACD Enzyme** and ***Open 0 Haz**.
2. Copy and create a new protocol:
 - a. Change the ***ACD Enzyme** incubation time to **15 MIN** per step for two steps and the temperature to **40°C**.
 - b. Keep ***Open 0 Haz** incubation time at **10 MIN** and temperature at **Ambient**. Save the protocol with a new name such as **ACD 30min PretreatPro**.
3. Refer to **Appendix C** for detailed instructions on editing an Enzyme treatment protocol.
4. Run all the other parts of the assay as usual and change only this reagent.
5. The final Slide setup for an FFPE sample should look like the following considering proper HIER condition choice.
 - a. Staining: Choose an edited LS multiplex/multiomics staining protocol as created in Chapter 6.
 - b. Preparation: Select ***Bake and Dewax**.
 - c. HIER: Choose ***ACD HIER 15 min with ER2 (95)**.
 - d. Enzyme: Choose the protocol created in step 2.
 - e. Probe Application: Select ***RNAscope 2.5 LSx Probe Application**.
 - f. Denaturation: Select ***....**.
 - g. Hybridization: Select ***RNAscope 2.5 LSx Hybridization**.
 - h. Probe Removal: Select ***RNAscope 2.5 LSx Probe Removal**.
 - i. Select **Add slide**.
6. After adding all the slides to the study, select **Close** to return to the Slide setup screen.
7. Proceed to **Chapter 7. Run the RNAscope 2.5 LS Assay**.

Note: For all protease-free workflows using LS PretreatPro and involving a low RNA expressor and/or a dense tissue like liver or spleen, we recommend boosting the ISH signal by increasing the target retrieval (ER2) strength [duration, temperature, or volume] or increasing fluorophore concentration to achieve preferred results.

K

Appendix K. Slide Setup for Additional Tissue Types – for RNA targets only

Alternatively prepared samples can be stained on the BOND RX using the following slide setup parameters.

Note: Choose appropriate staining and hybridization related protocols depending on whether you are using the mock probe workflow (Chapter 5) or the standard probe workflow (Chapter 6).

Fixed-frozen tissues

As described in **Chapter 4**, these tissues need a gentle target retrieval step.

1. In Slide setup, select the following:
 - a. Staining: Choose the appropriate protocol for the chemistry and workflow you are using.
 - b. Preparation: Select *----.
 - c. HIER: Choose ***ACD HIER 5 min with ER2 (95)**. See **Appendix B** to create this protocol.
 - d. Enzyme: Select the appropriate protocol for the chemistry and workflow you are using; ***ACD 15min Protease** or **RNAscope LSx Enzyme**.
 - e. Probe Application: Select ***RNAscope 2.5 LSx Probe Application**.
 - f. Denaturation: Choose *....
 - g. Hybridization: Choose the appropriate protocol for the chemistry and workflow you are using; **ACD 1 min Hybridization** or ***RNAscope 2.5 LSx Hybridization**.
 - h. Probe Removal: Select ***RNAscope 2.5 LSx Probe Removal**.
2. Protease incubation time may need to be adjusted but start with **15 MINS** as that works for most tissues.

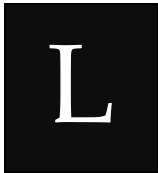
Fresh-frozen tissues

As described in **Chapter 4**, these tissues do NOT need a target retrieval. Instead, permeabilize the tissue at ambient temperature with a stronger protease such as RNAscope LS Protease IV (Cat. No. 322140).

1. In Slide setup, please skip the following steps: 1) Bake or Bake and Dewax 2) Heat retrieval. Choose the following instead:
 - a. Staining: Select the appropriate protocol for the chemistry and workflow you are using.
 - b. Preparation: Choose *----.
 - c. HIER: Choose *----.
 - d. Enzyme: Select **ACD 30min RT with LS Protease IV[†]**.
 - e. Probe Application: Select ***RNAscope 2.5 LSx Probe Application**.
 - f. Denaturation: Select *....

- g. Hybridization: Choose the appropriate protocol for the chemistry and workflow you are using: **ACD 1 min Hybridization** or ***RNAscope 2.5 LSx Hybridization**.
- h. Probe Removal: Select ***RNAscope 2.5 LSx Probe Removal**.
 - [†]See **Appendix G** to edit the protease protocol.

Note: Start your run immediately after setting it up. Do not use a delayed start. This causes poor protease spreadability and negatively impacts results. When the run is complete, the BOND RX rinses the slides every 10 minutes which can impact the counterstain. Set up the instrument as late in the day as possible. Rinsing does not affect the RNAscope signal and counterstaining can be repeated offline in the morning if needed.



Appendix L. Safety

Chemical safety

 **WARNING!** **GENERAL CHEMICAL HANDLING.** To minimize hazards, ensure laboratory personnel read and practice the general safety guidelines for chemical usage, storage, and waste provided below, and consult the relevant SDS for specific precautions and instructions:

- Read and understand the Safety Data Sheets (SDSs) provided before you store, handle, or work with any chemicals or hazardous materials. To obtain SDSs, visit <http://www.acdbio.com/technical-support/user-manuals>.
- Minimize contact with chemicals. Wear appropriate personal protective equipment when handling chemicals (for example, safety glasses, gloves, or protective clothing).
- Minimize the inhalation of chemicals. Do not leave chemical containers open. Use only with adequate ventilation (for example, fume hood).
- Characterize (by analysis if necessary) the waste generated by the particular applications, reagents, and substrates used in your laboratory.
- Ensure that the waste is stored, transferred, transported, and disposed of according to all local, state/provincial, and/or national regulations.
- **IMPORTANT!** Radioactive or biohazardous materials may require special handling, and disposal limitations may apply.

Biological hazard safety

 **WARNING!** **BIOHAZARD.** Biological samples such as tissues, body fluids, infectious agents, and blood of humans and other animals have the potential to transmit infectious diseases. Follow all applicable local, state/provincial, and/or national regulations. Wear appropriate protective equipment, which includes but is not limited to: protective eyewear, face shield, clothing/lab coat, and gloves. All work should be conducted in properly equipped facilities using the appropriate safety equipment (for example, physical containment devices). Individuals should be trained according to applicable regulatory and company/institution requirements before working with potentially infectious materials. Read and follow the applicable guidelines and/or regulatory requirements in the following:

In the U.S.:

- U.S. Department of Health and Human Services guidelines published in Biosafety in Microbiological and Biomedical Laboratories found at www.cdc.gov/biosafety
- Occupational Safety and Health Standards, Bloodborne Pathogens (29 CFR§1910.1030)
- Your company's/institution's Biosafety Program protocols for working with/handling potentially infectious materials
- Additional information about biohazard guidelines is available at www.cdc.gov/

In the EU:

- Check local guidelines and legislation on biohazard and biosafety precaution and refer to the best practices published in the World Health Organization (WHO) Laboratory Biosafety Manual, third edition
- Registration, Evaluation, Authorisation and Restriction of Chemicals (REACH)

Documentation and Support

Obtaining SDSs

Safety Data Sheets (SDSs) are available at: <https://acdbio.com/technical-support/user-manuals>. For the SDSs of chemicals not distributed by Advanced Cell Diagnostics, contact the chemical manufacturer.

Obtaining support

For the latest services and support information, go to: <https://acdbio.com/technical-support/support-overview>.

At the website, you can:

- Access telephone and fax numbers to contact Technical Support and Sales facilities.
- Search through frequently asked questions (FAQs).
- Submit a question directly to Technical Support.
- Search for user documents, SDSs, application notes, citations, training videos, and other product support documents.
- Find out information about customer training events.

Contact information

Advanced Cell Diagnostics, Inc.
7707 Gateway Blvd Suite 200
Newark, CA 94560
Toll Free: 1-877-576-3636
Direct: 1-510-576-8800
Fax: 1-510-576-8801
Information: info.acd@bio-techne.com
Orders: orders.acd@bio-techne.com
Support Email: support.acd@bio-techne.com

Limited product warranty

Advanced Cell Diagnostics, Inc. and/or its affiliate(s) warrant their products as set forth in the ACD General Terms and Conditions of Sale found on the ACD website. If you have any questions, please contact Advanced Cell Diagnostics at: <https://acdbio.com/about/contact>.

Headquarters

7707 Gateway Blvd Suite 200, Newark, CA 94560
Phone 1-510-576-8800 Toll Free 1-877-576-3636

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